

## MiR-21-5p and miR-126a-3p levels in plasma and circulating angiogenic cells: relationship with type 2 diabetes complications

Fabiola Olivieri<sup>1,2</sup>, Liana Spazzafumo<sup>3</sup>, Massimiliano Bonafè<sup>4</sup>, Rina Recchioni<sup>2</sup>, Francesco Prattichizzo<sup>1</sup>, Fiorella Marcheselli<sup>2</sup>, Luigina Micolucci<sup>1</sup>, Emanuela Mensà<sup>2</sup>, Angelica Giuliani<sup>1</sup>, Gabriele Santini<sup>1</sup>, Mirko Gobbi<sup>1</sup>, Raffaella Lazzarini<sup>2</sup>, Massimo Boemi<sup>5</sup>, Roberto Testa<sup>6</sup>, Roberto Antonicelli<sup>7</sup>, Antonio Domenico Procopio<sup>1,2</sup> and Anna Rita Bonfigli<sup>8</sup>

<sup>1</sup> Department of Clinical and Molecular Sciences, DISCLIMO, Università Politecnica delle Marche, Ancona, Italy

<sup>2</sup> Center of Clinical Pathology and Innovative Therapy, INRCA-IRCCS National Institute, Ancona, Italy

<sup>3</sup> Center of Biostatistics, INRCA-IRCCS National Institute, Ancona, Italy

<sup>4</sup> Department of Experimental, Diagnostic and Specialty Medicine, DIMES, University of Bologna, Bologna, Italy

<sup>5</sup> Metabolic Diseases and Diabetology Unit, INRCA-IRCCS National Institute, Ancona, Italy

<sup>6</sup> Experimental Models in Clinical Pathology, INRCA-IRCCS National Institute, Ancona, Italy

<sup>7</sup> UTIC-Cardiology Unit, INRCA-IRCCS National Institute, Ancona, Italy

<sup>8</sup> Scientific Direction, INRCA-IRCCS, National Institute, Ancona, Italy

**Correspondence to:** Fabiola Olivieri, *email:* f.olivieri@univpm.it

**Keywords:** miR-21, miR-126, type 2 diabetes, diabetes complication, circulating miRNAs, Gerotarget

**Received:** September 10, 2015

**Accepted:** October 02, 2015

**Published:** October 19, 2015

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

### ABSTRACT

**Innovative biomarkers are required to manage type 2 diabetic patients (T2DM). We focused our study on miR-126-3p and miR-21-5p levels, as biomarkers of endothelial function and inflammation. MiRNAs levels were measured in plasma from 107 healthy subjects (CTR) and 193 diabetic patients (T2DM), 76 without (T2DM NC) and 117 with (T2DM C) complications.**

**When diabetic complication were analysed as a whole, miR-126-3p and miR-21-5p levels declined significantly from CTR to T2DM NC and T2DM C patients. When miRNAs levels were related to specific complications, significantly higher miR-21-5p levels ( $0.46 \pm 0.44$  vs.  $0.26 \pm 0.33$ ,  $p < 0.001$ ) and significant lower miR-126-3p levels ( $0.21 \pm 0.21$  vs.  $0.28 \pm 0.22$ ,  $p = 0.032$ ) were found in T2DM with previous major cardiovascular events (MACE) vs. all the others T2DM patients.**

**To confirm these results we focused on circulating angiogenic cells (CACs) from a subgroup of 10 CTR, 15 T2DM NC and 15 T2DM patients with MACE. CACs from T2DM patients expressed higher miR-21-5p and lower miR-126-3p levels than CACs from CTR. Furthermore, CACs from T2DM + MACE showed the highest levels of miR-21-5p.**

**Circulating miR-21-5p and miR-126-3p emerge as dynamic biomarkers of systemic inflammatory/angiogenic status. Their expression levels in CACs from T2DM with MACE suggest a shift from a proangiogenic to a proinflammatory profile.**

### INTRODUCTION

Type 2 diabetes mellitus (T2DM) is an increasingly serious public health problem, since the number of affected adults and the proportion of T2DM patients with diabetic complications is expected to grow substantially in

future decades [1]. The common and severe complications of T2DM are silent killers and are often irreversible at the time of diagnosis [2]. Genetic variability does not account for the current T2DM epidemic nor for its complications. Therefore, innovative strategies for stratifying patients with T2DM complications using blood-

based biomarkers are being sought all over the world [3, 4]. The underlying pathogenesis of T2DM complications is not easily elucidated, because diabetic complications are characterized by heterogeneous phenotypes. A recent meta-analysis of randomised controlled trials has demonstrated that the benefit-risk ratio of intensive glucose lowering treatment, administered to prevent diabetic vascular complications, remains uncertain [5]. This suggests that the parameters currently used to monitor T2DM progression do not adequately predict the probability of developing complications. It is therefore of the utmost importance to identify new prognostic biomarkers capable of establishing evidence-based clinical practice recommendations for patients who are at increased risk of T2DM-related complications. Recently, epigenetic alterations have emerged as key factors in age-related diseases (ARDs), including T2DM and its complications [3, 6]. Factors associated with diabetic complications, including hyperglycaemia, oxidative stress and inflammatory factors, can modulate epigenetic mechanisms, altering gene expression in target cells, especially endothelial and immune cells. MicroRNAs (miRNAs) are a broad class of small, non-coding RNAs involved in the modulation of gene expression, either silencing or activating transcription *via* sequence-specific DNA/RNA binding [7]. Initially believed to leak passively from dead cells, miRNAs released into the bloodstream are emerging as an organized and controlled secretion process involving different shuttles [8-13]. Such mediators of epigenetic information have been extensively investigated in recent years as potential biomarkers of ARDs, including T2DM [14-19].

MiR-21 and miR-126 - now miR-21-5p and miR-126-3p, based on the latest miRBase release (V.21) - are

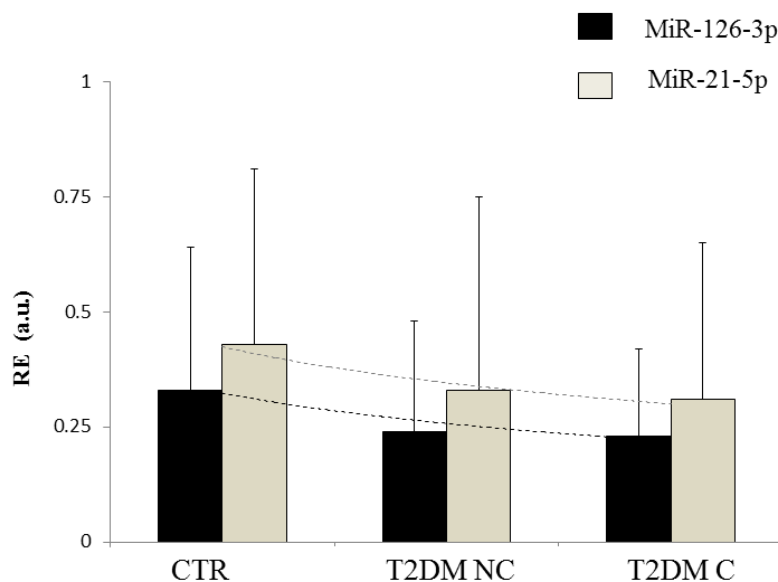
expressed mainly by cells involved in the modulation of inflammatory response and vascular homeostasis, and therefore are expected to be significantly modulated in T2DM and its complications [20]. MiR-126-3p is involved in modulating vascular regeneration and mobilization of hematopoietic stem/progenitor cells, and was therefore classified as an “angiomiR”; it was one of the first miRNAs to be found in reduced circulating concentrations in T2DM patients [14-16]. Its downregulation has been detected in endothelial tissues of diabetic mice with peripheral artery disease as well as in endothelial progenitor cells from diabetic patients and in human umbilical vein endothelial cells (HUVECs) cultured under hyperglycaemic conditions [16, 21, 22].

MiR-21-5p was initially classified as an “oncomiR”, since it is deregulated in almost all cancer types [23]. More recently it has also been implicated in other roles, including the modulation of inflammation and tissue repair, and has therefore also been classified as an “inflammamiR” [24-30].

We focused our study on miR-126-3p and miR-21-5p levels in plasma and CACs from healthy subjects and diabetic patients, to investigate their potentiality as diabetic biomarkers in relation to the main diabetic complications .

## RESULTS

The chemical-clinical, and anthropometric characteristics of the studied groups, 107 healthy CTR subjects and 193 T2DM patients, 76 patients without (T2DM NC) and 117 patients with (T2DM C) diabetic complications, are reported in table 1. Since T2DM group



**Figure 1: Circulating levels of miR-21-5p and miR-126-3p in CTR subjects and T2DM patients.** Linear trend for miR-126-3p,  $F = 9.51$ ,  $p = 0.002$ ; linear trend for miR-21-5p:  $F = 5.33$ ,  $p = 0.02$ . CTR: healthy control subjects; T2DM NC: patients without diabetic complications; T2DM C: patients with diabetic complications. RE = relative expression. Dotted line shows the exponential trend.

**Table 1: Chemical, clinical, and anthropometric characteristics of CTR subjects and patients without and with T2DM complications**

Variables	CTR (N=107)	T2DM NC (n=76)	T2DM C (n=117)
Age, (yrs)	64.25±7.56	65.56±6.96	66.51±7.48#
Males, n (%)	49 (45.79)	36 (47.37)	69 (58.97)*#
BMI, kg/m <sup>2</sup>	26.67±5.4	28.47±4.34	28.57±3.46*
Glucose, mg/dL	92.23±8.41	154.63±40.78*	178.89±56.78*#
HbA1c, %	5.96±0.41	7.34±1.28*	7.77±1.19*#
Insulin, mcU/mL	5.66±3.75	6.63±4.65	6.78±4.73
HDL cholesterol, mg/dL	59.29±15.34	55.94±17.85	51.43±15.14#
Total cholesterol, mg/dL	212.72±42.21	215.17±37.35	202.19±39.20
Creatinine, mg/dL	0.82±0.22	0.84±0.17	1.01±0.43*#
Hs-CRP, mg/L	2.52±3.77	3.79±4.25	3.54±4.24
MiR-126-3p, RE	0.33±0.31	0.24±0.24*	0.23±0.19*
MiR-21, RE	0.43±0.42	0.33±0.42*	0.31±0.34*

T2DM NC: patients without diabetic complications; T2DM C: patients with diabetic complications; RE=relative expression. The chi square test was applied for dichotomous variables. ANCOVA adjusted for age and sex or for variables significantly correlating with miRNAs was applied for continuous variables: \*p<0.05, reference group: CTR; #p<0.05, reference group: T2DM NC (diabetic patients without diabetic complications).

**Table 2: Circulating miR-21-5p and miR-126-3p levels in T2DM patients with different diabetic complications**

Diabetes complications	MiR-21 levels		
	Patients with a specific complication vs. all the other patients	Mean ± SD	p
neuropathy	37 vs. 156	0.21±0.27 vs. 0.34±0.39	0.089
nephropathy	27 vs. 166	0.26±0.28 vs. 0.33±0.39	0.489
chronic renal failure	10 vs. 183	0.39±0.41 vs. 0.31±0.38	0.375
retinopathy	84 vs. 109	0.27±0.30 vs. 0.35±0.43	0.165
lower limb arteriopathy	11 vs. 182	0.37±0.33 vs. 0.31±0.38	0.534
MACE	56 vs. 137	0.46±0.44 vs. 0.26±0.33	<b>&lt;0.001</b>
	MiR-126 levels		
neuropathy	37 vs. 156	0.18±0.20 vs. 0.24±0.22	0.122
nephropathy	27 vs. 166	0.20±0.16 vs. 0.23±0.22	0.512
chronic renal failure	10 vs. 183	0.23±0.19 vs. 0.23±0.22	0.993
retinopathy	84 vs. 109	0.23±0.20 vs. 0.23±0.23	0.863
lower limb arteriopathy	11 vs. 182	0.23±0.21 vs. 0.29±0.24	0.351
MACE	56 vs. 137	0.21±0.21 vs. 0.28±0.22	<b>0.032</b>

ANCOVA adjusted for age, sex, and number of complications. Levels lower than 0.05 in bold.

included a greater proportion of males than the CTR group, T2DM C patients were older than CTR subjects and males were a greater proportion than among T2DM NC patients as well as CTR subjects, all subsequent analyses were adjusted for age and sex.

MiR-126-3p and miR-21-5p declined significantly from CTR to T2DM NC and T2DM C (linear trend for miR-126-3p, F = 9.51, p = 0.002; linear trend for miR-21-5p: F = 5.33, p = 0.02; miR-126, ANCOVA adjusted for age and sex; F-test = 3.82, p = 0.023; miR-21-5p, ANCOVA adjusted for age, sex, glucose, and HbA1c; F-test = 4.52, p = 0.012) (Figure 1). However, the

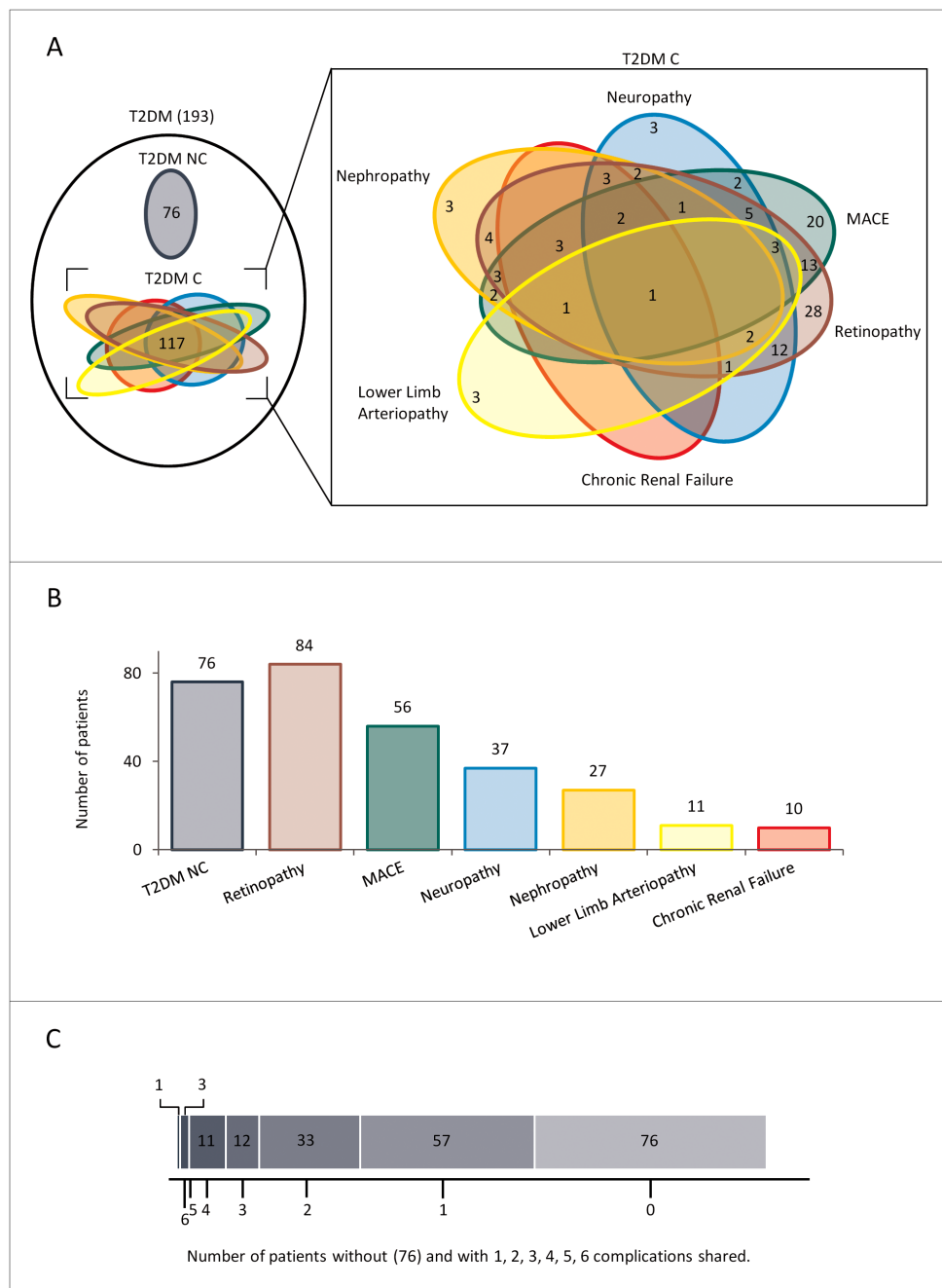
differences in miR-21-5p and miR-126-3p levels were not significant between patients without and with diabetes complications (ANCOVA adjusted for age, sex, glucose and HbA1c: F-test = 0.005, p = 0.945 and F-test = 0.612, p = 0.435, respectively).

Analysis in relation to the different T2DM complications showed no significant differences for neuropathy, nephropathy, chronic renal failure, retinopathy, or lower limb arteriopathy (table 2). However, the circulating levels of both miRNAs were significantly different between patients with and without MACE (miR-21-5p: 56 patients with vs. 137 patients without

MACE,  $0.46 \pm 0.44$  vs.  $0.26 \pm 0.33$ ,  $p < 0.001$ ; miR-126-3p:  $0.21 \pm 0.21$  vs.  $0.28 \pm 0.22$ ,  $p = 0.032$ ) (table 2). Multivariate covariance analysis with miR-21-5p and miR-126-3p as the dependent variables confirmed the significance of MACE complications (Wilk's lambda = 0.943,  $p = 0.007$ ).

Since among the 117 T2DM C patients enrolled for our study, 60 patients had more than one complications (Figure 2), to confirm the significant differences in circulating miR-21-5p and miR-126-3p found in patients

with MACE compared with those suffering from other complications, the total 193 T2DM patients were divided into 4 groups: no complications (NC,  $n = 76$ ), MACE only (MACE,  $n = 18$ ); MACE and other complications (MACE+OC,  $n = 38$ ); complications other than MACE (OC,  $n = 62$ ). Circulating miR-21-5p and miR-126-3p levels were significantly different in the 4 groups of T2DM patients (Figure 3) (ANCOVA adjusted for age, sex, glucose, and HbA1c; miR-21-5p: F-test = 8.375,  $p <$



**Figure 2: Overview of diabetic patients without or with complications.** A. Venn diagram showing the overlaps between different complications (T2DM C). B. Histogram displaying the size of each group of T2DM. C. Charts showing number of T2DM patients without complications and with one or more complications.

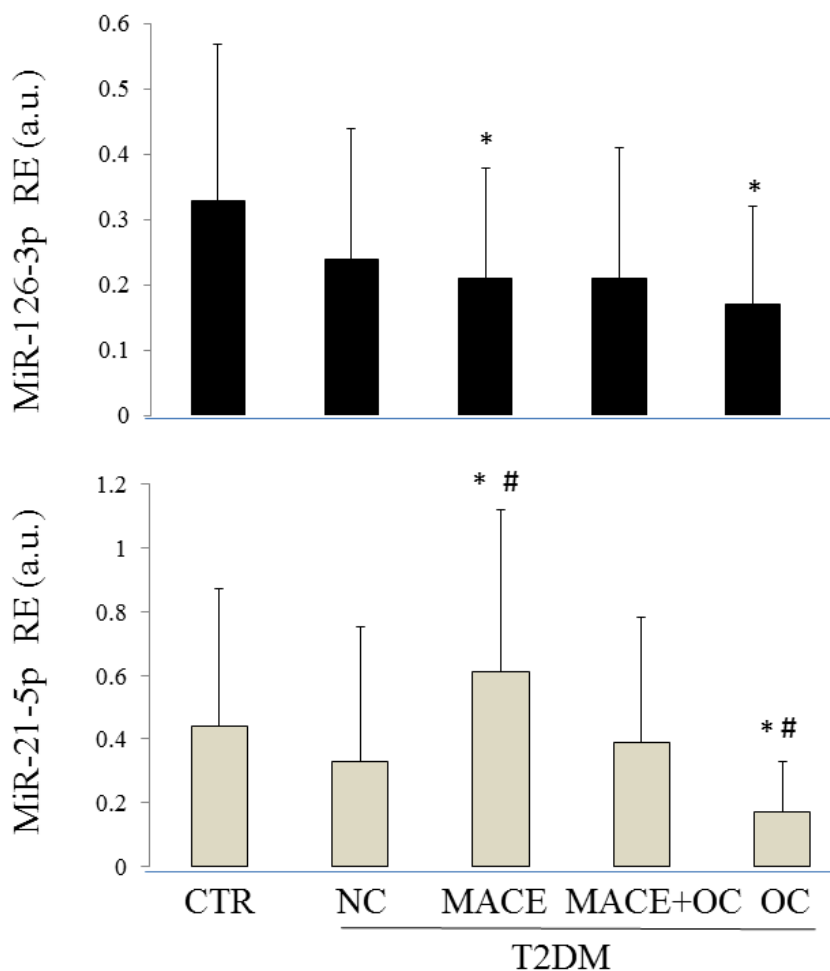
0.001; miR-126-3p: F-test = 6.655,  $p = 0.002$ ).

MiR-21-5p levels remained significantly higher in patients with MACE and significantly lower in diabetic patients + OC compared with patients without complications even after application of Bonferroni's correction, ( $p < 0.005$ ).

Circulating miR-21-5p and miR-126-3p were compared also between CTR and T2DM patients groups and were found to be significantly different (ANCOVA adjusted for age, sex, BMI, glucose, and HbA1c; circulating miR-21-5p: F-test = 6.44,  $p < 0.001$ ; circulating miR-126-3p: F-test = 4.55,  $p = 0.001$ ). Application of Bonferroni's correction using CTR as reference group confirmed that miR-21-5p levels were significantly higher and miR-126-3p levels were significantly lower in patients

with MACE. MiR-126-3p and miR-21-5p levels were both significantly lower in patients with complications other than MACE (T2DM + OC).

Correlation analysis, controlled for sex and age, between circulating miR-21-5p and miR-126-3p and the variables that were found to be significantly different between T2DM patients and healthy CTR showed weak but significant positive correlation between miR-21-5p and BMI (partial correlation coefficient, PCC; 0.12,  $p = 0.049$ ) and a weak but significant negative correlation between miR-126-3p and glycaemia parameters (miR-126-3p/glucose, PCC, -0.13,  $p = 0.038$ ; miR-126-3p/HbA1c, PCC, -0.13,  $p = 0.033$ ). Notably, a positive correlation was observed between circulating miR-21-5p and glycaemia parameters (glucose and HbA1c levels) when correlation



**Figure 3: Circulating levels of miR-21-5p and miR-126-3p in CTR and T2DM patients.** CTR: healthy control subjects; T2DM NC: patients without diabetic complications; T2DM C: patients with diabetic complications; RE = relative expression; MACE: T2DM patients with a previous MACE; MACE+OC (other complications): T2DM patients with a previous MACE and at least another diabetic complication; OC (other complications): T2DM patients with complications other than MACE. RE: relative expression; CTR: healthy control subjects; NC: diabetic patients without diabetic complications; MACE: T2DM patients with a previous MACE; MACE+OC (other complications): T2DM patients with a previous MACE and at least another diabetic complication; OC (other complications): patients with diabetic complications other than MACE. MiR-21-5p: ANCOVA adjusted for age, sex, BMI, glucose, and HbA1c; F-test = 6.44,  $p < 0.001$ ; ANCOVA adjusted for age and sex; F-test = 8.375,  $p < 0.001$ . MiR-126-3p: ANCOVA adjusted for age, sex, glucose, and HbA1c; F-test = 4.55,  $p = 0.01$ ; miR-126-3p: ANCOVA adjusted for age and sex; F-test = 6.655,  $p = 0.002$ . #Bonferroni's correction, alpha = 0.05/10,  $p < 0.005$ , reference group: T2DM NC; \*Bonferroni's correction, alpha = 0.05/10,  $p < 0.005$ , reference group: CTR.

analysis was confined to T2DM patients (miR-21/glucose, PCC, 0.14,  $p = 0.049$ ; miR-21-5p/HbA1c, PCC, 0.15,  $p = 0.045$ ).

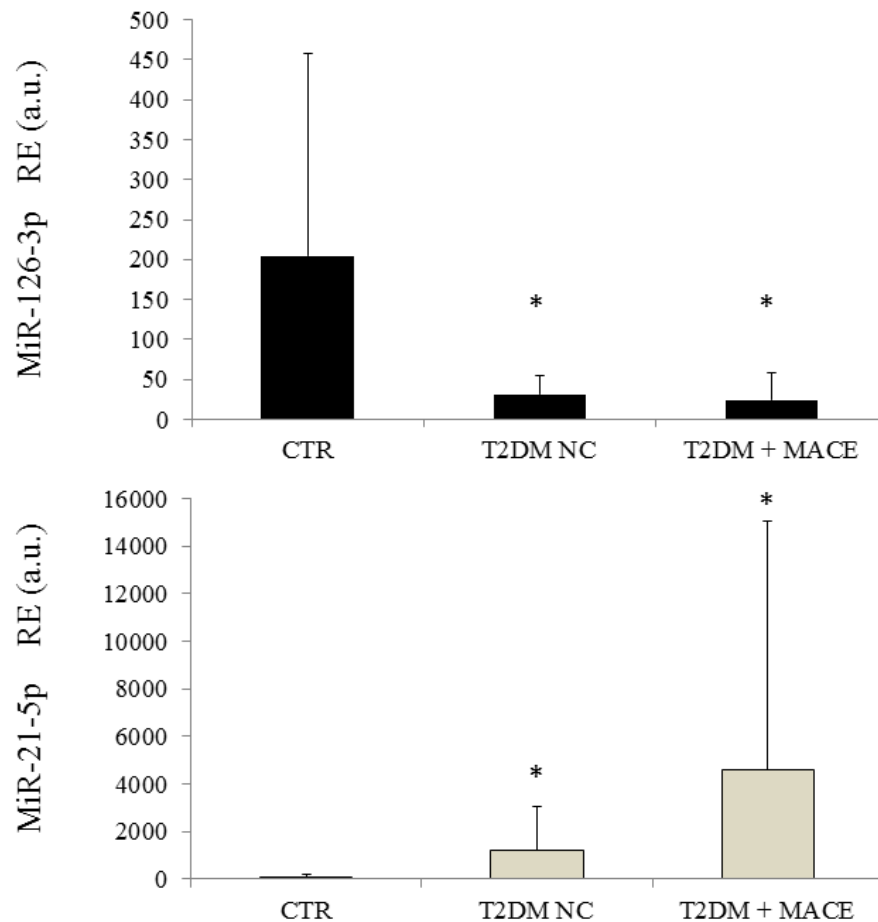
Since it has been reported that aspirin treatment may affect circulating levels of miRNAs [31], we asked our MACE patients whether they were taking aspirin treatment and found that they accounted for only 3 %.

To confirm the results obtained on plasma we analysed miR-126-3p and miR-21-5p expression levels in CACs from a subgroups of CTR, T2DM NC and T2DM + MACE. MiR-126-3p levels were significantly lower in CACs from T2DM NC and T2DM+MACE compared to CTR and miR-21-5p levels were higher in CACs from T2DM, both T2DM+MACE and T2DM NC, compared with CTR (Figure 4). Notably, the highest expression levels of miR-21 were observed in CACs from T2DM+MACE patients.

## DISCUSSION

Fasting blood glucose is the current diagnostic criterion for diabetes, being globally accepted for almost a decade. However, information regarding insulin resistance

and type 2 diabetes has increased exponentially during the last years, suggesting the existence of complex networks that regulate energy balance, in close association with the immune system and chronic inflammation [32]. A broader view of this paradigm, prompted us to research for new potential biomarkers, such as circulating miR-126-3p and miR-21-5p, since these miRNAs were associated with endothelial dysfunction and inflammation, two milestones of T2DM progression. According with previous results, we found that both miRNAs levels decreased from CTR subjects to diabetic patients [15]. An innovative aspect of our study was to analyse circulating miRNAs levels according to the presence of diabetes complications as a whole and grouped as specific complications. We observed that miR-126-3p and miR-21-5p levels declined significantly from CTR to T2DM non complicated and T2DM complicated patients. When the analysis focused on patients with specific complications, significantly higher miR-21-5p and lower miR-126-3p levels were seen in T2DM patients with previous episode of myocardial infarction (MACE). These observations are in accordance with the results of a previous prospective study showing that higher circulating and microvesicles-contained miR-



**Figure 4: MiR-21-5p and miR-126-3p expression levels in CACs from CTR, T2DM NC and T2DM +MACE patients.** T2DM NC: patients without diabetic complications; T2DM + MACE: T2DM patients with a previous MACE; RE = relative expression.



126-3p levels are protective against MACE [20]. Notably, we analysed in our retrospective study only a group of T2DM patients survived to the previous MACE.

Moreover, our and others previous data suggest that MACE is a potent inducer of miR-21-5p release in the circulation [25, 33-35]. MiR-21 transfer/ hyperexpression has been hypothesized as mechanism promoting cardiomyocyte survival during acute ischemic events, and cardiac fibroblasts proliferation after the acute event [36-38].

However, it was recently highlighted an link between miR-21 expression levels and angiogenic properties of endothelial cells; miR-21 hyper-expression suppresses endothelial progenitor cells (EPC) proliferation [39] and contribute to endothelial cell senescence [40]. Moreover, we observed a significant positive correlation between circulating miR-21-5p and BMI. Recently it was reported that obesity is related to miR-21-5p expression [41] and to pro-angiogenic potential inducing a modulation of miRNAs secretion [42], suggesting a potential link between circulating miR-21 levels, BMI and pro-angiogenic properties.

When we assessed miR-126-3p and miR-21-5p levels in CACs, a reduced miR-126-5p levels and an increased miR-21-3p expression levels were observed in T2DM patients (both T2DM NC and T2DM + MACE) compared to CTR. Previous data on endothelial progenitor cells from diabetic patients showed a reduced miR-126 expression [24], and an overexpression of miR-21 was reported in response to high glucose in endothelial cells [43].

Since miR-21 and miR-126 modulate inflammation and angiogenic pathways, our results suggest that CACs from T2DM are “proinflammatory cells” rather than “proangiogenic cells”. These conditions could depend from an increased “senescence status” of these circulating cells reached under hyperglycaemic conditions. The exposure to high glucose levels could induce the accumulation of dysfunctional endothelial senescent cells in T2DM patients shifting the transcriptional program of CACs from a “pro-angiogenic” to a “pro-inflammatory” profile, and increasing in turn the risk of developing micro and macrovascular complications. Our previous observation that telomere shortening in circulating cells from T2DM patients is related to the number of diabetic complications and that senescent CACs are present in patients with previous cardiac ischemic events [44, 45], further support this hypothesis.

## LIMITATION

Since several tissues are involved in diabetic complications, it is conceivable that all provide a relative contribution to circulating miRNA signatures. However, analysing plasma content of miRNAs it is impossible to establish which cell type contributed exactly to the

circulating levels of a specific miRNA.

Further studies are required to identify and validate circulating miRNA-based signatures associated with T2DM complications in longitudinal studies.

## CONCLUSIONS

Epigenetic mechanisms, including miRNA deregulation, probably partly explain the increased risk of developing diabetic complications over time despite multifactorial interventions with glucose-lowering, lipid-lowering and anti-hypertensive drugs [46, 47]. This study provides proof of principle that circulating-based miRNAs signatures are dynamic biomarkers of the health status as a whole, and therefore significant modulations are expected depending from comorbidities.

We recommend that the presence of comorbidities must be taken into account when circulating miRNA-based signatures are proposed as diagnostic or prognostic biomarkers for the major ARDs.

## MATERIALS AND METHODS

### Patients

A total of 193 T2DM patients from central Italy and 107 healthy control subjects (CTRs) gave their informed consent to be enrolled in the study. The study protocol was approved by the Ethics Committee of INRCA-IRCCS (Ancona, Italy). T2DM was diagnosed according to American Diabetes Association Criteria [20]. Inclusion criteria were: a body mass index (BMI) < 40 kg/m<sup>2</sup>, age 35 to 85 years, and ability and willingness to provide a written informed consent and to comply with study requirements. The information collected included information on vital signs, anthropometric data, medical history and behaviours, and exercise.

The presence/absence of diabetic complications was established as follows:

- retinopathy was defined as dilated pupils detected on funduscopy and/or fluorescence angiography;

-incipient nephropathy was a urinary albumin excretion rate > 30 mg / 24 h and normal creatinine clearance;

- chronic renal failure was defined as an estimated glomerular filtration rate < 60 mL/min per 1.73 m<sup>2</sup>;

-neuropathy was established by electromyography;

-ischemic heart disease was diagnosed by clinical history and/or ischaemic electrocardiographic alterations; these patients had had ST- or non-ST elevation myocardial infarction, which was defined as a major acute cardiac event (MACE). Mean time from the MACE was 9±8 years;

-peripheral vascular disease, including

arteriosclerosis obliterans and cerebrovascular disease, was diagnosed based on history, physical examination, and Doppler imaging.

Of the 193 T2DM patients with at least one complication, 37 had neuropathy, 11 had lower limb arteriopathy, 56 had MACE, 27 had nephropathy, 10 had chronic renal failure, and 84 had retinopathy.

Control samples (CTR) were selected among the husbands and wives of T2DM patients enrolled for the study. As microRNAs are epigenetic biomarkers, their expression is expected to be related with lifestyle and environment. Therefore, the choice of CTR among people living with diabetic patients is expected to minimize the effect of environment as confounding factor.

All study subjects reported dietary habits consistent with a Mediterranean-style diet.

### Laboratory assays

Blood concentrations of total and HDL cholesterol, triglycerides, fasting glucose, HbA1c, fasting insulin, creatinine, and hs-CRP were measured by standard procedures.

### Circulating Angiogenic Cells (CACs) isolation

CACs were isolated from 15 mL of heparinized peripheral blood. Peripheral blood mononuclear cells (PBMC) were isolated by density-gradient centrifugation with Ficoll (Ficoll-Paque™ PLUS, GE Healthcare Bio-Sciences Uppsala-Sweden), within 2 hours after blood collection.  $5 \times 10^6$  PBMC were plated on 24-well fibronectin-coated plate (BD Biosciences, Mountain View, CA) and maintained in endothelial basal medium (EBM; Clonetics-Lonza, Walkersville, MD USA) supplemented with EGM SingleQuots and 20% FCS for 4 days. After 4 days in culture, non-adherent cells were removed by PBS wash, while adherent cells were lysed directly in the culture wells for RNA purification according to the Total RNA Extraction kit (NORGEN-Germany). The CAC phenotype was confirmed in fluorescence microscopy by cellular uptake of acetylated LDL (DiI-acLDL, Molecular probes) and binding of FITC-conjugated lectin from *Ulex europaeus* (UEA-1, Sigma).

### RNA isolation and RT-qPCR assay

Prior to any therapeutic procedures, 1 ml of whole blood from each participant was collected into EDTA tubes after overnight fasting. Samples were immediately centrifuged at 3000 g for 10 min at room temperature, then centrifuged at 10,000 g for 5 min at 4 °C, and subsequently stored at -80°C until analysis. Samples were frozen within 3 h of plasma separation.

After two subsequent spins, total RNA was extracted from 100 µl of plasma using an RNA purification kit (Norgen Biotek Corporation, Thorold, ON, Canada) that isolates enriched miR species. The synthetic *Caenorhabditis elegans* miR, cel-miR-39, was spiked into plasma before RNA extraction. Only samples with cel-miR-39 recovery > 95 % were used in subsequent analyses.

MiRNAs were reverse-transcribed with the TaqMan MicroRNA Reverse Transcription Kit (Life Technologies, Carlsbad, CA, United States) as recommended by the manufacturer. MiRNA-specific TaqMan MicroRNA Assays (Life Technologies) were used for plasma miRNAs.

Data were analysed by the  $2^{-\Delta C_t}$  method. Relative expression corresponded to the  $2^{-\Delta C_t}$  value. For analysis of miRNA expression levels, external normalization to cel-miR-39 was applied.

### Statistical analysis

Summarized data are shown as mean±SD or as frequency (%). Analysis of covariance (ANCOVA) was used to compare the mean differences in chemical, clinical, and anthropometric variables after adjustment for age and sex (CTR subjects, and patients without and with diabetes complications). Multivariate analysis of covariance with miR-21-5p and miR-126-3p as the two dependent variables was performed to confirm the statistical significance of MACE complications.

Partial correlation, adjusted for age and sex, was used to test for correlations between miRNAs and the others variables. The association of T2DM complications with circulating miR-21-5p and miR-126-3p was also assessed for linear trend.

Data analysis was performed using IBM SPSS Statistics for Windows, version 20 (IBM Corp, Armonk, NY, USA). The significance level was a P value < 0.05. Bonferroni's correction for multiple testing was applied,  $\alpha = 0.05/10$ ,  $p < 0.005$ .

### ACKNOWLEDGMENTS

The authors are grateful to Word Designs for the language revision ([www.silviamodena.com](http://www.silviamodena.com)).

### FUNDING

This work was supported by grants from:  
-Grande Oriente d'Italia (GOI), Massoneria Italiana, Collegio delle Marche, Italy;  
-TERPAGE project POR Marche FESR 2007- 2013 Italy to FO;  
-Università Politecnica delle Marche, Italy, to ADP and FO.



## CONFLICTS OF INTERESTS

None of the authors have competing interests.

## Authors' contributions

GS, RL, AG, EM, LM processed plasma samples and carried out extraction and quantification of circulating microRNAs from plasma. RR, FM and FP processed and analysed miRNAs expression in CACs. MB, RT and RA recruited blood samples and managed the database. LS was responsible for statistical analysis. FO and ARB designed the study and drafted the manuscript. ADP and MB revised the manuscript critically. All authors read and approved the final manuscript.

## REFERENCES

- Huang ES, Laiteerapong N, Liu JY, John PM, Moffet HH, Karter AJ. Rates of complications and mortality in older patients with diabetes mellitus: the diabetes and aging study. *JAMA Intern Med.* 2014;174:251-258. doi: 10.1001/jamainternmed.2013.12956.
- Beltrami C, Angelini TG, Emanuelli C. Noncoding RNAs in diabetes vascular complications. *J Mol Cell Cardiol.* 2014;pii: S0022-2828(14)00426-X. doi: 10.1016/j.yjmcc.2014.12.014.
- Raciti GA, Longo M, Parrillo L, Ciccarelli M, Mirra P, Ungaro P, Formisano P, Miele C, Béguinot F. Understanding type 2 diabetes: from genetics to epigenetics. *Acta Diabetol.* 2015; 52(5):821-7. doi: 10.1007/s00592-015-0741-0.
- Wu T, Xie G, Ni Y, Liu T, Yang M, Wei H, Jia W, Ji G. Serum metabolite signatures of type 2 diabetes mellitus complications. *J Proteome Res.* 2015;14:447-56. doi: 10.1021/pr500825y.
- Boussageon R, Bejan-Angoulvant T, Saadatian-Elahi M, Lafont S, Bergeonneau C, Kassai B, Erpeldinger S, Wright JM, Gueyffier F, Cornu C. Effect of intensive glucose lowering treatment on all cause mortality, cardiovascular death, and microvascular events in type 2 diabetes: meta-analysis of randomised controlled trials. *BMJ.* 2011;343:d4169. doi: 10.1136/bmj.d4169.
- Reddy MA, Zhang E, Natarajan R. Epigenetic mechanisms in diabetic complications and metabolic memory. *Diabetologia.* 2015;58:443-55. doi: 10.1007/s00125-014-3462-y.
- Roberts TC. The MicroRNA Biology of the Mammalian Nucleus. *Mol Ther Nucleic Acids.* 2014;3:e188. doi: 10.1038/mtna.2014.40.
- Collino F, Deregibus MC, Bruno S, Sterpone L, Aghemo G, Viltono L, Tetta C, Camussi G. Microvesicles derived from adult human bone marrow and tissue specific mesenchymal stem cells shuttle selected pattern of miRNAs. *PLoS One.* 2010;5:e11803. doi: 10.1371/journal.pone.0011803.
- Zernecke A, Bidzhekov K, Noels H, Shagdarsuren E, Gan L, Denecke B, Hristov M, Köppel T, Jahantigh MN, Lutgens E, Wang S, Olson EN, Schober A, Weber C. Delivery of microRNA-126 by apoptotic bodies induces CXCL12-dependent vascular protection. *Sci Signal.* 2009;2:ra81. doi: 10.1126/scisignal.2000610.
- Vickers KC, Palmisano BT, Shoucri BM, Shamburek RD, Remaley AT. MicroRNAs are transported in plasma and delivered to recipient cells by high-density lipoproteins. *Nat Cell Biol.* 2011;13:423-33. doi: 10.1038/ncb2210.
- Arroyo JD, Chevillet JR, Kroh EM, Ruf IK, Pritchard CC, Gibson DF, Mitchell PS, Bennett CF, Pogosova-Agadjanian EL, Stirewalt DL, Tait JF, Tewari M. Argonaute 2 complexes carry a population of circulating microRNAs independent of vesicles in human plasma. *Proc Natl Acad Sci U S A.* 2011;108:5003-8. doi: 10.1073/pnas.1019055108.
- Turchinovich A, Weiz L, Langheinz A, Burwinkel B. Characterization of extracellular circulating microRNA. *Nucleic Acids Res.* 2011;39:7223-33. doi: 10.1093/nar/gkr254.
- Turchinovich A, Burwinkel B. Distinct AGO1 and AGO2 associated miRNA profiles in human cells and blood plasma. *RNA Biol.* 2012;9:1066-75. doi: 10.4161/ma.21083.
- Zhu H, Leung SW. Identification of microRNA biomarkers in type 2 diabetes: a meta-analysis of controlled profiling studies. *Diabetologia.* 2015;58(5):900-11.
- Zampetaki A, Kiechl S, Drozdov I, Willeit P, Mayr U, Prokopi M, et al. Plasma microRNA profiling reveals loss of endothelial miR-126 and other microRNAs in type 2 diabetes. *Circ Res.* 2010;107:810-817.
- Olivieri F, Bonafè M, Spazzafumo L, Gobbi M, Prattichizzo F, Recchioni R, Marcheselli F, La Sala L, Galeazzi R, Rippo MR, Fulgenzi G, Angelini S, Lazzarini R, Bonfigli AR, Bruguè F, Tiano L, Genovese S, Ceriello A, Boemi M, Franceschi C, Procopio AD, Testa R. Age- and glycemia-related miR-126-3p levels in plasma and endothelial cells. *Aging (Albany NY).* 2014;6:771-87.
- Liu Y, Gao G, Yang C, Zhou K, Shen B, Liang H, Jiang X. The role of circulating microRNA-126 (miR-126): a novel biomarker for screening prediabetes and newly diagnosed type 2 diabetes mellitus. *Int J Mol Sci.* 2014;15:10567-77. doi: 10.3390/ijms150610567.
- Zhang T, Lv C, Li L, Chen S, Liu S, Wang C, Su B. Plasma miR-126 is a potential biomarker for early prediction of type 2 diabetes mellitus in susceptible individuals. *Biomed Res Int.* 2013;2013:761617. doi: 10.1155/2013/761617.
- Guay C, Regazzi R. Circulating microRNAs as novel biomarkers for diabetes mellitus. *Nat Rev Endocrinol.* 2013;9:513-21. doi: 10.1038/nrendo.2013.86
- Jansen F, Yang X, Proebsting S, Hoelscher M, Przybilla D, Baumann K, Schmitz T, Dolf A, Endl E, Franklin

- BS, Sinning JM, Vasa-Nicotera M, Nickenig G, Werner N. MicroRNA expression in circulating microvesicles predicts cardiovascular events in patients with coronary artery disease. *J Am Heart Assoc.* 2014;3:e001249.
21. Togliatto G, Trombetta A, Dentelli P, Gallo S, Rosso A, Cotogni P, Granata R, Falcioni R, Delale T, Ghigo E, Brizzi MF. Unacylated ghrelin induces oxidative stress resistance in a glucose intolerance and peripheral artery disease mouse model by restoring endothelial cell miR-126 expression. *Diabetes.* 2015;64:1370-82.
  22. Meng S, Cao JT, Zhang B, Zhou Q, Shen CX, Wang CQ. Downregulation of microRNA-126 in endothelial progenitor cells from diabetes patients, impairs their functional properties, *via* target gene Spred-1. *J Mol Cell Cardiol.* 2012;53:64-72. doi: 10.1016/j.yjmcc.2012.04.003.
  23. Kumarswamy R, Volkmann I, Thum T. Regulation and function of miRNA-21 in health and disease. *RNA Biol.* 2011;8:706-713. doi: 10.4161/rna.8.5.16154.
  24. Olivieri F, Procopio AD, Montgomery RR. Effect of aging on microRNAs and regulation of pathogen recognition receptors. *Curr Opin Immunol.* 2014;29:29-37. doi: 10.1016/j.coi.2014.03.006.
  25. Olivieri F, Rippo MR, Monsurrò V, Salvioli S, Capri M, Procopio AD, Franceschi C. MicroRNAs linking inflammaging, cellular senescence and cancer. *Ageing Res Rev.* 2013;12:1056-1068. doi: 10.1016/j.arr.2013.05.001.
  26. Olivieri F, Spazzafumo L, Santini G, Lazzarini R, Albertini MC, Rippo MR, Galeazzi R, Abbatecola AM, Marcheselli F, Monti D, Ostan R, Cevenini E, Antonicelli R, Franceschi C, Procopio AD. Age-related differences in the expression of circulating microRNAs: miR-21 as a new circulating marker of inflammaging. *Mech Ageing Dev.* 2012;133:675-685. doi: 10.1016/j.mad.2012.09.004.
  27. Zhong X, Chung ACK, Chen HY, Dong Y, Meng XM, Li R, et al. miR-21 is a key therapeutic target for renal injury in a mouse model of type 2 diabetes. *Diabetologia* 2013;56:663-674.
  28. Duygu B, Da Costa Martins PA. miR-21: a star player in cardiachypertrophy. *Cardiovasc Res.* 2015;105(3):235-7. doi: 10.1093/cvr/cvv026.
  29. Zhou J, Zhang J. Identification of miRNA-21 and miRNA-24 in plasma as potential early stage markers of acute cerebral infarction. *Mol Med Rep.* 2014;10:971-976. doi: 10.3892/mmr.2014.2245.
  30. Recchioni R, Marcheselli F, Olivieri F, Ricci S, Procopio AD, Antonicelli R. Conventional and novel diagnostic biomarkers of acute myocardial infarction: a promising role for circulating microRNAs. *Biomarkers.* 2013;18:547-558. doi: 10.3109/1354750X.2013.833294.
  31. de Boer HC, van Solingen C, Prins J, Duijs JM, Huisman MV, Rabelink TJ, van Zonneveld AJ. Aspirin treatment hampers the use of plasma microRNA-126 as a biomarker for the progression of vascular disease. *Eur Heart J.* 2013;34:3451-7. doi: 10.1093/eurheartj/eh007.
  32. Paneni F, Costantino S, Volpe M, Lüscher TF, Cosentino F. Epigenetic signatures and vascular risk in type 2 diabetes: a clinical perspective. *Atherosclerosis.* 2013;230:191-197. doi: 10.1016/j.atherosclerosis.2013.07.003
  33. Olivieri F, Antonicelli R, Lorenzi M, D'Alessandra Y, Lazzarini R, Santini G, Spazzafumo L, Lisa R, La Sala L, Galeazzi R, Recchioni R, Testa R, Pompilio G, Capogrossi MC, Procopio AD. Diagnostic potential of circulating miR-499-5p in elderly patients with acute non ST-elevation myocardial infarction. *Int J Cardiol.* 2013;167:531-6. doi: 10.1016/j.ijcard.2012.01.075.
  34. Wang F, Long G, Zhao C, Li H, Chaugai S, Wang Y, Chen C, Wang DW. Atherosclerosis-related circulating miRNAs as novel and sensitive predictors for acute myocardial infarction. *PLoS One.* 2014;9:e105734. doi: 10.1371/journal.pone.0105734. eCollection 2014.
  35. D'Alessandra Y, Carena MC, Spazzafumo L, Martinelli F, Bassetti B, Devanna P, Rubino M, Marenzi G, Colombo GI, Achilli F, Maggolini S, Capogrossi MC, Pompilio G. Diagnostic potential of plasmatic MicroRNA signatures in stable and unstable angina. *PLoSOne.* 2013;8:e80345. doi: 10.1371/journal.pone.0080345. eCollection 2013.
  36. Yang Q, Yang K, Li A. microRNA-21 protects against ischemia-reperfusion and hypoxia-reperfusion-induced cardiocyte apoptosis *via* the phosphatase and tensin homolog/Akt-dependent mechanism. *Mol Med Rep.* 2014;9:2213-20. doi: 10.3892/mmr.2014.2068.
  37. Tu Y, Wan L, Fan Y, Wang K, Bu L, Huang T, Cheng Z, Shen B. Ischemic postconditioning-mediated miRNA-21 protects against cardiac ischemia/reperfusion injury *via* PTEN/Akt pathway. *PLoS One.* 2013;8:e75872. doi: 10.1371/journal.pone.0075872. eCollection 2013.
  38. Thum T, Gross C, Fiedler J, Fischer T, Kissler S, Bussen M, Galuppo P, Just S, Rottbauer W, Frantz S, Castoldi M, Soutschek J, Koteliansky V, Rosenwald A, Basson MA, Licht JD, Pena JT, Rouhanifard SH, Muckenthaler MU, Tuschl T, Martin GR, Bauersachs J, Engelhardt S. MicroRNA-21 contributes to myocardial disease by stimulating MAP kinase signalling in fibroblasts. *Nature.* 2008;456:980-4. doi: 10.1038/nature07511.
  39. Zuo K, Li M, Zhang X, Lu C, Wang S, Zhi K, He B. MiR-21 suppresses endothelial progenitor cell proliferation by activating the TGFβ signaling pathway *via* downregulation of WWP1. *Int J Clin Exp Pathol.* 2015;8:414-22.
  40. Dellago H, Preschitz-Kammerhofer B, Terlecki-Zaniewicz L, Schreiner C, Fortschegger K, Chang MW, Hackl M, Monteforte R, Kühnel H, Schosserer M, Gruber F, Tschachler E, Scheideler M, Grillari-Voglauer R, Grillari J, Wieser M. High levels of oncomiR-21 contribute to the senescence-induced growth arrest in normal human cells and its knock-down increases the replicative lifespan. *Aging Cell.* 2013;12:446-58.
  41. Chartoumpakis DV, Zaravinos A, Ziros PG, Iskrenova RP, Psyrogiannis AI, Kyriazopoulou VE, Habeos IG. Differential expression of microRNAs in adipose tissue

after long-term high-fat diet-induced obesity in mice. *PLoS One*. 2012;7:e34872.

42. Togliatto G, Dentelli P, Gili M, Gallo S, Deregibus C, Biglieri E, Iavello A, Santini E, Rossi C, Solini A, Camussi G, Brizzi MF. Obesity reduces the pro-angiogenic potential of adipose tissue stem cell-derived extracellular vesicles (EVs) by impairing miR-126 content: impact on clinical applications. *Int J Obes (Lond)*. 2015. doi: 10.1038/ijo.2015.123.
43. Zeng J, Xiong Y, Li G, Liu M, He T, Tang Y, Chen Y, Cai L, Jiang R, Tao J. MiR-21 is overexpressed in response to high glucose and protects endothelial cells from apoptosis. *Exp Clin Endocrinol Diabetes*. 2013;121:425-430. doi: 10.1055/s-0033-1345169.
44. Olivieri F, Antonicelli R, Recchioni R, Mariotti S, Marcheselli F, Lisa R, Spazzafumo L, Galeazzi R, Caraceni D, Testa R, Latini R, Procopio AD. Telomere/telomerase system impairment in circulating angiogenic cells of geriatric patients with heart failure. *Int J Cardiol*. 2013;164:99-105.
45. Testa R, Olivieri F, Sirolla C, Spazzafumo L, Rippo MR, Marra M, Bonfigli AR, Ceriello A, Antonicelli R, Franceschi C, Castellucci C, Testa I, Procopio AD. Leukocyte telomere length is associated with complications of type 2 diabetes mellitus. *Diabet Med*. 2011;28:1388-94.
46. Prattichizzo F, Giuliani A, Ceka A, Rippo MR, Bonfigli AR, Testa R, Procopio AD, Olivieri F. Epigenetic mechanisms of endothelial dysfunction in type 2 diabetes. *Clin Epigenetics*. 2015;7:56. doi: 10.1186/s13148-015-0090-4. eCollection 2015.
47. Prattichizzo F, Bonafe M, Ceka A, Giuliani A, Rippo MR, Re M, Antonicelli R, Procopio AD, Olivieri F. Endothelial cell senescence and inflammaging: microRNAs as biomarkers and innovative therapeutic tools. *Curr Drug Targets*. 2015.