Research Paper

Transcriptomic analysis identifies four novel receptors potentially linking endometrial cancer with polycystic ovary syndrome and generates a transcriptomic atlas

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ABSTRACT

Polycystic Ovary Syndrome (PCOS) is associated with a 3 to 4-fold increased risk of endometrial cancer (EC), but molecular mechanisms are unclear. Upregulation of the IGF1 gene in PCOS endometrium may increase EC risk, but this is uncertain. We aimed to investigate links between EC and PCOS, by analysing publicly available transcriptomic data. The NCBI Gene Expression Omnibus was used to identify relevant studies. Differentially expressed genes (DEGs) were identified and analysed using Metascape to identify pathways of interest. PCOS DEGs that encode proteins secreted into blood were identified using the Human Protein Atlas blood protein database. EC DEGs that are cellular receptors were identified using EcoTyper. These were intersected to identify which EC receptors interact with PCOS secreted proteins. Seven receptors were identified in EC but only PTPRF, ITGA2, ITGA3 and ITGB4 genes were expressed on epithelial cells. Pathway enrichment of these genes showed that the major and common pathway involved was that of the PI3K-AKT signalling pathway which was consistent with a link between PCOS and EC. However, IGF1 was down regulated in PCOS and EC. These findings hold significant promise for improving our understanding of mechanistic pathways leading to EC in PCOS.

INTRODUCTION

Endometrial cancer (EC) is the second most diagnosed cancer in women globally, with 417,000 new cases and 97,000 deaths in 2020 [1]. Obesity is a known risk factor for EC and obesity assessed by body mass index (BMI) at diagnosis is associated with worse survival among women diagnosed with EC [2]. EC is histologically divided into two subgroups: endometrioid (type 1) and non-endometrioid (type 2), with the endometrioid group oestrogen-responsive, making up about 80% of cases and often with a good outcome. The non-endometrioid group is less common, less oestrogen sensitive and associated with a poorer outcome [3].

The most frequently altered mutation in type 1 EC, is the PI3K/AKT/mammalian target of the rapamycin (mTOR) (P1K3CA) pathway [4]. Elevated IGF-1 and insulin levels have been shown to directly stimulate cell proliferation through activation of the P1K3CA pathway. Higher levels of phosphorylated IGF-I/insulin receptor gene have also been associated with EC risk factors such as diabetes in postmenopausal women [5]. The precise association between serum IGF-1 and EC is still uncertain and results from case control studies are conflicting [6, 7]. Prospective analyses are more useful than case-control studies when looking at predictors of disease risk, however these are sparse and have shown no association with total IGF1 [8, 9].

Polycystic Ovary Syndrome (PCOS), the most common female endocrinopathy (3.1–20% of women of reproductive age), is associated with obesity, infrequent menstrual periods, infertility, hirsutism, enlarged ovaries with many small follicles on ultrasound imaging, increased risks of type-2 diabetes in later life and a 3 to 4-fold

increased risk of EC [10]. Mechanisms thought to increase EC risk in PCOS include raised oestrogen levels, obesity and anovulatory menstrual cycles/lack of progesterone/ infrequent shedding of the endometrium but questions have been raised about these hypotheses [11]. Increased EC risk and endometrial dysfunction causing miscarriages in PCOS have also been linked to insulin resistance and the associated elevation in serum IGF1 in PCOS [12]. IGF-1 and IGF-2 have also been shown to promote EC cell proliferation, while metformin inhibited this proliferation. Metformin is a biguanide drug that is a first-line treatment for type 2 diabetes that has beneficial effects on various markers of the metabolic syndrome [13].

IGF1 may therefore increase EC risk in PCOS. However, this requires validation, and it is important to investigate potential confounders which affect insulin resistance (e.g., obesity). In a pilot-study [14] in Nottingham, UK, in which 102 women were recruited, they found for the first time that IGF1, IGFBP1 and PTEN gene expression were significantly up-regulated in the endometrium of PCOS, and EC women compared to controls [14]. The changes in gene expression were independent of BMI, waist hip ratio, oestradiol, and androgen levels. Protein validation test in the serum samples in the three groups were consistent with the gene findings. Genes related to the IGF1/IGFBP pathway may therefore contribute to increased risk of EC in women with PCOS. There is however a need for additional validation of these findings given the previous reports showing that serum IGF-I, IGFBP-1, IGFBP-3, and insulin levels seem unrelated to EC risk [7].

There is thankfully now an increasing amount of publicly available transcriptomic data from studies carried out in women with PCOS and endometrial cancer. Bioinformatic analyses of these data offer the potential to investigate research questions, without the need for *in-vivo* studies. The original aim of this study was to investigate the links between EC and PCOS, by analysing publicly available transcriptomic data and investigate IGF-1 and IGFBP gene expression in the endometrium of women with PCOS and EC compared with normal endometrium. Our conclusions at this stage do not support a link between IGF-1 and IGFBP genes in PCOS and EC. However, we have identified four novel receptors which may underpin the risk of EC in PCOS, and we believe our findings provide sufficient evidence to form the basis for a transcriptomic atlas to underpin future research into the links between PCOS and EC and the molecular mechanisms underpinning both diseases.

RESULTS

At the time of search, NCBI GEO search yielded 813 results for the keyword 'polycystic ovary' and 3021 results for 'endometrial cancer' in Homo sapiens datasets. Of the polycystic ovary results, 37 were expression profiling by array and 18 were by high throughout sequencing, while in endometrial cancer there were 90 expression profiling by array and 58 by high throughput sequencing. Furthermore, of the 127-expression profiling by array studies identified, only 11 were of patients and healthy controls without or prior to any drug treatment.

Several differentially expressed genes in polycystic ovary syndrome patients encode proteins secreted into the blood which have receptors in endometrial cancer cells

The cell sources and location of DEGs in polycystic ovary syndrome and endometrial cancer are different. For PCOS DEGs to exert their effect on endometrial cancer, these cells must secrete proteins into the blood that leaves the ovary and goes to the epithelial lining of the uterus. The identified DEGs of PCOS were checked against the protein atlas blood secretome database and protein coding genes were identified (Supplementary Table 1) as well as the corresponding epithelial receptors for the DEGs from endometrial cancer (Supplementary Table 2).

The protein encoding DEGs identified in PCOS were intersected with EC receptor DEGs to identify interacting ligands (Figure 1A). There were over 20 different combinations, with most intersections including one gene and several spanning more than one gene (Supplementary File 1 and Table 4 within Supplementary File 1). Most of the intersections occurred within the same disease state rather than between PCOS and EC. Similarly, the receptors found to be differentially expressed in EC were linked to that of the receptors targeted by PCOS ligands (Figure 1B). There were 14 different intersections, with many ligands targeting receptors in the ovary rather than in the uterus. 7 receptors were identified in endometrial cancer – AXL, DDR2, TLR4, PTPRF, ITGA2, ITGA3 and ITGB4 –, but only the latter four were expressed on epithelial cells (source of EC).

Most of the ligands in PCOS were down regulated in patients, with receptors being either up regulated or down regulated in EC (Figure 2A). Pathway enrichment of these genes show that the major and common pathway involved was that of the PI3K-AKT signalling pathway (Figure 2B). Other pathways related to these receptors include processes involving cellular adhesion and proliferation.

Other findings on the association between PCOS and EC

IGF1 is down regulated in epithelial cells of the endometrium of polycystic ovary syndrome patients

GSE48301 [15] looked at different endometrial cell types of PCOS and healthy women. The reanalysis of data showed that 3 DEGs were considered statistically significant. These genes are IGF1, ALDH1A1, MGP, which have different roles such as being a growth factor or are involved in alcohol metabolic pathways (Supplementary File 2 and Table 1 within Supplementary File 2). However, the top 100 genes were taken and analysed for gene enrichment analysis. The findings are shown in Figure 3. UCH proteinases are involved in an important pathway that mediates protein-protein interactions. Another pathway of interest was the STAT5 activation downstream of FLT3 ITD mutants, as STAT5 contributes to oncogenesis and other pathway members involve several PI3K-AKT pathways (Figure 3; Supplementary File 2 and Tables 2 and 3 within Supplementary File 2).

Figure 1: Intersecting the data for all (**A**) ligands from PCOS with its receptors in EC and (**B**) receptors found in endometrial cancer with ligands in PCOS.

Findings which form the basis for a transcriptomic atlas to underpin future research into PCOS

Table 1 outlines the findings which support the formation of a PCOS transcriptomic atlas. We also found DEGs in common between GSE8157 and GSE6798. There were 54 DEGs in common in skeletal muscle of polycystic patients (Supplementary Table 5), of which 44 were consistently up regulated and 7 were down regulated. However, it is of interest that Lipoprotein Lipase (LPL) and the insulin receptor (INSR) are both down regulated

Figure 2: (A) The link between PCOS ligands that are differentially expressed and the target receptors in endometrial cancer that are differentially expressed. (**B**) Pathway enrichment of the target receptors in endometrial cancer.

in skeletal muscle. The combined effect of these could explain the observed elevated serum levels of triglycerides found in polycystic patients.

Findings which form the basis for a transcriptomic atlas to underpin future research into endometrial cancer

Table 2 outlines the findings which support the formation of an EC transcriptomic atlas. While most of the endometrial cancer studies had different inclusion criteria, all studies included stage I tumours derived from endometroid cells. All DEGs were intersected to identify commonalities (Supplementary Figures 6, 7 and Supplementary Table 6) and GSE 115810 [22] was the only dataset to share some genes with some of the other datasets identified. However, the expression of these genes was different between each dataset (Supplementary File 1 and Supplementary Tables 1–3). We identified 12 genes that were shared between endometrial cancer of different tumour grades and in deceased patients. Of those genes, only 1 (CORO1B) was commonly up regulated in all datasets, 10 were commonly down regulated and one gene up regulated in one data set and down regulated in the other.

DISCUSSION

The aim of this study was to investigate the possible molecular associations between PCOS and EC by analysing data from publicly available transcriptomic data sets. We set out to further investigate findings from previous research showing that IGF-1 and IGFBP gene expression in the endometrium of women with PCOS and EC was increased compared with normal endometrium and identify other novel genes that may be commonly expressed. We were unable to confirm these specific findings from our data analysis at this stage, especially as this was an *in-silico* analysis and laboratory validation is required to support any hypothesis that can be generated from this analysis. Both IGF1 and INSR were however down regulated in PCOS and EC patients, when we looked at the DEGs that were common between all PCOS datasets and all EC datasets, except for one dataset (GSE178671) where it was up regulated in recurrent endometrial cancer stage IB in comparison to non-recurrent endometrial cancer stage IB. This finding goes against the findings from the pilot study in the UK [14] where IGF1, IGFBP1 and PTEN gene expression were significantly up regulated in the endometrium of PCOS and EC women compared to controls, so more studies are required to decipher the exact role of the insulin signalling pathway in EC risk in PCOS.

On the other hand, several of the DEGs were related to insulin and insulin-related pathways such as IGFALS, IGFB5, SYNAP5, IGF2BP2-AS1, and IGFBP4. Pathways affected by or involving insulin include GO:1990314 cellular response to insulin-like growth factor stimulus, hsa04910 Insulin signalling pathway, hsa04931 insulin resistance. It is of interest that in the study GSE178671, where recurrent and non-recurrent endometrial cancer were compared, the DEGs were involved in the IGFR1 cascade signalling, signalling by type 1 insulin-like growth factor-1 receptor, IRS-related events triggered by IGF1R, all of which are within the MAPK signalling pathway (Supplementary File 11 and Table 3 within Supplementary File 11). The MAPK signalling pathway when exposed to growth factors stimulates the cells to proliferate and grow and is a known tumour suppressor pathway. Changes in the insulin signalling pathway may therefore underpin the association between PCOS and EC as insulin resistance is a key feature of PCOS, though the precise molecular mechanisms which underpin this association remains unclear.

Despite this uncertainty, we believe our work has identified some novel findings and provide sufficient data to form the basis for a transcriptomic atlas to improve our understanding of the underlying mechanisms as well as underpin future research into PCOS and EC.

Table 1: Findings which form the basis for a PCOS transcriptomic atlas

Similar transcriptomic atlases have been published for other disease states e.g., COVID-19 [26]. For example, one interesting finding was the identification of four receptors present on epithelial cells of endometrial cancer, potentially linking EC with PCOS. To find out how PCOS may be promoting EC, one must first find out what PCOS

is putting into the blood stream that is not found in the same amounts as in healthy blood. Ideally this should be metabolites as well as excreted gene products (proteins and possibly RNAs).

From our transcriptomic atlas, the focus was on extracellular proteins. Using the human protein atlas, we

Table 2: Findings which could form the basis for an EC transcriptomic atlas

successfully identified which DEGs encoded proteins secreted in the blood, and their receptors on the epithelial lining of the uterus were identified using the online tool, EcoTyper. Four receptors (PTPRF, ITGA2, ITGA3 and ITGB4) were identified as differentially expressed in EC with ligands or gene products secreted by PCOS. While these receptors appeared to be either up or down regulated in endometrial cancer, their ligands appear to be mostly down regulated by PCOS.

Next, it was important to determine which of these molecules were known to be signalling molecules and what their effects were in terms of altering gene expression or enzyme activity (i.e., compile an effector list). Following this, it was important to determine which gene expression changes in EC were linked to a cancerous state. In terms of the identified receptors, three – ITGA2, ITGA3 and ITGB4 – appeared to be involved in the PI3K-AKT signalling pathway, which is a known oncogenic pathway that promotes various cell functions such as proliferation, cell survival and angiogenesis. PTPRF, the fourth gene, was an up regulated receptor gene in EC and

involved in various pathways linked to cellular adhesion and organization. Its ligand, NID1, a gene that plays a role in cellular adhesion and extracellular matrix organization, appeared to be down regulated in PCOS. Subsequent research to decipher the precise mechanisms by which these receptors might play a role in the association between EC and PCOS should ideally find out from the literature what molecules cause these gene expression changes, and in a pathway analysis, these can be linked with their target affected genes.

The results of this study also provided data to form the basis for a transcriptomic atlas to underpin future research into the molecular mechanisms underpinning PCOS and/or EC. For example, in PCOS, we found that in the study GSE106724 [16] that looked at the long noncoding RNA (lncRNA) and mRNA of ovarian granulosa cells of PCOS patients with either normal testosterone levels or hyperandrogenism, in comparison to control patients, the lncRNA, 'AC142528.1-001', was up regulated in all conditions of PCOS regardless of testosterone and androgen levels, and the lncRNA

'LOC100506990' was down regulated in both PCOS patients with hyperandrogenism and normal testosterone levels. This could form the basis for a novel PCOS biomarker and further illustrates the value in publishing our transcriptomic atlas at this stage.

Other findings were the association between DEGs involved in activation of GTPase activity receptor and pathways, such as response to growth factor stimulus and lutein granulosa cells in PCOS (GSE98595) [17]; pathways associated with cell adhesion, RNA splicing and response to growth factors in PCOS theca cells (GSE1615) [18]; embryo development ending in birth or egg hatching, followed by respiratory tube development and oocytes in Metaphase II in PCOS patients GSE40400 [19]; response to hormone stimulation (including the response to insulin pathway) in skeletal muscle of insulin resistant PCOS women (GSE8157) [20], the finding of a pathway involved in positive regulation of ryanodine-sensitive calcium-release channel activity in DEGs identified in the skeletal muscle of obese PCOS women GSE6798 [21] and the down regulation of LPL gene in skeletal muscle in PCOS patients (GSE6798).

Furthermore, when we investigated the pathways associated with the 54 DEGs in skeletal muscle of PCOS patients from two different studies (GSE8157 [20] and GSE6798 [21]), gene enrichment of the up regulated genes revealed that the genes were involved in: (1) activation of RAC1 downstream of NMDARs, (2) behaviour and (3) multicellular organismal homeostasis. Of these three signalling pathways, RAC1 has been found to be hyperactivated in human cancers and promotes tumour initiation, progression, and metastatic dissemination [27], and may be consistent with a link between PCOS and EC.

Findings which form the basis for a transcriptomic atlas to underpin future research into EC included the association between cellular adhesion and grades 2 and 3 EC (GSE115810) [22], microtubule-based movement in stage 1 EC (GSE17025 [26], an association between cell adhesion and migration and Wnt signalling with recurrent endometrial cancer (GSE178671) [24], the regulation of nuclear beta catenin signalling and EC survival (GSE21882) [25] and the identification of CORO1B gene as the only up regulated gene common to all datasets of EC of different tumour grades and in deceased patients, which may be involved in cell motility.

The use of gene-enrichment analysis (GEA) to identify potential pathways was a strength and helpful step to enable us to investigate our research question. We are also unaware of a similar approach to decipher the possible molecular links between PCOS and EC. A potential limitation however is that results from GEA can sometimes be misleading. The differences identified may

Figure 4: Visual representation of the datasets used in this study.

reflect the fact that each disease is caused by different factors (input signals) and DEG changes in the same direction might be coincidental or due to some other factor exerting the same change in both diseases. The same change in gene expression in both diseases might not have anything to do with one (PCOS) causing a change in the other (EC). Having identified a potential pathway, it is then crucial to look at what is happening to the expression levels of all the components. Several members of a pathway might have elevated transcript levels while others might have reduced levels or have elevated levels of inhibitory factors which nullify the effectiveness of the pathway. Further interrogation of the data presented in our transcriptomic atlas is therefore required.

Our transcriptomic atlas allows research groups globally to independently undertake further data analyses to investigate the molecular links between PCOS and EC. We identified four novel receptors which may underpin the risk of EC in PCOS, but there may well be others. Finally, both IGF1 and INSR were down regulated in PCOS and EC patients, which was not consistent with previous research. Therefore, more studies are required to decipher the exact role of the insulin signalling pathway in EC risk in PCOS.

MATERIALS AND METHODS

Institutional review board approval was not required for this study as we have used publicly available data, and the study was conducted between February and May 2022. The Gene Expression Omnibus (GEO) database from NCBI was used to identify transcriptomic studies containing the keywords 'polycystic ovary' and 'Endometrial cancer' [28]. Of the identified studies, only datasets containing expression profiling by array, and gene expression profiling by high throughput sequencing were selected for analysis.

Differentially expressed genes (DEGs) were identified using the built-in GEO2R program with the limma (Linear Models for Micorarray Analysis) R package and applying forced normalization on all data. Comparisons used for analysis were between healthy controls and cases, or different grades of tumours (Supplementary Figure 12 and Supplementary Table 10). Genes with an adjusted *P* value of ≤ 0.05 were considered as statistically significant. All DEGs were analysed using the Metascape database, a gene enrichment and analysis resource, for identification of pathways [29, 30].

PCOS DEGs encoding proteins that are secreted in the blood were identified using the Human Protein Atlas blood protein database [31]. The endometrial cancer DEGs that are epithelial receptors were identified using EcoTyper [32]. Furthermore, this tool was used to identify the receptors for the blood protein encoding PCOS genes, and the ligands for the EC receptors. An up-set plot was generated using R studio (R version 4.2.1) to identify the common secreted ligands and common receptors in PCOS and endometrial cancer. Figure 4 is a visual representation of the datasets used in this study.

Abbreviations

PCOS: Polycystic Ovary Syndrome; EC: Endometrial Cancer; DEGs: Differentially expressed genes; NCBI: National Center for Biotechnology Information; GEO: Gene Expression Omnibus; LncRNA: long noncoding RNA; mRNA: messenger RNA.

Data availability statement

Data used in this study was obtained from NCBI Gene Expression Omnibus database. All PMID and GEO accession numbers are provided. Gene descriptions and Metascape data is provided in Supplementary Files.

Author contributions

Conceptualization, W.A. and M.H.; methodology, F.A. and M.H.; formal analysis, F.A. and M.H; investigation, F.A. and M.H.; writing—original draft preparation, F.A. and W.A.; writing—review and editing, F.A., W.A, C.H., M.H; supervision, W.A., C.H., and M.H; project administration, W.A. and M.H; All authors have read and agreed to the published version of the manuscript.

CONFLICTS OF INTEREST

Authors have no conflicts of interest to declare.

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REFERENCES

- 1. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin. 2021; 71:209– 49. <https://doi.org/10.3322/caac.21660>. [[PubMed](https://pubmed.ncbi.nlm.nih.gov/33538338)]
- 2. Arem H, Irwin ML. Obesity and endometrial cancer survival: a systematic review. Int J Obes (Lond). 2013; 37:634–39. <https://doi.org/10.1038/ijo.2012.94>. [[PubMed](https://pubmed.ncbi.nlm.nih.gov/22710929)]
- 3. Wang X, Glubb DM, O'Mara TA. 10 Years of GWAS discovery in endometrial cancer: Aetiology, function and translation. EBioMedicine. 2022; 77:103895. [https://doi.](https://doi.org/10.1016/j.ebiom.2022.103895) [org/10.1016/j.ebiom.2022.103895](https://doi.org/10.1016/j.ebiom.2022.103895). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/35219087)]
- 4. Cheung LW, Hennessy BT, Li J, Yu S, Myers AP, Djordjevic B, Lu Y, Stemke-Hale K, Dyer MD, Zhang F, Ju Z, Cantley

LC, Scherer SE, et al. High frequency of PIK3R1 and PIK3R2 mutations in endometrial cancer elucidates a novel mechanism for regulation of PTEN protein stability. Cancer Discov. 2011; 1:170–85. [https://doi.org/10.1158/2159-8290.](https://doi.org/10.1158/2159-8290.CD-11-0039) [CD-11-0039](https://doi.org/10.1158/2159-8290.CD-11-0039). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/21984976)]

- 5. Merritt MA, Strickler HD, Einstein MH, Yang HP, Sherman ME, Wentzensen N, Brouwer-Visser J, Cossio MJ, Whitney KD, Yu H, Gunter MJ, Huang GS. Insulin/IGF and sex hormone axes in human endometrium and associations with endometrial cancer risk factors. Cancer Causes Control. 2016; 27:737–48. [https://doi.org/10.1007/s10552-016-](https://doi.org/10.1007/s10552-016-0751-4) [0751-4](https://doi.org/10.1007/s10552-016-0751-4). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/27125830)]
- 6. Ayabe T, Tsutsumi O, Sakai H, Yoshikawa H, Yano T, Kurimoto F, Taketani Y. Increased circulating levels of insulin-like growth factor-I and decreased circulating levels of insulin-like growth factor binding protein-1 in postmenopausal women with endometrial cancer. Endocr J. 1997; 44:419–24. <https://doi.org/10.1507/endocrj.44.419>. [[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/9279518)
- 7. Weiderpass E, Brismar K, Bellocco R, Vainio H, Kaaks R. Serum levels of insulin-like growth factor-I, IGF-binding protein 1 and 3, and insulin and endometrial cancer risk. Br J Cancer. 2003; 89:1697–704. [https://doi.org/10.1038/](https://doi.org/10.1038/sj.bjc.6601312) [sj.bjc.6601312.](https://doi.org/10.1038/sj.bjc.6601312) [\[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/14583772)
- 8. Gunter MJ, Hoover DR, Yu H, Wassertheil-Smoller S, Manson JE, Li J, Harris TG, Rohan TE, Xue X, Ho GY, Einstein MH, Kaplan RC, Burk RD, et al. A prospective evaluation of insulin and insulin-like growth factor-I as risk factors for endometrial cancer. Cancer Epidemiol Biomarkers Prev. 2008; 17:921–29. [https://doi.](https://doi.org/10.1158/1055-9965.EPI-07-2686) [org/10.1158/1055-9965.EPI-07-2686](https://doi.org/10.1158/1055-9965.EPI-07-2686). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/18398032)]
- 9. Lukanova A, Zeleniuch-Jacquotte A, Lundin E, Micheli A, Arslan AA, Rinaldi S, Muti P, Lenner P, Koenig KL, Biessy C, Krogh V, Riboli E, Shore RE, et al. Prediagnostic levels of C-peptide, IGF-I, IGFBP -1, -2 and -3 and risk of endometrial cancer. Int J Cancer. 2004; 108:262-68. [https://](https://doi.org/10.1002/ijc.11544) doi.org/10.1002/ijc.11544. [[PubMed](https://pubmed.ncbi.nlm.nih.gov/14639613)]
- 10. Haoula Z, Salman M, Atiomo W. Evaluating the association between endometrial cancer and polycystic ovary syndrome. Hum Reprod. 2012; 27:1327–31. [https://doi.org/10.1093/](https://doi.org/10.1093/humrep/des042) [humrep/des042](https://doi.org/10.1093/humrep/des042). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/22367984)]
- 11. Hardiman P, Pillay OC, Atiomo W. Polycystic ovary syndrome and endometrial carcinoma. Lancet. 2003; 361:1810–12. [https://doi.org/10.1016/s0140-](https://doi.org/10.1016/s0140-6736(03)13409-5) [6736\(03\)13409-5.](https://doi.org/10.1016/s0140-6736(03)13409-5) [\[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/12781553)
- 12. Luo L, Wang Q, Chen M, Yuan G, Wang Z, Zhou C. IGF-1 and IGFBP-1 in peripheral blood and decidua of early miscarriages with euploid embryos: comparison between women with and without PCOS. Gynecol Endocrinol. 2016; 32:538–42. <https://doi.org/10.3109/09513590.2016.1138459>. [[PubMed](https://pubmed.ncbi.nlm.nih.gov/27174569)]
- 13. Lee TY, Martinez-Outschoorn UE, Schilder RJ, Kim CH, Richard SD, Rosenblum NG, Johnson JM. Metformin as a Therapeutic Target in Endometrial Cancers. Front Oncol.

2018; 8:341. [https://doi.org/10.3389/fonc.2018.00341.](https://doi.org/10.3389/fonc.2018.00341) [\[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/30211120)

- 14. Shafiee MN, Seedhouse C, Mongan N, Chapman C, Deen S, Abu J, Atiomo W. Up-regulation of genes involved in the insulin signalling pathway (IGF1, PTEN and IGFBP1) in the endometrium may link polycystic ovarian syndrome and endometrial cancer. Mol Cell Endocrinol. 2016; 424:94– 101.<https://doi.org/10.1016/j.mce.2016.01.019>. [[PubMed](https://pubmed.ncbi.nlm.nih.gov/26802879)]
- 15. Piltonen TT, Chen J, Erikson DW, Spitzer TL, Barragan F, Rabban JT, Huddleston H, Irwin JC, Giudice LC. Mesenchymal stem/progenitors and other endometrial cell types from women with polycystic ovary syndrome (PCOS) display inflammatory and oncogenic potential. J Clin Endocrinol Metab. 2013; 98:3765–75. [https://doi.](https://doi.org/10.1210/jc.2013-1923) [org/10.1210/jc.2013-1923](https://doi.org/10.1210/jc.2013-1923). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/23824412)]
- 16. Jin L, Yang Q, Zhou C, Liu L, Wang H, Hou M, Wu Y, Shi F, Sheng J, Huang H. Profiles for long non-coding RNAs in ovarian granulosa cells from women with PCOS with or without hyperandrogenism. Reprod Biomed Online. 2018; 37:613-23. [https://doi.org/10.1016/j.rbmo.2018.08.005.](https://doi.org/10.1016/j.rbmo.2018.08.005) [\[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/30224242)
- 17. Ferrero H, Díaz-Gimeno P, Sebastián-León P, Faus A, Gómez R, Pellicer A. Dysregulated genes and their functional pathways in luteinized granulosa cells from PCOS patients after cabergoline treatment. Reproduction. 2018; 155:373–81. [https://doi.org/10.1530/REP-18-0027.](https://doi.org/10.1530/REP-18-0027) [\[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/29439093)
- 18. Wood JR, Nelson-Degrave VL, Jansen E, McAllister JM, Mosselman S, Strauss JF 3rd. Valproate-induced alterations in human theca cell gene expression: clues to the association between valproate use and metabolic side effects. Physiol Genomics. 2005; 20:233–43. [https://doi.](https://doi.org/10.1152/physiolgenomics.00193.2004) [org/10.1152/physiolgenomics.00193.2004](https://doi.org/10.1152/physiolgenomics.00193.2004). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/15598877)]
- 19. Huang X, Hao C, Shen X, Liu X, Shan Y, Zhang Y, Chen L. Differences in the transcriptional profiles of human cumulus cells isolated from MI and MII oocytes of patients with polycystic ovary syndrome. Reproduction. 2013; 145:597– 608. [https://doi.org/10.1530/REP-13-0005.](https://doi.org/10.1530/REP-13-0005) [\[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/23603633)
- 20. Skov V, Glintborg D, Knudsen S, Tan Q, Jensen T, Kruse TA, Beck-Nielsen H, Højlund K. Pioglitazone enhances mitochondrial biogenesis and ribosomal protein biosynthesis in skeletal muscle in polycystic ovary syndrome. PLoS One. 2008; 3:e2466. [https://doi.](https://doi.org/10.1371/journal.pone.0002466) [org/10.1371/journal.pone.0002466.](https://doi.org/10.1371/journal.pone.0002466) [\[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/18560589)
- 21. Skov V, Glintborg D, Knudsen S, Jensen T, Kruse TA, Tan Q, Brusgaard K, Beck-Nielsen H, Højlund K. Reduced expression of nuclear-encoded genes involved in mitochondrial oxidative metabolism in skeletal muscle of insulin-resistant women with polycystic ovary syndrome. Diabetes. 2007; 56:2349–55. [https://doi.org/10.2337/db07-](https://doi.org/10.2337/db07-0275) [0275](https://doi.org/10.2337/db07-0275). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/17563058)]
- 22. Hermyt E, Zmarzły N, Grabarek B, Kruszniewska-Rajs C, Gola J, Jęda-Golonka A, Szczepanek K, Mazurek U, Witek A. Interplay between miRNAs and Genes Associated

with Cell Proliferation in Endometrial Cancer. Int J Mol Sci. 2019; 20:6011.<https://doi.org/10.3390/ijms20236011>. [[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/31795319)

- 23. Day RS, McDade KK, Chandran UR, Lisovich A, Conrads TP, Hood BL, Kolli VS, Kirchner D, Litzi T, Maxwell GL. Identifier mapping performance for integrating transcriptomics and proteomics experimental results. BMC Bioinformatics. 2011; 12:213. [https://doi.](https://doi.org/10.1186/1471-2105-12-213) [org/10.1186/1471-2105-12-213.](https://doi.org/10.1186/1471-2105-12-213) [[PubMed](https://pubmed.ncbi.nlm.nih.gov/21619611)]
- 24. de Andrade DAP, da Silva LS, Laus AC, de Lima MA, Berardinelli GN, da Silva VD, Matsushita GM, Bonatelli M, da Silva ALV, Evangelista AF, Carvalho JP, Reis RM, Dos Reis R. A 4-Gene Signature Associated With Recurrence in Low- and Intermediate-Risk Endometrial Cancer. Front Oncol. 2021; 11:729219. [https://doi.org/10.3389/](https://doi.org/10.3389/fonc.2021.729219) [fonc.2021.729219](https://doi.org/10.3389/fonc.2021.729219). [[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/34485158)
- 25. Levan K, Partheen K, Osterberg L, Olsson B, Delle U, Eklind S, Horvath G. Identification of a gene expression signature for survival prediction in type I endometrial carcinoma. Gene Expr. 2010; 14:361–70. [https://doi.org/1](https://doi.org/10.3727/105221610x12735213181242) [0.3727/105221610x12735213181242](https://doi.org/10.3727/105221610x12735213181242). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/20635577)]
- 26. Alqutami F, Senok A, Hachim M. COVID-19 Transcriptomic Atlas: A Comprehensive Analysis of COVID-19 Related Transcriptomics Datasets. Front Genet. 2021; 12:755222. <https://doi.org/10.3389/fgene.2021.755222>. [\[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/35003208)
- 27. Colón-Bolea P, García-Gómez R, Casar B. RAC1 Activation as a Potential Therapeutic Option in Metastatic Cutaneous Melanoma. Biomolecules. 2021; 11:1554. [https://doi.](https://doi.org/10.3390/biom11111554) [org/10.3390/biom11111554](https://doi.org/10.3390/biom11111554). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/34827551)]
- 28. Barrett T, Wilhite SE, Ledoux P, Evangelista C, Kim IF, Tomashevsky M, Marshall KA, Phillippy KH, Sherman PM, Holko M, Yefanov A, Lee H, Zhang N, et al. NCBI GEO: archive for functional genomics data sets--update. Nucleic Acids Res. 2013; 41:D991–95. [https://doi.org/10.1093/nar/](https://doi.org/10.1093/nar/gks1193) [gks1193.](https://doi.org/10.1093/nar/gks1193) [\[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/23193258)
- 29. Zhou Y, Zhou B, Pache L, Chang M, Khodabakhshi AH, Tanaseichuk O, Benner C, Chanda SK. Metascape provides a biologist-oriented resource for the analysis of systemslevel datasets. Nat Commun. 2019; 10:1523. [https://doi.](https://doi.org/10.1038/s41467-019-09234-6) [org/10.1038/s41467-019-09234-6](https://doi.org/10.1038/s41467-019-09234-6). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/30944313)]
- 30. Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, Simonovic M, Doncheva NT, Morris JH, Bork P, Jensen LJ, Mering CV. STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. Nucleic Acids Res. 2019; 47:D607–13. [https://doi.](https://doi.org/10.1093/nar/gky1131) [org/10.1093/nar/gky1131](https://doi.org/10.1093/nar/gky1131). [[PubMed](https://pubmed.ncbi.nlm.nih.gov/30476243)]
- 31. Uhlén M, Karlsson MJ, Hober A, Svensson AS, Scheffel J, Kotol D, Zhong W, Tebani A, Strandberg L, Edfors F, Sjöstedt E, Mulder J, Mardinoglu A, et al. The human secretome. Sci Signal. 2019; 12:eaaz0274. [https://doi.](https://doi.org/10.1126/scisignal.aaz0274) [org/10.1126/scisignal.aaz0274.](https://doi.org/10.1126/scisignal.aaz0274) [\[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/31772123)
- 32. Luca BA, Steen CB, Matusiak M, Azizi A, Varma S, Zhu C, Przybyl J, Espín-Pérez A, Diehn M, Alizadeh AA, van de Rijn M, Gentles AJ, Newman AM. Atlas of clinically distinct cell states and ecosystems across human solid tumors. Cell. 2021; 184:5482-96.e28. [https://doi.](https://doi.org/10.1016/j.cell.2021.09.014) [org/10.1016/j.cell.2021.09.014.](https://doi.org/10.1016/j.cell.2021.09.014) [\[PubMed\]](https://pubmed.ncbi.nlm.nih.gov/34597583)