Review

Peroxisome proliferator-activated receptors (PPARs) are potential drug targets for cancer therapy

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ABSTRACT

Peroxisome-proliferator-activated receptors (PPARs) are nuclear hormone receptors including PPARα, PPARδ and PPARγ, which play an important role in regulating cancer cell proliferation, survival, apoptosis, and tumor growth. Activation of PPARs by endogenous or synthetic compounds regulates tumor progression in various tissues. Although each PPAR isotype suppresses or promotes tumor development depending on the specific tissues or ligands, the mechanism is still unclear. In this review, we summarized the regulative mechanism of PPARs on cancer progression.

INTRODUCTION

As the nuclear hormone receptor, peroxisome-proliferator-activated receptors (PPARs) consist of PPARα, PPARδ and PPARγ, which are ligand-activated transcription factors. Ligand binding and activation of PPARs heterodimerize with retinoid X receptor (RXRs) and regulate gene transcription. Although PPARs/RXRs bind to the peroxisome-proliferator response element (PPRE, consensus sequence 5′-AGGTCA N AGGTCA-3′, N being any nucleotide) of target gene promoter regions, the each PPAR isotype consensus PPRE motif is different [1–5]. PPARs play a critical role in regulation of obesity, diabetes, atherosclerosis and cancer [6–9]. Even though the PPARs family contains PPARα, PPARγ and PPARδ, they serve as different functions in tumor development. Increasing evidences show that PPARα [2, 10–12] or PPARγ [7, 8, 13] inhibits tumor progression, which acts as tumor suppressors, while some reports show that PPARα is associated with tumor progression [14–16]. In contrast, PPARδ promotes tumor development [3, 6, 17]. PPARδ is associated with ulcerative colitis (UC) and Crohn›s disease (CD), which is involved in the progression of colorectal cancer (CRC) [18, 19]. Endogenous or synthetic ligands can activate PPARδ resulting in inflammation and cancer depending on the specific ligands and tissue types [20–22]. Therefore, PPARs can be activated by endogenous or synthetic ligands, subsequently PPARs dependently or independently regulate tumor progression depending on the conditions. In this review, we discussed the progress of PPARs on tumor development.

PPARα

Lack of PPARα expressions are associated with shorter breast cancer-specific survival [23]. Our previous investigation shows that PPARα induces Bcl2 degradation leading to increased SW480 colonic cancer cell apoptosis in response to chemotherapeutic agents [10]. Glut1 plays a critical role in glucose uptake to regulate cancer cell metabolism, which is widely expressed in most types of cancer cells [24, 25]. PPARα can directly inhibit Glut1 transcription by binding Glut1 potential PPRE motif [2]. The synthetic ligands of PPARα including fenofibrate, clofibrate and wyeth14,643 suppress cell proliferation by inducing apoptosis and cell cycle arrest involved in inhibition of NFκB [26] and activation of caspase-3 [26, 27]. More importantly, the combination of wyeth-14,643 and bezafibrate significantly suppresses lung cancer cell growth [12]. In addition, N-Acetyl-Cysteine (NAC)/PPARα signaling suppresses Non-small cell lung cancer (NSCLC) cell growth involved in increased the expression of p53 [28]. Although fenofibrate promotes breast cancer cell apoptosis via NFκB-mediated activation

of caspase-3 and expression of Bad, which is independent of PPARα activity [27], clofibrate or wyeth14,643 induces hepatocarcinoma HepG2 cell apoptosis [29] and inhibits tumor progression [11] in a PPARα-dependent manner. Moreover, fenofibrate suppresses Huh7 hepatocarcinoma cell proliferation by increasing C-terminal modulator protein (CTMP) expression [27]. In addition to the inhibition of PPARα on tumor progression, PPARα−/− mice inhibit tumorigenesis involved in increased endogenous angiogenesis inhibitor thrombospondin-1(TSP-1) [14]. Endogenous PPARα ligand arachidonic acid (AA) enhances breast cancer cell proliferation by up-regulation of cyclin E levels [30]. Nesterified fatty acids (NEFAs) activate PPARα-mediated hepatocarcinogenesis [31]. Therefore, PPARα antagonist MK886 and NXT629 inhibit chronic lymphocytic leukemia (CLL) cell proliferation [15, 16]. Other reports show that clofibrate promotes ovarian and prostate cancer progression independent of PPARα [32]. These findings suggest that different agonists play diversity functions on tumor progression, sometimes they serves as reverse roles, which depends on the tissue types or PPARα ligands (Figure 1). The discrepancy is associated with the dose of ligands or types of these ligands. Therefore, it is necessary to synthesize the suitable ligands for cancer treatment, which will provide a new drug target for cancer treatment.

PPARδ

Increasing literatures show that aberrant expression of PPARδ is associated with pro-inflammatory response and tumor progression [3, 17]. Consistent with this, overexpression of PPARδ causes AOM-induced colon tumorigenesis [33], and ultraviolet (UV)-induced PPARδ expression leads to Src activation and EGFR/ERK signaling-mediated skin cancer in mice. In contrast, PPARδ−/− mice inhibit DSS-induced colonic inflammation and colitis-associated tumor growth [20], which is associated with inhibition of VEGF expression [34]. Since 14-3-3ε interacts with Bad leading to inhibition of cell apoptosis [35], PPARδ activation by PGI2, COX-2-derived prostacyclin, directly induces 14-3-3ε gene expression [36]. COX-2 inhibitors (COXIBs, indomethacin, SC-236 and isoliquiritigenin) suppress PPARδ signaling-mediated cell proliferation and tumorigenesis [17]. Wnt/β-catenin/ signaling promotes tumorigenesis by inducing PPARδ expression [18, 37], which is associated with PPARδ-mediated cyclin E1 and VEGF expression [38–40]. In contrast, APC inhibits PPARδ transcription activity [18, 41]. PPARδ induces VEGF expression leading to PPARδ activation by VEGF/PI3K/Akt pathway [40, 42, 43], suggesting that activation of PPARδ undergoes a feedback loop [20, 40]. In contrast, PPARδ-mediated tumor development is inhibited by nitric oxide donating aspirin (NO-ASA) [44]. In addition to PPARδ-mediated tumor progression, PPARδ ligand GW0742 reduces colon or breast cancer event [45, 46], this event is reversed in PPARδ−/− mice [47]. PPARδ promotes HARS-induced senescence leading to inhibition of tumorigenesis [48]. Consistent with this, silence of PPARδ results in cell proliferation and tumor growth [49]. Clinical observations show that although PPARδ protein levels are lower
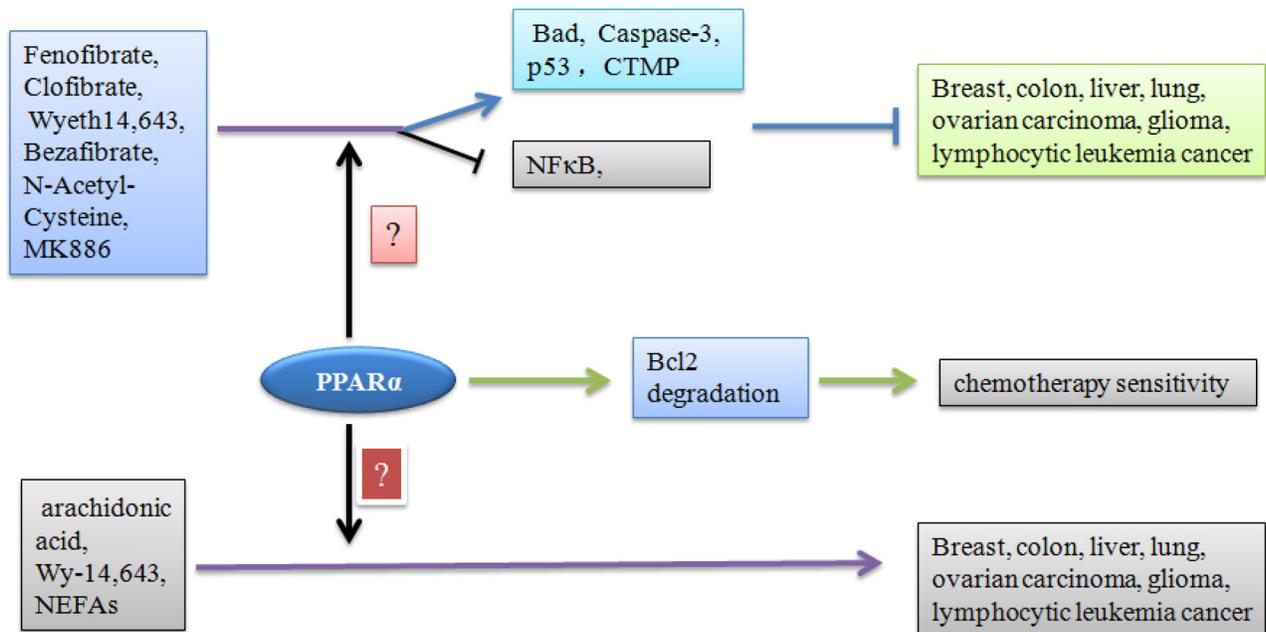

Figure 1: Effect of PPARα ligands on tumor progression. Agonists regulate different types of tumor progression in a PPARα dependent or independent manner. In addition, PPARα destructs Bcl2 function leading to increased chemotherapy sensitivity of cancer cells.
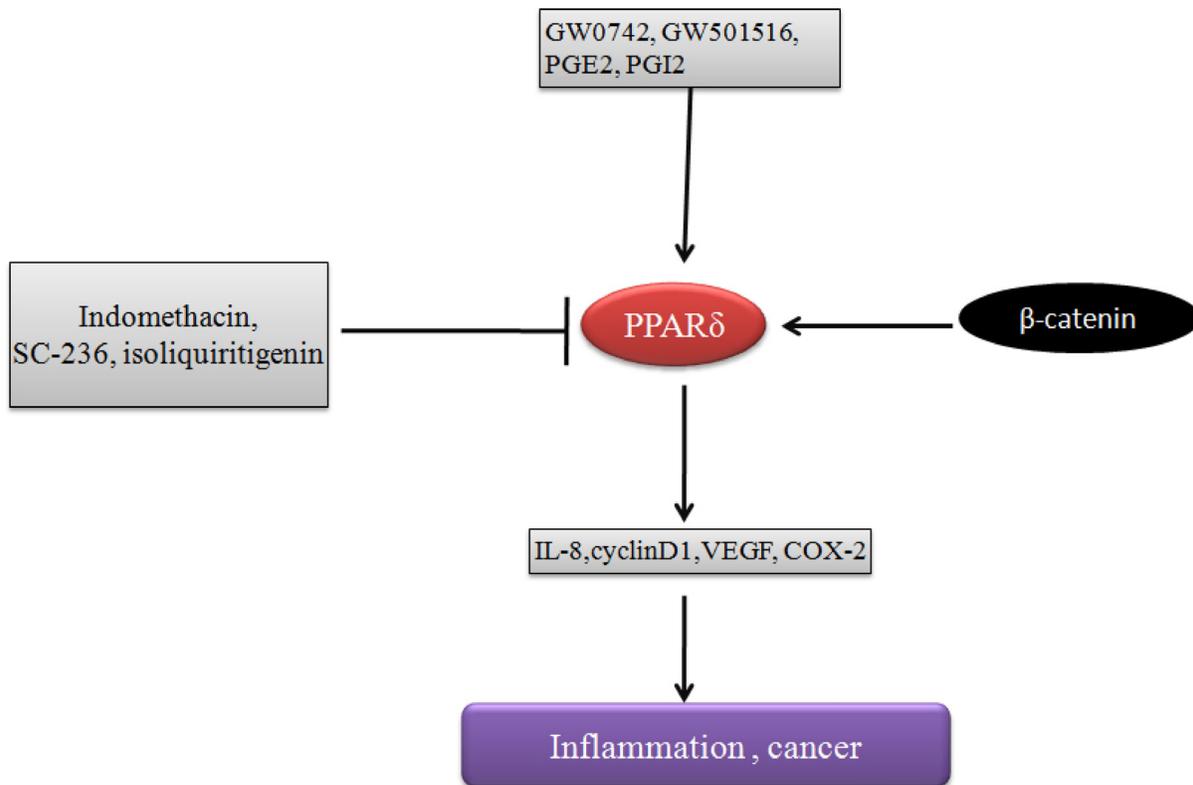
Figure 2: PPARδ promotes tumor development. Agonists of PPARδ promote inflammation and tumor development by inducing cyclin D1, IL-8, VEGF, COX-2 expression, which is inhibited by the inhibitors of COX-2 such as indomethacin, SC-236, isoliquiritigenin.
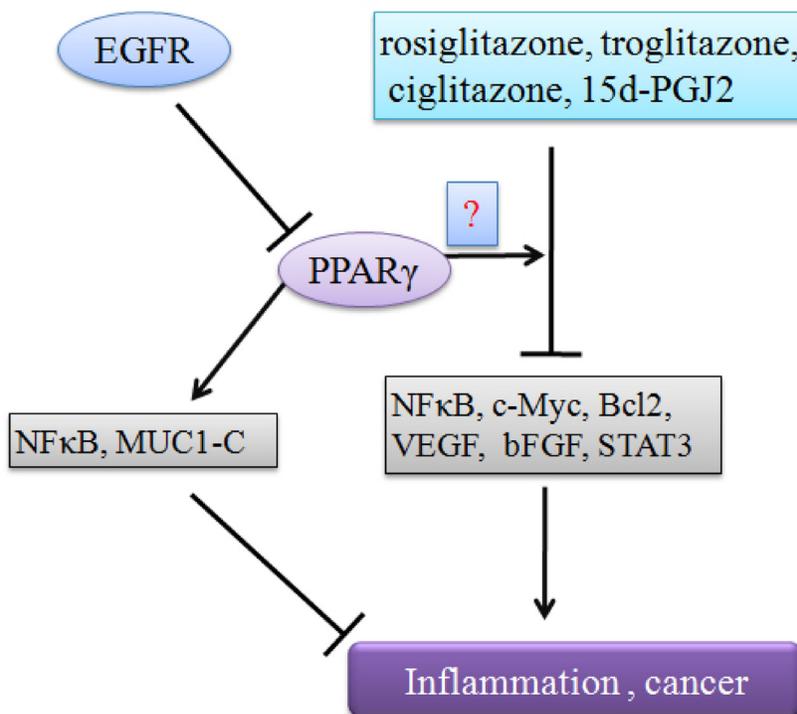

Figure 3: PPARγ negatively regulates tumor progression. Agonists regulate tumor progression in a PPARγ dependent or independent manner, which are involved in inhibition of NFκB, c-Myc, Bcl2, VEGF, bFGF, STAT3. In addition, EGFR can terminate PPARγ antitumor function.

in human colon adenocarcinomas [50], high PPARδ protein levels are benefit of colorectal cancer patients [51]. However, increasing evidences show that PPARδ promotes tumor growth [17, 20, 21, 34, 39, 40]. Taken together, PPARδ regulates tumor progression involved in multiple signaling pathways (Figure 2). It needs to further determine the physical mechanism of PPARδ on tumor development.

PPARγ

PPARγ plays an important role in inflammation, glucose metabolism and cancer [7–9]. While some clinical observations show that PPARγ expression levels are high in advanced prostate cancer (APC) tissues, ovarian, prostate and testicular carcinoma tissues [52–55], it is unclear whether the high levels of PPARγ correlate with favorite outcome in cancer patients. However, other clinical observations show that high PPARγ protein levels are benefit of colonic cancer, cervical carcinoma, follicular thyroid tumor, and esophageal cancer [9]. Consistent with this, overexpression of PPARγ inhibits cell proliferation and tumor growth, but this is reversed in PPARγ silenced cancer cells or activated EGFR signaling [7–9, 13]. PPARγ natural ligand 15-Deoxy-Δ-Prostaglandin J2(15d-PGJ$_2$) induces cell apoptosis involved in inhibition of NFκB (nuclear factor-κB) [56]. In addition, some synthetic ligands such asrosiglitazone, troglitazone and ciglitazone suppress cell proliferation by inducing apoptosis, that is involved in reduced c-Myc, Bcl2, VEGF, and bFGF expression [9]. Moreover, ciglitazone increases the effective of cisplatin on human ovarian cancer treatment [57]. However, ciglitazone and troglitazone suppress ovarian cancer cell proliferation as well as rosiglitazone induces MCF-7 breast cancer cell or pancreatic cancer cell apoptosis independent of PPARγ activity [58–60]. In addition, 15d-PGJ2 and rosiglitazone independent of PPARγ inhibit Janus Kinase (JAK)- signal transducer and activator of transcription (STAT) pathway [61]. These findings suggest that although some ligands show anti-tumor activity, they are independent of PPARγ activity with different mechanism (Figure 3). In addition, overexpression or silence of PPARγ suggests that it indeed inhibits tumor growth [7–9]. Therefore, there is a need to develop and test selective PAPRγ ligands.

Potential therapeutic targets for cancer

Increasing literatures show that PPARα or PPARγ can inhibit tumor progression by multiple pathways, which can be the potential therapeutic targets for cancer treatment, while some agonists suppress tumor progression in a PPARα/γ- independent manner (Figure 1, Figure 3). In contrast, PPARδ can promote tumor progression, so the antagonists of PPARδ may be the potential therapeutic targets for cancer treatment (Figure 2). Taken together, there is a need to develop and test selective PPARs ligands because of some agonists or antagonists independent of PPARs activity on effect of tumor development.

CONFLICTS OF INTEREST

None.

REFERENCES

1. Palmer CN, Hsu MH, Griffin HJ, Johnson EF. Novel sequence determinants in peroxisome proliferator signaling. J Biol Chem. 1995; 270:16114–21.

2. You M, Jin J, Liu Q, Xu Q, Shi J, Hou Y. PPARα Promotes Cancer Cell Glut1 Transcription Repression. J Cell Biochem. 2017; 118:1556–62.

3. Zhang W, Xu Y, Xu Q, Shi H, Shi J, Hou Y. PPARδ promotes tumor progression via activation of Glut1 and SLC1-A5 transcription. Carcinogenesis. 2017; 38:748–55.

4. Khozoie C, Borland MG, Zhu B, Baek S, John S, Hager GL, Shah YM, Gonzalez FJ, Peters JM. Analysis of the peroxisome proliferator-activated receptor-β/δ (PPARβ/δ) cistrome reveals novel co-regulatory role of ATF4. BMC Genomics. 2012; 13:665.

5. Venkatachalam G, Kumar AP, Yue LS, Pervaiz S, Clement MV, Sakharkar MK. Computational identification and experimental validation of PPRE motifs in NHE1 and MnSOD genes of human. BMC Genomics. 2009; 10:S5.

6. Michalik L, Desvergne B, Wahli W. Peroxisome-proliferator-activated receptors and cancers: complex stories. Nat Rev Cancer. 2004; 4:61–70.

7. Hou Y, Moreau F, Chadee K. PPARγ is an E3 ligase that induces the degradation of NFκB/p65. Nat Commun. 2012; 3:1300.

8. Hou Y, Gao J, Xu H, Xu Y, Zhang Z, Xu Q, Zhang C. PPARγ E3 ubiquitin ligase regulates MUC1-C oncoprotein stability. Oncogene. 2014; 33:5619–25.

9. Zhang Z, Xu Y, Xu Q, Hou Y. PPARγ against tumors by different signaling pathways. Onkologie. 2013; 36:598–601.

10. Gao J, Liu Q, Xu Y, Gong X, Zhang R, Zhou C, Su Z, Jin J, Shi H, Shi J, Hou Y. PPARα induces cell apoptosis by destructing Bcl2. Oncotarget. 2015; 6:44635–42. https://doi.org/10.18632/oncotarget.5988.

11. Pozzi A, Ibanez MR, Gatica AE, Yang S, Wei S, Mei S, Falck JR, Capdevila JH. Peroxisomal proliferator-activated receptor-alpha-dependent inhibition of endothelial cell proliferation and tumorigenesis. J Biol Chem. 2007; 282:17685–95.

12. Skrypnyk N, Chen X, Hu W, Su Y, Mont S, Yang S, Gangadhariah M, Wei S, Falck JR, Jat JL, Zent R, Capdevila JH, Pozzi A. PPARα activation can help prevent and treat non-small cell lung cancer. Cancer Res. 2014; 74:621–31.

13. Xu Y, Jin J, Zhang W, Zhang Z, Gao J, Liu Q, Zhou C, Xu Q, Shi H, Hou Y, Shi J. EGFR/MDM2 signaling promotes NF-κB activation via PPARγ degradation. Carcinogenesis. 2016; 37:215–22.

14. Kaipainen A, Kieran MW, Huang S, Butterfield C, Bielenberg D, Mostoslavsky G, Mulligan R, Folkman J, Panigrahy D. PPARalpha deficiency in inflammatory cells suppresses tumor growth. PLoS One. 2007; 2:e260.

15. Spaner DE, Lee E, Shi Y, Wen F, Li Y, Tung S, McCaw L, Wong K, Gary-Gouy H, Dalloul A, Ceddia R, Gorzcynski R. PPAR-alpha is a therapeutic target for chronic lymphocytic leukemia. Leukemia. 2013; 27:1090–99.

16. Messmer D, Lorrain K, Stebbins K, Bravo Y, Stock N, Cabrera G, Correa L, Chen A, Jacintho J, Chiorazzi N, Yan XJ, Spaner D, Prasit P, Lorrain D. A Selective Novel Peroxisome Proliferator-Activated Receptor (PPAR)-α Antagonist Induces Apoptosis and Inhibits Proliferation of CLL Cells In Vitro and In Vivo. Mol Med. 2015; 21:410–19.

17. You M, Yuan S, Shi J, Hou Y. PPARδ signaling regulates colorectal cancer. Curr Pharm Des. 2015; 21:2956–59.

18. He TC, Chan TA, Vogelstein B, Kinzler KW. PPARdelta is an APC-regulated target of nonsteroidal anti-inflammatory drugs. Cell. 1999; 99:335–45.

19. Peters JM, Morales JL, Gonzalez FJ. Modulation of gastrointestinal inflammation and colorectal tumorigenesis by peroxisome proliferator-activated receptor-β/δ (PPARβ/δ). Drug Discov Today Dis Mech. 2011; 8:e85–93.

20. Wang D, Fu L, Ning W, Guo L, Sun X, Dey SK, Chaturvedi R, Wilson KT, DuBois RN. Peroxisome proliferator-activated receptor δ promotes colonic inflammation and tumor growth. Proc Natl Acad Sci USA. 2014; 111:7084–89.

21. Hollingshead HE, Borland MG, Billin AN, Willson TM, Gonzalez FJ, Peters JM. Ligand activation of peroxisome proliferator-activated receptor-beta/delta (PPARbeta/delta) and inhibition of cyclooxygenase 2 (COX2) attenuate colon carcinogenesis through independent signaling mechanisms. Carcinogenesis. 2008; 29:169–76.

22. Berger J, Leibowitz MD, Doebber TW, Elbrecht A, Zhang B, Zhou G, Biswas C, Cullinan CA, Hayes NS, Li Y, Tanen M, Ventre J, Wu MS, et al. Novel peroxisome proliferator-activated receptor (PPAR) gamma and PPARdelta ligands produce distinct biological effects. J Biol Chem. 1999; 274:6718–25.

23. Baker BG, Ball GR, Rakha EA, Nolan CC, Caldas C, Ellis IO, Green AR. Lack of expression of the proteins GMPR2 and PPARα are associated with the basal phenotype and patient outcome in breast cancer. Breast Cancer Res Treat. 2013; 137:127–37.

24. Zhao FQ, Keating AF. Functional properties and genomics of glucose transporters. Curr Genomics. 2007; 8:113–28.

25. Young CD, Lewis AS, Rudolph MC, Ruehle MD, Jackman MR, Yun UJ, Ilkun O, Pereira R, Abel ED, Anderson SM. Modulation of glucose transporter 1 (GLUT1) expression levels alters mouse mammary tumor cell growth *in vitro* and *in vivo*. PLoS One. 2011; 6:e23205.

26. Zak Z, Gelebart P, Lai R. Fenofibrate induces effective apoptosis in mantle cell lymphoma by inhibiting the TNFalpha/NF-kappaB signaling axis. Leukemia. 2010; 24:1476–86.

27. Li T, Zhang Q, Zhang J, Yang G, Shao Z, Luo J, Fan M, Ni C, Wu Z, Hu X. Fenofibrate induces apoptosis of triple-negative breast cancer cells via activation of NF-κB pathway. BMC Cancer. 2014; 14:96.

28. Hann SS, Zheng F, Zhao S. Targeting 3-phosphoinositide-dependent protein kinase 1 by N-acetyl-cysteine through activation of peroxisome proliferators activated receptor alpha in human lung cancer cells, the role of p53 and p65. J Exp Clin Cancer Res. 2013; 32:43.

29. Maggiora M, Oraldi M, Muzio G, Canuto RA. Involvement of PPARα and PPARγ in apoptosis and proliferation of human hepatocarcinoma HepG2 cells. Cell Biochem Funct. 2010; 28:571–77.

30. Chang NW, Wu CT, Chen DR, Yeh CY, Lin C. High levels of arachidonic acid and peroxisome proliferator-activated receptor-alpha in breast cancer tissues are associated with promoting cancer cell proliferation. J Nutr Biochem. 2013; 24:274–81.

31. Tanaka N, Moriya K, Kiyosawa K, Koike K, Aoyama T. Hepatitis C virus core protein induces spontaneous and persistent activation of peroxisome proliferator-activated receptor alpha in transgenic mice: implications for HCV-associated hepatocarcinogenesis. Int J Cancer. 2008; 122:124–31.

32. Wang S, Hannafon BN, Zhou J, Ding WQ. Clofibrate induces heme oxygenase 1 expression through a PPARα-independent mechanism in human cancer cells. Cell Physiol Biochem. 2013; 32:1255–64.

33. Zuo X, Xu M, Yu J, Wu Y, Moussalli MJ, Manyam GC, Lee SI, Liang S, Gagea M, Morris JS, Broaddus RR, Shureiqi I. Potentiation of colon cancer susceptibility in mice by colonic epithelial PPAR-δ/β overexpression. J Natl Cancer Inst. 2014; 106:dju052.

34. Zuo X, Peng Z, Moussalli MJ, Morris JS, Broaddus RR, Fischer SM, Shureiqi I. Targeted genetic disruption of peroxisome proliferator-activated receptor-delta and colonic tumorigenesis. J Natl Cancer Inst. 2009; 101:762–67.

35. Liou JY, Ghelani D, Yeh S, Wu KK. Nonsteroidal anti-inflammatory drugs induce colorectal cancer cell apoptosis by suppressing 14-3-3epsilon. Cancer Res. 2007; 67:3185–91.

36. Wu KK, Liou JY. Cyclooxygenase inhibitors induce colon cancer cell apoptosis Via PPARdelta —> 14-3-3epsilon pathway. Methods Mol Biol. 2009; 512:295–307.

37. Pollock CB, Rodriguez O, Martin PL, Albanese C, Li X, Kopelovich L, Glazer RI. Induction of metastatic gastric

cancer by peroxisome proliferator-activated receptorδ activation. PPAR Res. 2010; 2010:571783.

38. Nagy TA, Wroblewski LE, Wang D, Piazuelo MB, Delgado A, Romero-Gallo J, Noto J, Israel DA, Ogden SR, Correa P, Cover TL, Peek RM Jr. β-Catenin and p120 mediate PPARδ-dependent proliferation induced by Helicobacter pylori in human and rodent epithelia. Gastroenterology. 2011; 141:553–64.

39. Hwang I, Kim J, Jeong S. β-Catenin and peroxisome proliferator-activated receptor-δ coordinate dynamic chromatin loops for the transcription of vascular endothelial growth factor A gene in colon cancer cells. J Biol Chem. 2012; 287:41364–73.

40. Jeong E, Koo JE, Yeon SH, Kwak MK, Hwang DH, Lee JY. PPARδ deficiency disrupts hypoxia-mediated tumorigenic potential of colon cancer cells. Mol Carcinog. 2014; 53:926–37.

41. Foreman JE, Sorg JM, McGinnis KS, Rigas B, Williams JL, Clapper ML, Gonzalez FJ, Peters JM. Regulation of peroxisome proliferator-activated receptor-beta/delta by the APC/beta-CATENIN pathway and nonsteroidal antiinflammatory drugs. Mol Carcinog. 2009; 48:942–52.

42. Wang D, Wang H, Shi Q, Katkuri S, Walhi W, Desvergne B, Das SK, Dey SK, DuBois RN. Prostaglandin E(2) promotes colorectal adenoma growth via transactivation of the nuclear peroxisome proliferator-activated receptor delta. Cancer Cell. 2004; 6:285–95.

43. Wang D, Wang H, Guo Y, Ning W, Katkuri S, Wahli W, Desvergne B, Dey SK, DuBois RN. Crosstalk between peroxisome proliferator-activated receptor delta and VEGF stimulates cancer progression. Proc Natl Acad Sci USA. 2006; 103:19069–74.

44. Ouyang N, Williams JL, Rigas B. NO-donating aspirin isomers downregulate peroxisome proliferator-activated receptor (PPAR)delta expression in APC(min/+) mice proportionally to their tumor inhibitory effect: implications for the role of PPARdelta in carcinogenesis. Carcinogenesis. 2006; 27:232–39.

45. Marin HE, Peraza MA, Billin AN, Willson TM, Ward JM, Kennett MJ, Gonzalez FJ, Peters JM. Ligand activation of peroxisome proliferator-activated receptor beta inhibits colon carcinogenesis. Cancer Res. 2006; 66:4394–401.

46. Yao PL, Morales JL, Zhu B, Kang BH, Gonzalez FJ, Peters JM. Activation of peroxisome proliferator-activated receptor-β/δ (PPAR-β/δ) inhibits human breast cancer cell line tumorigenicity. Mol Cancer Ther. 2014; 13:1008–17.

47. Hollingshead HE, Morimura K, Adachi M, Kennett MJ, Billin AN, Willson TM, Gonzalez FJ, Peters JM. PPARbeta/delta protects against experimental colitis through a ligand-independent mechanism. Dig Dis Sci. 2007; 52:2912–19.

48. Zhu B, Ferry CH, Blazanin N, Bility MT, Khozoie C, Kang BH, Glick AB, Gonzalez FJ, Peters JM. PPARβ/δ promotes HRAS-induced senescence and tumor suppression by potentiating p-ERK and repressing p-AKT signaling. Oncogene. 2014; 33:5348–59.

49. Yang L, Zhou J, Ma Q, Wang C, Chen K, Meng W, Yu Y, Zhou Z, Sun X. Knockdown of PPAR δ gene promotes the growth of colon cancer and reduces the sensitivity to bevacizumab in nude mice model. PLoS One. 2013; 8:e60715.

50. Foreman JE, Chang WC, Palkar PS, Zhu B, Borland MG, Williams JL, Kramer LR, Clapper ML, Gonzalez FJ, Peters JM. Functional characterization of peroxisome proliferator-activated receptor-β/δ expression in colon cancer. Mol Carcinog. 2011; 50:884–900.

51. Yang L, Zhang H, Zhou ZG, Yan H, Adell G, Sun XF. Biological function and prognostic significance of peroxisome proliferator-activated receptor δ in rectal cancer. Clin Cancer Res. 2011; 17:3760–70.

52. Rogenhofer S, Ellinger J, Kahl P, Stoehr C, Hartmann A, Engehausen D, Wieland WF, Müller SC, Hofstädter F, Walter B. Enhanced expression of peroxisome proliferate-activated receptor gamma (PPAR-γ) in advanced prostate cancer. Anticancer Res. 2012; 32:3479–83.

53. Zhang GY, Ahmed N, Riley C, Oliva K, Barker G, Quinn MA, Rice GE. Enhanced expression of peroxisome proliferator-activated receptor gamma in epithelial ovarian carcinoma. Br J Cancer. 2005; 92:113–19.

54. Kubota T, Koshizuka K, Williamson EA, Asou H, Said JW, Holden S, Miyoshi I, Koeffler HP. Ligand for peroxisome proliferator-activated receptor gamma (troglitazone) has potent antitumor effect against human prostate cancer both *in vitro* and *in vivo*. Cancer Res. 1998; 58:3344–52.

55. Hase T, Yoshimura R, Mitsuhashi M, Segawa Y, Kawahito Y, Wada S, Nakatani T, Sano H. Expression of peroxisome proliferator-activated receptors in human testicular cancer and growth inhibition by its agonists. Urology. 2002; 60:542–47.

56. Fujita M, Tohji C, Honda Y, Yamamoto Y, Nakamura T, Yagami T, Yamamori M, Okamura N. Cytotoxicity of 15-deoxy-Δ(12,14)-prostaglandin J(2) through PPARγ-independent pathway and the involvement of the JNK and Akt pathway in renal cell carcinoma. Int J Med Sci. 2012; 9:555–66.

57. Yokoyama Y, Xin B, Shigeto T, Mizunuma H. Combination of ciglitazone, a peroxisome proliferator-activated receptor gamma ligand, and cisplatin enhances the inhibition of growth of human ovarian cancers. J Cancer Res Clin Oncol. 2011; 137:1219–28.

58. Al-Alem L, Southard RC, Kilgore MW, Curry TE. Specific thiazolidinediones inhibit ovarian cancer cell line proliferation and cause cell cycle arrest in a PPARγ independent manner. PLoS One. 2011; 6:e16179.

59. Green DR. Apoptotic pathways: the roads to ruin. Cell. 1998; 94:695–98.

60. Ricciardi S, Mey V, Nannizzi S, Pasqualetti G, Crea F, Del Tacca M, Danesi R. Synergistic cytotoxicity and molecular interaction on drug targets of sorafenib and gemcitabine in human pancreas cancer cells. Chemotherapy. 2010; 56:303–12.

61. Park EJ, Park SY, Joe EH, Jou I. 15d-PGJ2 and rosiglitazone suppress Janus kinase-STAT inflammatory signaling through induction of suppressor of cytokine signaling 1 (SOCS1) and SOCS3 in glia. J Biol Chem. 2003; 278:14747–52.