

# Mitochondrial DNA sequencing and large-scale genotyping identifies *MT-ND4* gene mutation m.11696G>A associated with idiopathic oligoasthenospermia

Juan Ji<sup>1,2,3,\*</sup>, Miaofei Xu<sup>1,2,\*</sup>, Zhenyao Huang<sup>1,2,\*</sup>, Lei Li<sup>4</sup>, Hongxiang Zheng<sup>4</sup>, Shuping Yang<sup>4</sup>, Shilin Li<sup>4</sup>, Li Jin<sup>4</sup>, Xiufeng Ling<sup>1,3</sup>, Yankai Xia<sup>1,2</sup>, Chuncheng Lu<sup>1,2</sup> and Xinru Wang<sup>1,2</sup>

<sup>1</sup>State Key Laboratory of Reproductive Medicine, Institute of Toxicology, Nanjing Medical University, Nanjing 210029, China

<sup>2</sup>Key Laboratory of Modern Toxicology of Ministry of Education, School of Public Health, Nanjing Medical University, Nanjing 210029, China

<sup>3</sup>Department of Children Health Care, Nanjing Maternity and Child Health Care Hospital Affiliated to Nanjing Medical University, Nanjing 210029, China

<sup>4</sup>State Key Laboratory of Genetic Engineering and Ministry of Education Key Laboratory of Contemporary Anthropology, School of Life Sciences, Fudan University, Shanghai 200433, China

\*The first three authors have contributed equally to this study and they should be regarded as joint first authors.

**Correspondence to:** Chuncheng Lu, **email:** chunchengl@njmu.edu.cn  
Xinru Wang, **email:** xrwang@njmu.edu.cn

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## ABSTRACT

Genetic variants of mitochondrial DNA (mtDNA) were implicated to be associated with male infertility. Our previous whole mitochondrial genome sequencing and association study has identified two susceptibility mtDNA variants for oligoasthenospermia in Han Chinese men. In this study, we tested promising associations in an extended validation using 670 idiopathic oligoasthenospermia cases and 793 healthy controls to identify additional risk variants. We found that the genetic variant of m.11696G>A showed significantly higher frequency in the case group than that in the control group (odds ratio (OR) 2.21, 95% CI 1.21-4.04) ( $P=7.90\times 10^{-3}$ ). To elucidate the exact role of the genetic variants in spermatogenesis, two main sperm parameters (sperm count and motility) were taken into account. We found that m.11696G>A was associated with low sperm motility, with the OR of 2.38 (95% CI 1.27-4.46) ( $P=5.22\times 10^{-3}$ ). These results advance our understanding of the genetic susceptibility to oligoasthenospermia and more functional studies are needed to provide insights into its pathogenic mechanism.

## INTRODUCTION

Infertility is one of the most frequently diagnosed diseases in reproductive health area, and male-related problems account for approximately half of all infertility cases [1-2]. A significant proportion of idiopathic male infertility is accompanied by quantitative and/or qualitative abnormalities [3, 4]. Although several genetic factors of nuclear genome have been reported to be involved in spermatogenic impairment [5, 6], only few

genetic variants of mitochondrion genome have been identified to be associated with spermatogenesis and sperm maturation [7].

As mitochondria are the major source of ATP, they play critical roles in spermatogenesis, differentiation and optimal functioning of germ cells [8]. The 16,569 bp circular human mtDNA encodes two tRNAs, 22 rRNAs, and 13 polypeptides, which are necessary for the proper assembly and function of the mitochondrial complexes of oxidative phosphorylation (OXPHOS) [9, 10]. Sequence

polymorphisms in the human mtDNA are significantly related with the geographic origin of the indigenous populations. These mtDNA variants form clusters of related mtDNA haplotypes defined as mtDNA haplogroups [11]. Certain mtDNA haplogroups are associated with specific disease phenotypes, such as Alzheimer's disease and Parkinson's disease [12, 13]. Several studies have reported that mutations in the mtDNA resulted in either functionless or malfunctioning proteins, hence affected sperm motility in different population [14]. Hence, in this study, we hypothesize that mitochondrial genetic variants may be associated with idiopathic oligoasthenospermia in Han Chinese men.

Recently, we conducted a two-stage study to systematically elucidate the potential role of mtDNA genetic variants in oligoasthenospermia based on using next-generation sequencing (NGS) in the discovery phase and SNPscan in the follow-up validation phase [15]. Two mtDNA genetic variants, m.16179C>T and m.12361A>G, were identified to be associated with low sperm count or motility [15].

Here we evaluated promising associations in an extended validation using 670 cases and 793 controls. We focused on the single nucleotide polymorphisms (SNPs) that have *P* values ranging from 0.1-0.2 in the solexa sequencing stage, reported in our previous study [15]. These results will advance our understanding of the susceptibility to oligoasthenospermia in Han Chinese men.

## RESULTS

### Characteristics of the study population

The final population consisted of 1463 Han Chinese subjects, composed of 793 fertile controls and 670 patients. The distributions of selected characteristics among the case and control subjects were presented in Table 1. No significant differences were identified between the case group and the control group regarding all selected variables (including age, smoking, drinking, tea consumption and BMI).

### Mitochondrial DNA haplogroup distribution in case group and control group

To assess whether some genetic background are predisposing to or protecting against spermatogenic impairment, we first investigated the mtDNA haplogroups distributions among the case and control groups in the population. Twelve main haplogroups, including A, B, C, D, F, G, M\*, M7, M8, M9, N\* and N9 (Supplementary Figure 1), were detected in our study subjects. The detailed mtDNA haplogroups distributions were shown in Table 2. Compared to the control group, no significant distribution difference was found between these two groups. These results demonstrated that the genetic background (mainly mtDNA haplogroups) would not influence the susceptibility of studied subjects to spermatogenic impairment.

### Validation of oligoasthenospermia susceptible mtDNA variants

Through NGS of mtDNA genome, six SNPs met the selection criteria for the validation stage (Table 3). Additive models of logistic regression analyses were used to estimate the *P* values of association analyses. For the exploratory purpose and due to a relatively small sample size in this analysis,  $0.1 < P \leq 0.2$  were considered statistically suggestive. The frequency distribution of these six SNPs identified in the whole mitochondrial genome sequencing and the validations were shown in Table 4. The frequency distribution of m.11696G>A was at a significantly increased risk of oligoasthenospermia compared with the controls (OR 2.21, 95%CI 1.21-4.04) ( $P = 7.90 \times 10^{-3}$ ). To the other genetic variants, no significant differences of distribution frequencies were identified between the two groups.

To uncover the exact role of the genetic variants in spermatogenesis, two main sperm parameters (sperm count and motility) were taken into consideration. According to these two parameters, the cases were further classified into two subgroups respectively. As shown in Table 4, the frequency of m.11696G>A was higher in the case group (with sperm concentration  $< 15 \times 10^6/\text{ml}$ ) than that in the control group (OR 2.25; 95 % CI 1.15-4.42) ( $P = 1.56 \times 10^{-2}$ ), although the significant differences were not retained after Bonferroni adjustment. As to the sperm motility, logistic regression analysis revealed that only m.11696G>A was associated with a significantly increased risk of asthenospermia (characterized by reduced sperm motility, with sperm motility  $< 40\%$  motile sperm) [16] with the OR value 2.38 (95% CI 1.27-4.46) ( $P = 5.22 \times 10^{-3}$ ) (Table 5). Due to the low occur frequency and limited sample size in this study, although m.3398C>T showed decreased risk of asthenospermia ( $P = 8.40 \times 10^{-3}$ ), validations are still needed in a larger population (Table 5).

## DISCUSSION

The role of mitochondria in spermatogenesis has been extensively researched [17, 18]. However, former studies paid close attention to single or specific genetic variants of mitochondrial genes participated in spermatogenesis [19, 20]. To systematically explore the role of whole mtDNA genome on spermatogenesis, NGS was applied in 233 idiopathic oligoasthenospermia cases and 233 healthy controls, and susceptible genetic variants were evaluated with SNPscan in 670 cases and 793 controls.

It has been demonstrated that mtDNA haplogroup R was a strong independent predictor of sperm motility [21], while others thought that there was no effect of mtDNA haplotype on sperm velocity [22]. Considering above, the frequency distribution of mtDNA haplogroups between the two groups were firstly investigated to study whether population heterogeneity were confounders in identifying candidate mtDNA genetic variants on spermatogenic

**Table 1: The distributions of selected variables among cases and control subjects**

Variables	Frequency	Control fertility/ normozoospermia (n = 793)	Oligoasthenospermia (n = 670)	P
<sup>a</sup> Age(years), mean±SD	-	28.76±3.55	28.52±4.20	2.36×10 <sup>-1</sup>
<sup>b</sup> Smoking	Ever	404 (50.95%)	342 (51.04)	1.00
	Never	389 (49.05%)	328 (48.96)	
<sup>b</sup> Drinking	Ever	412 (51.95%)	352 (52.54)	8.33×10 <sup>-1</sup>
	Never	381 (48.05%)	318 (47.46)	
<sup>b</sup> Tea consumption	Ever	437 (55.11%)	368 (54.93)	9.58×10 <sup>-1</sup>
	Never	356 (44.89%)	302 (45.07)	
<sup>a</sup> BMI, mean±SD	-	23.62±3.92	23.70±3.04	6.67×10 <sup>-1</sup>

<sup>a</sup> Independent-samples T-test was used to test for differences in continuous variables such as age and body mass index (BMI) between the cases and controls.

<sup>b</sup> Two-sided chi-squared test was used to test the differences of categorical variables such as drinking, smoking status and tea consumption between cases and controls.

**Table 2: Distributions of mtDNA haplogroups among the control group and the case group**

mtDNA haplogroups	Case infertility/ oligoasthenospermia (n = 670)		Control fertility/ normozoospermia (n = 793)		OR (95%CI) <sup>a</sup>	P <sup>a</sup>
	n	%	n	%		
A	30	4.48%	34	4.29%	1.05 (0.63-1.73)	8.59×10 <sup>-1</sup>
B	101	15.07%	104	13.11%	1.18 (0.88-1.58)	2.82×10 <sup>-1</sup>
C	25	3.73%	31	3.91%	0.95 (0.56-1.63)	8.60×10 <sup>-1</sup>
D	167	24.93%	188	23.71%	1.07 (0.84-1.36)	5.88×10 <sup>-1</sup>
F	88	13.13%	115	14.50%	0.89 (0.66-1.20)	4.51×10 <sup>-1</sup>
G	27	4.03%	33	4.16%	0.97 (0.58-1.63)	8.99×10 <sup>-1</sup>
M*	45	6.72%	57	7.19%	0.93 (0.62-1.39)	7.24×10 <sup>-1</sup>
M7	47	7.01%	60	7.57%	0.92 (0.62-1.37)	6.87×10 <sup>-1</sup>
M8	43	6.42%	42	5.30%	1.23 (0.79-1.90)	3.61×10 <sup>-1</sup>
M9	10	1.49%	15	1.89%	0.79 (0.35-1.76)	5.57×10 <sup>-1</sup>
N*	20	2.99%	20	2.52%	1.19 (0.63-2.23)	5.88×10 <sup>-1</sup>
N9	33	4.93%	37	4.67%	1.06 (0.65-1.71)	8.17×10 <sup>-1</sup>
Others	34	5.07%	57	7.19%	0.69 (0.45-1.07)	9.54×10 <sup>-2</sup>

<sup>a</sup> ORs and P value were obtained from multivariate logistic regression analysis.

**Table 3: Screening predisposed mtDNA variations through solexa sequencing**

Gene	Position	Variant	Amino acid	Case infertility/ oligoasthenospermia (233)	Control fertility/ normozoospermia (233)	OR (95%CI) <sup>a</sup>	P <sup>a</sup>
ND1	3398	T to C	M-T	233/0	230/3	-	1.24×10 <sup>-1</sup>
ND2	5263	C to T	A-V	226/7	229/4	2.37 (0.61-9.30)	1.69×10 <sup>-1</sup>
CO2	7805	G to A	V-I	230/3	233/0	-	1.24×10 <sup>-1</sup>
ATP6	9053	G to A	S-N	210/23	217/16	1.49 (0.76-2.89)	1.58×10 <sup>-1</sup>
ATP6	9128	T to C	I-T	229/4	225/8	0.49 (0.15-1.65)	1.91×10 <sup>-1</sup>
ND4	11696	G to A	V-I	220/13	227/6	1.12 (0.83-5.99)	1.01×10 <sup>-1</sup>

<sup>a</sup> ORs and P value were obtained from multivariate logistic regression analysis.

**Table 4: Association of six identified genetic variations in solexa sequence with oligoasthenospermia according to sperm concentration**

Position	Genotype	Control	Case/idiopathic infertility								
			Oligoasthenospermia			Sperm concentration					
			n (670)	OR (95%CI) <sup>a</sup>	P <sup>b</sup>	n (287)	OR (95%CI) <sup>a</sup>	P <sup>b</sup>	n (383)	OR (95%CI) <sup>a</sup>	P <sup>b</sup>
3398	T/C	783/10	666/4	0.47 (0.15-1.51)	2.82×10 <sup>-1</sup>	285/2	0.55 (0.12-2.52)	7.43×10 <sup>-1</sup>	381/2	0.41 (0.09-1.89)	3.56×10 <sup>-1</sup>
5263	C/T	784/9	660/10	1.33 (0.54-3.29)	5.38×10 <sup>-1</sup>	285/2	0.61 (0.13-2.85)	7.37×10 <sup>-1</sup>	375/8	1.86 (0.71-4.85)	2.02×10 <sup>-1</sup>
7805	G/A	792/1	666/4	4.76 (0.53-42.66)	1.85×10 <sup>-1</sup>	286/1	2.77 (0.17-44.42)	4.61×10 <sup>-1</sup>	380/3	6.25 (0.65-60.31)	1.04×10 <sup>-1</sup>
9053	G/A	742/51	615/55	1.30 (0.88-1.93)	1.91×10 <sup>-1</sup>	264/23	1.27 (0.76-2.11)	3.63×10 <sup>-1</sup>	351/32	1.33 (0.84-2.10)	2.27×10 <sup>-1</sup>
9128	T/C	781/12	663/7	0.69 (0.27-1.76)	4.30×10 <sup>-1</sup>	284/3	0.69 (0.19-2.45)	7.71×10 <sup>-1</sup>	379/4	0.69 (0.22-2.14)	6.02×10 <sup>-1</sup>
11696	G/A	776/17	<b>639/31</b>	<b>2.261</b> <b>(1.21-4.04)</b>	<b>7.90×10<sup>-3</sup></b>	274/13	2.17 (1.03-4.52)	3.50×10 <sup>-2</sup>	365/18	2.25 (1.15-4.42)	1.56×10 <sup>-2</sup>

<sup>a</sup> ORs were obtained from multivariate logistic regression analysis.

<sup>b</sup> P value was obtained from multivariate logistic regression analysis. The significance level was set to 0.05/6 = 0.008, using a Bonferroni correction.

impairment. In our study, most of these haplogroups are the subgroups of macro-haplogroup M and macro-haplogroup N. The haplogroup frequencies are in line with the frequencies in East Asia. And no significant differences were found in the mtDNA haplogroups distribution. It might be caused by investigating two different cases, asthenospermia and oligoasthenospermia, respectively. Also, Ruiz-Pesini supported that asthenozoospermia, but not oligozoospermia, was associated with mtDNA haplogroups in whites [23]. In other words, the genetic backgrounds may not affect our results of the present association study.

Our results demonstrated that genetic variant m.11696G>A was associated with risk of asthenospermia. The G-to-A transition at position 11696 (m.11696G>A)

in the *MT-ND4* gene resulting in the substitution of an isoleucine for valine at amino acid position 313 is located in the predicted transmembrane region (Figure 1) [24]. Besides, it was reported that this mutation was related to mitochondrial diseases such as Leber hereditary optic neuropathy with dystonia and deafness [25–27]. This LHON-associated mtDNA mutation was first identified to be heteroplasmy in a large Dutch family [24]. Interspecies comparison does not show significant conservation of this valine, and in most mammals a threonine residue is found at this position (Figure 2).

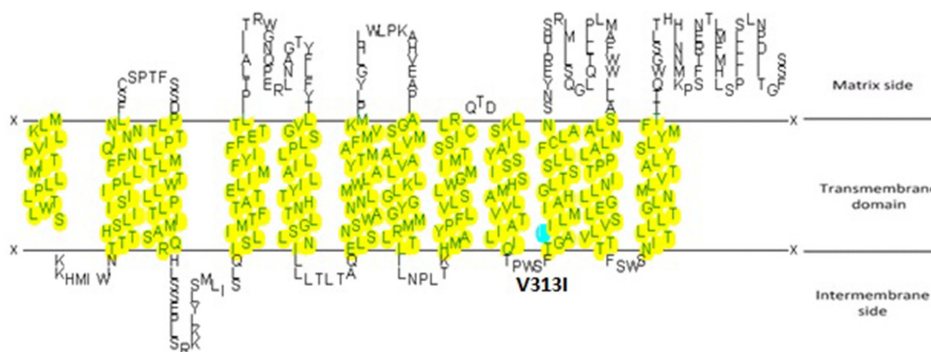
Complex I (NADH-ubiquinone oxidoreductase) is the major entry point of electrons into the electron transport chain and contributes to the establishment of

**Table 5: Association of six identified genetic variations in solexa sequence with oligoasthenospermia according to sperm motility**

Position	Genotype	Control	Case/idiopathic infertility								
			Oligoasthenospermia			Sperm motility					
			n (670)	OR (95%CI) <sup>a</sup>	P <sup>b</sup>	≥40 <sup>a</sup>		<40 <sup>a</sup>			
			n (166)	OR (95%CI) <sup>a</sup>	P <sup>b</sup>	n (504)	OR (95%CI) <sup>a</sup>	P <sup>b</sup>			
3398	T/C	783/10	666/4	0.47 (0.15-1.51)	2.82×10 <sup>-1</sup>	162/4	1.93 (0.60-6.24)	2.94×10 <sup>-1</sup>	504/0	-	8.40×10 <sup>-3</sup>
5263	C/T	784/9	660/10	1.33 (0.54-3.29)	5.38×10 <sup>-1</sup>	164/2	1.06 (0.23-4.96)	1.00	496/8	1.41 (0.54-3.67)	6.18×10 <sup>-1</sup>
7805	G/A	792/1	666/4	4.76 (0.53-42.66)	1.85×10 <sup>-1</sup>	166/0	-	1.00	500/4	6.33 (0.71-56.85)	7.81×10 <sup>-2</sup>
9053	G/A	742/51	615/55	1.30 (0.88-1.93)	1.91×10 <sup>-1</sup>	152/14	1.34 (0.72-2.48)	3.95×10 <sup>-1</sup>	463/41	1.29 (0.84-1.97)	2.44×10 <sup>-1</sup>
9128	T/C	781/12	663/7	0.69 (0.27-1.76)	4.30×10 <sup>-1</sup>	166/0	-	2.39×10 <sup>-1</sup>	497/7	0.92 (0.36-2.34)	1.00
11696	G/A	776/17	<b>639/31</b>	<b>2.261</b> <b>(1.21-4.04)</b>	<b>7.90×10<sup>-3</sup></b>	160/6	1.71 (0.66-4.41)	2.64×10 <sup>-1</sup>	<b>479/25</b>	<b>2.38</b> <b>(1.27-4.46)</b>	<b>5.22×10<sup>-3</sup></b>

<sup>a</sup> ORs and P value were obtained from multivariate logistic regression analysis.

<sup>b</sup> P value was obtained from multivariate logistic regression analysis. The significance level was set to 0.05/6 = 0.008, using a Bonferroni correction.



**Figure 1: The secondary structure changes of variant m. 11696G>A are predicted by the SOSUI system (<http://sosui.proteome.bio.tuat.ac.jp>). m.11696G>A would replace the amino acid residue valine at position 313 with isoleucine, which lies in the transmembrane region of the *MT-ND4* subunit.**

Homo sapiens (Human)	WSF TG A <b>V</b> ILMIAH
Patient	WSF T G A I ILMIAH
Pan troglodytes (Chimpanzee)	WSF T G A I ILMIAH
Canis lupus familiaris (Dog)	WSYMGA T ALMIAH
Equus caballus (Horse)	WSYMGA T ALMIAH
Mus musculus (Mouse)	WSFMGA T MLMIAH
Felis catus (Cat)	WSYMGA T ALMIAH
Equus asinus (Ass)	WSYMGA T ALMIAH
Oryctolagus cuniculus (Rabbit)	WSFMGA T ALMIAH
Gallus gallus (Chicken)	WSF SGA M ILMISH

▲  
**V313I(m.11696 G>A)**

**Figure 2: Alignment of the ND4 protein in different species showing the conservation of the amino acid 313.**

a proton gradient that is required for the bulk of cellular ATP synthesis [28]. This enzyme, has an overall L-shaped structure, with one arm, which contains all the subunits encoded by the mtDNA, buried in the mitochondrial inner membrane, and the other arm, which contains the catalytic center, protruding into the mitochondrial matrix [29]. Complex I is composed of 45 subunits, including mitochondrial encoded NADH dehydrogenase subunit (MTND) genes [30]. The seven MTND genes (*MTND1*, *MTND2*, *MTND3*, *MT-ND4L*, *MT-ND4*, *MTND5*, *MTND6*) comprise 38% of the total mtDNA, spanning over 6000 bases of the mitochondrial genome [31]. Complex I accepts electrons from NADH, transfers them to ubiquinone (coenzyme Q10) and uses the energy released to pump protons out across the mitochondrial inner membrane [32]. Defects of complex I are the most common biochemical abnormalities in patients with mitochondrial respiratory chain disorders. The first-ever mtDNA point mutation was described in an MTND gene, and over the past 17 years, lots of pathogenical point mutations in the *MT-ND4* gene have been reported [33–35].

In brief, our study demonstrated that genetic variant m.11696G>A increased the risk of asthenospermia. It is reasonable to surmise that some mtDNA genetic variants in the NADH dehydrogenase genes may cause spermatogenesis failure by decreasing activities of mitochondrial respiratory chain complexes. These findings may contribute to understanding the etiology of male infertility and further functional studies are still needed to support our findings.

## MATERIALS AND METHODS

### Study population and sample collection

This study was approved by the Ethics Review Board of Nanjing Medical University. The protocol and consent form were approved by the Institutional Review Board of Nanjing Medical University prior to the study. After the study procedures were explained and all questions were answered, all of the subjects signed informed consent forms.

We performed a two-stage case-control analysis. The whole mitochondrial genome solexa sequencing phase included 233 idiopathic oligoasthenospermia cases and 233 healthy controls which were consecutively recruited from Affiliated Hospitals of Nanjing Medical University (NJMU Infertile Study). The details of solexa sequencing were described in Lu et al 2015. For validations, we enlarged the sample size by testing 670 oligozoospermia patients and 793 controls, which were continuously enlisted from Renji Hospital. All infertile male subjects were genetically unrelated Han Chinese men and selected based on an andrological examination, including examination of medical history, physical examination, semen analysis, scrotal ultrasound, hormone

analysis, karyotyping and Y-chromosome microdeletion screening. Those with a history of cryptorchidism, vascular trauma, orchitis, obstruction of the vas deferens, abnormalities in chromosome number or microdeletions of the azoospermia factor region on the Y chromosome were excluded from the study [36]. All controls, which had a normal reproductive history and normal physical examination, had children within 1 year. After completing a questionnaire, each subject donated 5 ml of blood as a source of genomic DNA for further genotyping analysis.

### Semen analysis

Semen analysis for sperm concentration and motility was conducted on the basis of the World Health Organization (WHO) criteria [37]. To ensure the reliability of diagnosis, each subject was examined twice. The semen parameters (sperm concentration and sperm motility) were dichotomized based on WHO reference values. The controls consisted of proven fertile men with normal semen parameters. Considering the effect of mtDNA genetic variant on sperm concentration or sperm motility separately, we stratified the case group into two sub-groups respectively: concentration-group I (with sperm concentration  $\geq 15 \times 10^6/\text{ml}$ ) and concentration-group II (with sperm concentration  $< 15 \times 10^6/\text{ml}$ ), motility-group I (sperm motility  $\geq 40\%$  motile sperm) and motility-group II (sperm motility  $< 40\%$  motile sperm) [37].

### SNP selection and genotyping for validation

Through NGS of mtDNA genome, variations in each person were scored relative to the revised Cambridge reference sequence (rCRS) [38], and individual haplogroup was defined according to the reported East Asian mtDNA phylogenetic tree [39]. We selected SNPs from the same analysis as Lu et al 2015, meeting the following criteria for the validation: (i) SNPs had  $0.1 < P \leq 0.2$  in the comparison between 233 cases and 233 controls; (ii) SNPs were potentially functional.

Selected genetic variants were genotyped by a custom-by-design 48-Plex SNPscan™ Kit (Cat#:G0104; Genesky Biotechnologies Inc., Shanghai, China). This kit was developed according to patented SNP genotyping technology by Genesky Biotechnologies Inc., which was based on double ligation and multiplex fluorescence PCR [40]. In order to validate the genotyping accuracy using SNPscan™ Kit, five percent of the samples were randomly selected for repeat genotyping by single nucleotide extension using the Multiplex SNaPshot Kit (Applied Biosystems Inc., Foster City, CA, USA), and the concordance rates were more than 99%.

### Statistical analysis

Basic descriptive diversity statistics were calculated with DnaSP. The association analysis of stage one was

performed using PLINK (version 1.07; <http://pngu.mgh.harvard.edu/~purcell/plink/>). Statistical analysis of stage two was performed by Stata 10.0 (StataCorp LP, USA). Infertility risks were estimated with odds ratios (OR) and 95% confidence intervals (95% CI) using multivariate logistic regression. Two-sided tests were utilized and the Bonferroni adjustment for multiple testing was used. Thus if there are  $n$  tests in a particular set of interest and an investigation-wide 5% test is required, the applied  $P$  value for a truly significant result is calculated as  $0.05/n$ . Results will also be commented for which, although not attaining significance after applying the rather stringent Bonferroni adjustment, nevertheless returned a spot  $P$  value of  $< 5\%$ .

### Author contributions

CL and XW directed the study, obtained financial support and were responsible for study design. JJ, MX and ZH performed overall project management with, performed statistical analysis with LL and HZ drafted the initial manuscript. SY and SL were responsible for subject recruitment and sample preparation. LJ, YX and XL conceived of the study, and participated in its design and coordination and helped to draft the manuscript. All authors read and approved the final manuscript.

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### CONFLICTS OF INTEREST

The authors declare no conflicts of interest

### REFERENCES

1. Holden CA, McLachlan RI, Pitts M, Cumming R, Wittert G, Agius PA, Handelsman DJ, de Kretser DM. Men in Australia Telephone Survey (MATES): a national survey of the reproductive health and concerns of middle-aged and older Australian men. *Lancet*. 2005; 366:218-224.
2. Walsh TJ, Pera RR, Turek PJ. The genetics of male infertility. *Semin Reprod Med*. 2009; 27:124-136.
3. Ferlin A, Arredi B, Speltra E, Cazzadore C, Selice R, Garolla A, Lenzi A, Foresta C. Molecular and clinical characterization of Y chromosome microdeletions in infertile men: a 10-year experience in Italy. *J Clin Endocrinol Metab*. 2007; 92:762-770.
4. Hargreave TB. Genetic basis of male fertility. *Br Med Bull*. 2000; 56:650-671.
5. Bashamboo A, Ferraz-de-Souza B, Lourenco D, Lin L, Sebire NJ, Montjean D, Bignon-Topalovic J, Mandelbaum J, Siffroi JP, Christin-Maitre S, Radhakrishna U, Rouba H, Ravel C, et al. Human male infertility associated with mutations in NR5A1 encoding steroidogenic factor 1. *Am J Hum Genet*. 2010; 87:505-512.
6. O'Flynn O'Brien KL, Varghese AC, Agarwal A. The genetic causes of male factor infertility: a review. *Fertil Steril*. 2010; 93:1-12.
7. Selvi Rani D, Vanniarajan A, Gupta NJ, Chakravarty B, Singh L, Thangaraj K. A novel missense mutation C11994T in the mitochondrial ND4 gene as a cause of low sperm motility in the Indian subcontinent. *Fertil Steril*. 2006; 86:1783-1785.
8. Demain LA, Conway GS, Newman WG. Genetics of mitochondrial dysfunction and infertility. *Clin Genet*. 2017; 91:199-207.
9. Modica-Napolitano JS, Singh KK. Mitochondria as targets for detection and treatment of cancer. *Expert Rev Mol Med*. 2002; 4:1-19.
10. Modica-Napolitano JS, Singh KK. Mitochondrial dysfunction in cancer. *Mitochondrion*. 2004; 4:755-762.
11. Farha S, Hu B, Comhair S, Zein J, Dweik R, Erzurum SC, Aldred MA. Mitochondrial haplogroups and risk of pulmonary arterial hypertension. *PLoS One*. 2016; 11:e0156042.
12. Pyle A, Foltynie T, Tiangyou W, Lambert C, Keers SM, Allcock LM, Davison J, Lewis SJ, Perry RH, Barker R, Burn DJ, Chinnery PF. Mitochondrial DNA haplogroup cluster UKJT reduces the risk of PD. *Ann Neurol*. 2005; 57:564-567.
13. van der Walt JM, Dementieva YA, Martin ER, Scott WK, Nicodemus KK, Kroner CC, Welsh-Bohmer KA, Saunders AM, Roses AD, Small GW, Schmechel DE, Murali Doraiswamy P, Gilbert JR, et al. Analysis of European mitochondrial haplogroups with Alzheimer disease risk. *Neurosci Lett*. 2004; 365:28-32.
14. Bahrehmand Namaghi I, Vaziri H. Sperm mitochondrial DNA deletion in Iranian infertiles with asthenozoospermia. *Andrologia*. 2017; 49. doi: 10.1111/and.12627.
15. Lu C, Xu M, Wang R, Qin Y, Ren J, Wu W, Song L, Wang S, Zhou Z, Shen H, Sha J, Hu Z, Xia Y, et al. A genome-wide association study of mitochondrial DNA in Chinese men identifies two risk single nucleotide substitutions for idiopathic oligoasthenospermia. *Mitochondrion*. 2015; 24:87-92.
16. Liu SW, Li Y, Zou LL, Guan YT, Peng S, Zheng LX, Deng SM, Zhu LY, Wang LW, Chen LX. Chloride channels are involved in sperm motility and are downregulated in spermatozoa from patients with asthenozoospermia. *Asian J Androl*. 2017; 19:418-24. doi: 10.4103/1008-682X.181816.

17. Amaral S, Oliveira PJ, Ramalho-Santos J. Diabetes and the impairment of reproductive function: possible role of mitochondria and reactive oxygen species. *Curr Diabetes Rev.* 2008; 4:46-54.
18. Ramalho-Santos J, Varum S, Amaral S, Mota PC, Sousa AP, Amaral A. Mitochondrial functionality in reproduction: from gonads and gametes to embryos and embryonic stem cells. *Hum Reprod Update.* 2009; 15:553-572.
19. Baklouti-Gargouri S, Ghorbel M, Ben Mahmoud A, Mkaouar-Rebai E, Cherif M, Chakroun N, Sellami A, Fakhfakh F, Ammar-Keskes L. Identification of a novel m.9588G > a missense mutation in the mitochondrial COIII gene in asthenozoospermic Tunisian infertile men. *J Assist Reprod Genet.* 2014; 31:595-600.
20. Baklouti-Gargouri S, Ghorbel M, Ben Mahmoud A, Mkaouar-Rebai E, Cherif M, Chakroun N, Sellami A, Fakhfakh F, Ammar-Keskes L. A novel m.6307A>G mutation in the mitochondrial COXI gene in asthenozoospermic infertile men. *Mol Reprod Dev.* 2013; 80:581-587.
21. Feng GF, Zhang J, Feng LM, Shen NX, Li LJ, Zhu YM. Mitochondrial DNA haplogroup associated with sperm motility in the Han population. *Asian J Androl.* 2013; 15:630-633.
22. Mossman JA, Slate J, Birkhead TR. Mitochondrial haplotype does not affect sperm velocity in the zebra finch *Taeniopygia guttata*. *J Evol Biol.* 2010; 23:422-432.
23. Ruiz-Pesini E, Lapeña AC, Díez-Sánchez C, Pérez-Martos A, Montoya J, Alvarez E, Díaz M, Urriés A, Montoro L, López-Pérez MJ, Enriquez JA. Human mtDNA haplogroups associated with high or reduced spermatozoa motility. *Am J Hum Genet.* 2000; 67:682-696.
24. Xie S, Zhang J, Sun J, Zhang M, Zhao F, Wei QP, Tong Y, Liu X, Zhou X, Jiang P, Ji Y, Guan MX. Mitochondrial haplogroup D4j specific variant m.11696G > a (*MT-ND4*) may increase the penetrance and expressivity of the LHON-associated m.11778G > a mutation in Chinese pedigrees. *Mitochondrial DNA A DNA Mapp Seq Anal.* 2017; 28:434-41.
25. Lu J, Qian Y, Li Z, Yang A, Zhu Y, Li R, Yang L, Tang X, Chen B, Ding Y, Li Y, You J, Zheng J, et al. Mitochondrial haplotypes may modulate the phenotypic manifestation of the deafness-associated 12S rRNA 1555A>G mutation. *Mitochondrion.* 2010; 10:69-81.
26. Qu J, Li R, Zhou X, Tong Y, Yang L, Chen J, Zhao F, Lu C, Qian Y, Lu F, Guan MX. Cosegregation of the ND4 G11696A mutation with the LHON-associated ND4 G11778A mutation in a four generation Chinese family. *Mitochondrion.* 2007; 7:140-146.
27. Zhou X, Wei Q, Yang L, Tong Y, Zhao F, Lu C, Qian Y, Sun Y, Lu F, Qu J, Guan MX. Leber's hereditary optic neuropathy is associated with the mitochondrial ND4 G11696A mutation in five Chinese families. *Biochem Biophys Res Commun.* 2006; 340:69-75.
28. Beutner G, Alavian KN, Jonas EA, Porter GA Jr. The Mitochondrial Permeability Transition Pore and ATP Synthase. *Handb Exp Pharmacol.* 2016:1-26.
29. Mimaki M, Wang X, McKenzie M, Thorburn DR, Ryan MT. Understanding mitochondrial complex I assembly in health and disease. *Biochim Biophys Acta.* 2012; 1817:851-862.
30. Palmieri FM, Barton DL. Challenges of oral medications in patients with advanced breast cancer. *Semin Oncol Nurs.* 2007; 23:S17-S22.
31. Mitchell AL, Elson JL, Howell N, Taylor RW, Turnbull DM. Sequence variation in mitochondrial complex I genes: mutation or polymorphism? *J Med Genet.* 2006; 43:175-179.
32. Yano T, Magnitsky S, Ohnishi T. Characterization of the complex I-associated ubiquinone species: toward the understanding of their functional roles in the electron/proton transfer reaction. *Biochim Biophys Acta.* 2000; 1459:299-304.
33. Deschauer M, Bamberg C, Claus D, Zierz S, Turnbull DM, Taylor RW. Late-onset encephalopathy associated with a C11777A mutation of mitochondrial DNA. *Neurology.* 2003; 60:1357-1359.
34. Ellouze S, Augustin S, Bouaita A, Bonnet C, Simonutti M, Forster V, Picaud S, Sahel JA, Corral-Debrinski M. Optimized allotopic expression of the human mitochondrial ND4 prevents blindness in a rat model of mitochondrial dysfunction. *Am J Hum Genet.* 2008; 83:373-387.
35. Ji Y, Zhang AM, Jia X, Zhang YP, Xiao X, Li S, Guo X, Bandelt HJ, Zhang Q, Yao YG. Mitochondrial DNA haplogroups M7b1'2 and M8a affect clinical expression of leber hereditary optic neuropathy in Chinese families with the m.11778G>a mutation. *Am J Hum Genet.* 2008; 83:760-768.
36. Xia Y, Han Y, Wu B, Wang S, Gu A, Lu N, Bo J, Song L, Jin N, Wang X. The relation between urinary metabolite of pyrethroid insecticides and semen quality in humans. *Fertil Steril.* 2008; 89:1743-1750.
37. Cooper TG, Noonan E, von Eckardstein S, Auger J, Baker HW, Behre HM, Haugen TB, Kruger T, Wang C, Mbizvo MT, Vogelsong KM. World Health Organization reference values for human semen characteristics. *Hum Reprod Update.* 2010; 16:231-245.
38. Bandelt HJ, Kloss-Brandstatter A, Richards MB, Yao YG, Logan I. The case for the continuing use of the revised Cambridge Reference Sequence (rCRS) and the standardization of notation in human mitochondrial DNA studies. *J Hum Genet.* 2014; 59:66-77.
39. Kong QP, Bandelt HJ, Sun C, Yao YG, Salas A, Achilli A, Wang CY, Zhong L, Zhu CL, Wu SF, Torroni A, Zhang YP. Updating the East Asian mtDNA phylogeny: a prerequisite for the identification of pathogenic mutations. *Hum Mol Genet.* 2006; 15:2076-2086.
40. Chen X, Li S, Yang Y, Yang X, Liu Y, Hu W, Jin L, Wang X. Genome-wide association study validation identifies novel loci for atherosclerotic cardiovascular disease. *J Thromb Haemost.* 2012; 10:1508-1514.