

Updated meta-analysis of the role of *APOE* $\epsilon 2/\epsilon 3/\epsilon 4$ alleles in frontotemporal lobar degeneration

Wen-Hua Su^{1,2}, Zhi-Hong Shi^{1,2}, Shu-Ling Liu^{1,2}, Xiao-Dan Wang^{1,2}, Shuai Liu^{1,2} and Yong Ji^{1,2}

¹Department of Neurology, Tianjin Huanhu Hospital, Tianjin, China

²Tianjin Key Laboratory of Cerebral Vascular and Neurodegenerative Diseases, Tianjin Huanhu Hospital, Tianjin, China

Correspondence to: Yong Ji, **email:** jiyongtianjin@163.com

Keywords: *APOE*, FTL, allele, genotype, meta-analysis

Received: January 17, 2017

Accepted: April 11, 2017

Published: April 21, 2017

Copyright: Su et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License 3.0 (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

We performed an updated meta-analysis to assess the role of the $\epsilon 2/\epsilon 3/\epsilon 4$ alleles of Apolipoprotein E gene (*APOE*) in frontotemporal lobar degeneration (FTLD). The relevant articles were retrieved from PubMed, CENTRAL, EMBASE and Web of Science databases, and 51 eligible case-control studies with 5123 cases and 20566 controls were selected after screening according to inclusion and exclusion criteria. Our analysis demonstrated that *APOE* $\epsilon 4$ was associated with increased FTLD risk in all genetic models ($\epsilon 4$ vs. $\epsilon 3$ allele, $\epsilon 4$ vs. $\epsilon 2$ allele, $\epsilon 4$ vs. $\epsilon 2+\epsilon 3+\epsilon 4$ allele, $\epsilon 4$ vs. $\epsilon 2+\epsilon 3+\epsilon 4$ carrier, $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$, $\epsilon 3\epsilon 4$ vs. $\epsilon 3\epsilon 3$, $\epsilon 3\epsilon 4+\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$, $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3+\epsilon 3\epsilon 4$, all $P < 0.01$, odds ratio [OR] > 1). Subgroup analysis revealed significant association between *APOE* $\epsilon 4$ and FTLD ($P < 0.01$, OR > 1) for the Caucasian, Italian, population based (PB), $P > 0.05$ value of the Hardy-Weinberg Equilibrium (HWE), Newcastle-Ottawa scale score > 6 , and behavioral variant frontotemporal dementia (bvFTD) subgroups. However, there was no significant association between the *APOE* $\epsilon 2$ allele and FTLD ($P > 0.05$) in most genetic models and sub-group analyses. Begg's and Egger's tests also revealed no publication bias, and sensitivity analysis showed that our data analysis was robust. Thus our meta-analyses suggest that *APOE* $\epsilon 4$ is a genetic risk factor in patients with FTLD.

INTRODUCTION

Frontotemporal lobar degeneration (FTLD) is a common form of dementia that is characterized by focal atrophy of frontal and/or anterior temporal brain lobes [1]. The distinct clinical subtypes of FTLD include behavior variant frontotemporal dementia (bvFTD), semantic dementia (SD) and progressive non-fluent aphasia (PNFA) [2, 3]. Several genetic variants are associated with FTLD [4–6]. In the Italian population, C276T polymorphism of neuronal nitric oxide synthase (*nNOS*) gene is linked to increased susceptibility to sporadic FTLD [5]. Conversely, A2518G polymorphism in monocyte chemoattractant protein 1 (*MCP-1*) gene is a protective factor of sporadic FTLD [6].

Human Apolipoprotein E (*APOE*) gene that is located on chromosome 19 is involved in lipid homeostasis and is implicated in cardiovascular disease [7, 8]. Altered structure and function of ApoE protein is associated with neurodegenerative disorders such as Alzheimer's disease (AD) [8]. *APOE* gene has three common alleles ($\epsilon 2$, $\epsilon 3$

and $\epsilon 4$) and six related genotypes ($\epsilon 3\epsilon 3$, $\epsilon 3\epsilon 2$, $\epsilon 2\epsilon 2$, $\epsilon 3\epsilon 4$, $\epsilon 4\epsilon 4$, and $\epsilon 2\epsilon 4$) and distinct pathological roles have been attributed to all 3 alleles of *APOE*, namely, $\epsilon 2$, $\epsilon 3$, and $\epsilon 4$ [8]. The conclusions of various studies that have investigated the role of *APOE* polymorphism in FTLD have been inconsistent and contradictory. For instance, *APOE* $\epsilon 4$ was associated with increased FTLD risk in the Dutch population [9]. However, a negative association was reported between *APOE* polymorphism and FTLD risk in German patients [10]. In addition, genome wide association studies (GWAS) data of FTLD did not confirm a positive association with the *APOE* gene [11, 12].

So far, only two meta-analyses have reported on the relationship between *APOE* polymorphism and susceptibility to FTLD [13, 14]. Since many new studies have published on since 2013, we conducted an updated meta-analysis to reassess this association by systematically retrieving, screening and enrolling the available case-control studies to determine the association between *APOE* polymorphism and FTLD risk.

RESULTS

Selection criteria for eligible studies in the meta-analysis

Figure 1 shows the flow diagram of methodology used to search databases and select relevant studies based on “Preferred Reporting Items for Systematic Reviews and Meta-Analyses” (PRISMA). A total of 488 records were initially identified by searching four online databases, namely PubMed ($n = 74$), Cochrane Central Register of Controlled Trials (CENTRAL, $n = 0$), Excerpta Medica Database (EMBASE, $n = 290$) and Web of Science (WOS, $n = 124$). We removed 112 duplicate records after identifying them on Endnote. Further, 284 records that included case reports, posters, book articles, reviews, meeting abstracts ($n = 53$), non-FTLD, non-*ApoE*, non-clinical, non-mutation data ($n = 223$), and meta-analysis ($n = 8$) were also excluded. The remaining 92 full-text articles were then assessed for eligibility that resulted in excluding 41 articles for lack of relevant or control data. Finally, 51 case-control studies with 5123 cases and 20566 controls were included in our meta-analysis [5, 6, 9, 10, 13, 15–60]. The NOS assessment showed that three studies had a NOS score of 5 [39, 46, 47] and another three studies had a NOS score of 6 [26, 28, 32] indicating the medium-quality. The other 45 studies [5, 6, 9, 10, 13, 15–25, 27, 29–31, 33–38, 40–45, 48–60] were of high-quality with NOS scores > 6 . Supplementary Table 1 shows the characteristics of eligible studies.

APOE polymorphism and FTLN risk meta-analysis

The pooled values of OR and 95% confidence interval (CI) were analyzed by Mantel-Haenszel statistics to identify associations between *APOE* $\epsilon 2$, $\epsilon 3$, $\epsilon 4$ alleles and FTLN risks. As shown in Table 1, increased FTLN risk was observed in $\epsilon 4$ vs. $\epsilon 3$ allele model ($P < 0.001$, OR = 1.66, 95% CI = 1.35–2.03), $\epsilon 4$ vs. $\epsilon 2$ allele model ($P = 0.008$, OR = 1.52, 95% CI = 1.12–2.06), $\epsilon 4$ vs. $\epsilon 2+\epsilon 3+\epsilon 4$ allele model ($P < 0.001$, OR = 1.52, 95% CI = 1.31–1.76), $\epsilon 4$ vs. $\epsilon 2+\epsilon 3+\epsilon 4$ carrier model ($P < 0.001$, OR = 1.50, 95% CI = 1.32–1.70). Similarly, increased risk was observed for the genetic models of $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$ ($P < 0.001$, OR = 3.23, 95% CI = 2.27–4.60), $\epsilon 3\epsilon 4$ vs. $\epsilon 3\epsilon 3$ ($P < 0.001$, OR = 1.62, 95% CI = 1.25–2.10), $\epsilon 3\epsilon 4+\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$ ($P < 0.001$, OR = 1.70, 95% CI = 1.33–2.19), and $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3+\epsilon 3\epsilon 4$ ($P < 0.001$, OR = 2.82, 95% CI = 1.99–3.98) as shown in Table 1. These data demonstrated that the *APOE* $\epsilon 4$ allele increased FTLN susceptibility in a dose-dependent manner.

In contrast, *APOE* $\epsilon 2$ allele was not associated with FTLN risk. Our analyses for *APOE* $\epsilon 2$ showed significant difference only in the models of $\epsilon 2\epsilon 2$ vs. $\epsilon 3\epsilon 3$ ($P = 0.039$, OR = 1.74, 95% CI = 1.03–2.96) and $\epsilon 2\epsilon 2$ vs. $\epsilon 3\epsilon 3+\epsilon 3\epsilon 4$ ($P = 0.024$, OR = 1.84, 95% CI = 1.08–3.12), but not others (all $P > 0.05$). The forest plots for the allele models

of $\epsilon 4$ vs. $\epsilon 3$ and $\epsilon 2$ vs. $\epsilon 3$ are shown in Figures 2 and 3, respectively.

Subgroup analysis of *APOE* polymorphism and FTLN risk

Next, we performed a series of subgroup analyses based on ethnicity (Caucasian and Asian), country (Italy, China, USA and UK), source of control (PB and HB), clinical subtypes (bvFTD, SD, PNFA, FTLN MND-, FTLN MND+), HWE (P value of HWE > 0.05 and < 0.05), and NOS (score > 6 and ≤ 6). We observed that Caucasian, Italian, PB, P value of HWE > 0.05 , and NOS score > 6 subgroups for *APOE* $\epsilon 4$ demonstrated increased FTLN risk in the following models: $\epsilon 4$ vs. $\epsilon 3$ (Table 2, all $P < 0.01$, OR > 1); $\epsilon 4$ vs. $\epsilon 2$ (Table 2, all $P < 0.05$, OR > 1); $\epsilon 4$ vs. $\epsilon 2+\epsilon 3+\epsilon 4$ allele (Table 2, all $P < 0.001$, OR > 1); $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$ (Table 3, all $P < 0.001$, OR > 1); $\epsilon 3\epsilon 4$ vs. $\epsilon 3\epsilon 3$ (Table 3, all $P < 0.01$, OR > 1); $\epsilon 3\epsilon 4+\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$ (Table 4, all $P < 0.01$, OR > 1); and $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3+\epsilon 3\epsilon 4$ (Table 4, all $P < 0.01$, OR > 1). These data demonstrated that both $\epsilon 4\epsilon 4$ and $\epsilon 3\epsilon 4$ genotypes of *APOE* conferred increased susceptibility to FTLN in the Caucasian population, especially people of Italian origin.

Moreover, our analysis for *APOE* $\epsilon 4$ in Asian populations, especially Chinese individuals demonstrated enhanced FTLN risk for the allele (Table 2, $\epsilon 4$ vs. $\epsilon 3$, $P = 0.001$, OR = 2.04; $\epsilon 4$ vs. $\epsilon 2+\epsilon 3+\epsilon 4$, $P = 0.001$, OR = 1.94), heterozygote (Table 3, $\epsilon 3\epsilon 4$ vs. $\epsilon 3\epsilon 3$, $P = 0.001$, OR = 2.20), dominant (Table 4, $\epsilon 3\epsilon 4+\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$, $P = 0.001$, OR = 2.21) and carrier (Supplementary Table 2, $\epsilon 4$ vs. $\epsilon 2+\epsilon 3+\epsilon 4$ carrier, $P = 0.003$, OR = 1.92) models, but were not significant for homozygote (Table 3, $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$, $P = 0.068$) and recessive (Table 4, $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3+\epsilon 3\epsilon 4$, $P = 0.101$) models. These indicated that in the Asian population, including the Chinese individuals, the $\epsilon 3\epsilon 4$ genotype was linked to increased FTLN risk. The forest plots of subgroup analysis based on ethnicity for *APOE* $\epsilon 4$ under all genetic models were shown in Supplementary Figures 1–8.

In addition, stratified analysis of clinical subtypes (bvFTD, SD, PNFA, FTLN with or without motor neuron disease) showed that all genetic models were associated with increased bvFTD risk (Tables 2–4, Supplementary Table 2, all $P < 0.01$, OR > 1). This suggested that *APOE* $\epsilon 4$ was a risk factor for bvFTD.

In regard to *APOE* $\epsilon 2$, no significant differences were observed in the subgroup analyses for almost all genetic models (Supplementary Tables 2–5, $P > 0.05$). These findings further confirmed the negative genetic association between *APOE* $\epsilon 2$ and FTLN risks.

Heterogeneity, publication bias and sensitivity analysis

We assessed heterogeneity between studies by performing the Q statistic and I^2 tests. As shown in Table 1,

there was no heterogeneity among different studies for the following models: $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$, $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3+\epsilon 3\epsilon 4$, $\epsilon 2\epsilon 2$ vs. $\epsilon 3\epsilon 3$, $\epsilon 3\epsilon 2$ vs. $\epsilon 3\epsilon 3$, and $\epsilon 2\epsilon 2$ vs. $\epsilon 3\epsilon 3+\epsilon 3\epsilon 2$ (all P value of heterogeneity > 0.1 , $I^2 < 25\%$). Hence, we used the fixed-effect model for their analysis. The random-effect model was applied for others.

In addition, Begg's test and Egger's test analyses suggested absence of publication bias (Supplementary Table 6, all P value > 0.1). Begg's funnel plot of publication bias for $\epsilon 4$ vs. $\epsilon 3$ and $\epsilon 2$ vs. $\epsilon 3$ allele models are shown in Figure 4. Furthermore, sensitivity analysis was performed to evaluate the reliability of data and strengthen the validity of genetic relationship. We observed that similar pooled ORs were obtained when individual studies were omitted one by one, thereby indicating that the original statistical data were genuine and robust (Figure 5).

DISCUSSION

In 2002, Verpillat *et al.* [13] carried out a meta-analysis of 11 studies, and reported that *APOE* $\epsilon 2$ was associated with an increased risk of FTLD in the Caucasian population. However, in 2013, another meta-analysis based on 28 studies by Rubino *et al.* [14] in 2013 showed that FTLD susceptibility was associated with *APOE* $\epsilon 4$, but not $\epsilon 2$. These contradictory conclusions may have been a result of small and different sample sizes.

Recently, mutations in valosin-containing protein (*VCP*), progranulin (*GRN*), and the microtubule-associated protein tau (*MAPT*) genes were reported by us in 38 Chinese FTLD cases [61]. Further, our analysis of 62 Chinese FTLD patients and 381 sex- and age-matched elderly controls demonstrated significant association between FTLD susceptibility and *APOE* $\epsilon 4$, but not $\epsilon 2$ [36]. However, both conclusions were limited by small sample sizes. Therefore, to comprehensively assess the factors that are associated with FTLD, we enrolled 51 case-control studies and conducted an updated meta-analysis that also included subtype analyses of factors such as country, ethnicity, source of controls and clinical subtypes. Our data demonstrated a strong positive association between *APOE* $\epsilon 4$ and FTLD risks in the allele, homozygote, heterozygote, dominant recessive and carrier models. However, no statistically correlation was observed between *APOE* $\epsilon 2$ and FTLD risks, thereby confirming our previous finding [36] and partly agreeing with the results reported by Verpillat *et al.* [13].

FTLD and Alzheimer's disease (AD) are main contributors to dementia [62]. The molecular mechanisms underlying the role of *APOE* $\epsilon 4$ in the pathogenesis of FTLD and AD are unclear. *APOE* $\epsilon 4$ reduced the clearance of beta-amyloid ($A\beta$) that resulted in enhanced $A\beta$ deposition within the neurons in the AD mouse model [63, 64]. *APOE* $\epsilon 4$ was also associated with $A\beta$ deposition in the brain of a FTLD case [65]. Hence, the link between

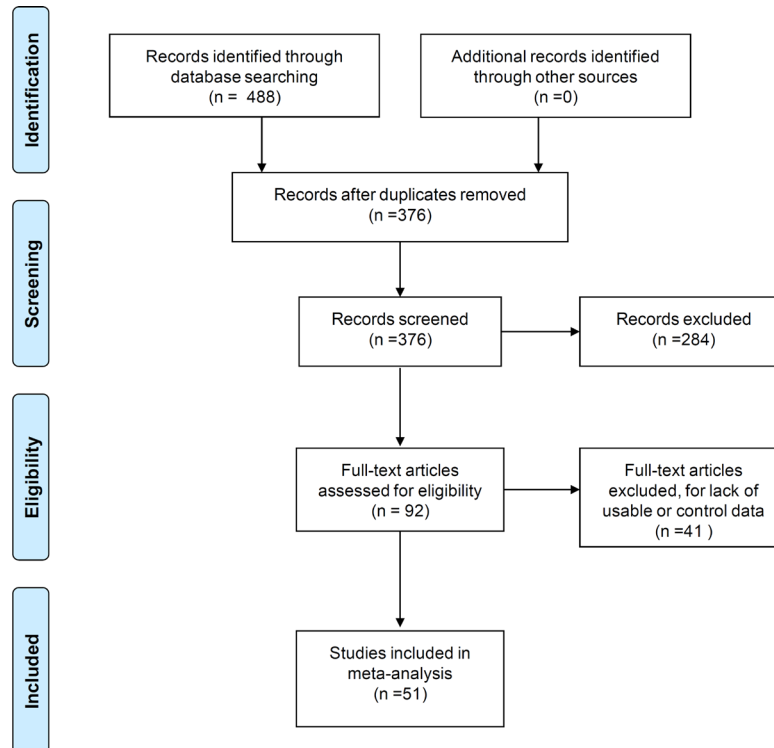


Figure 1: Flow diagram of database search and study selection.

Table 1: Meta-analysis for the association between *APOE* polymorphism and FTLTD risks

Comparison	Study number	Sample size (case/control)	Association Test		Heterogeneity		Model
			OR (95% CI)	<i>P</i>	<i>I</i> ²	<i>P</i>	
ε4 vs. ε3 allele	34	2072/13661	1.66 (1.35–2.03)	< 0.001	68.7%	< 0.001	Random
ε4 vs. ε2 allele	34	2072/13661	1.52 (1.12–2.06)	0.008	60.8%	< 0.001	Random
ε4 vs. ε2+ε3+ε4 allele	40	2417/15059	1.52 (1.31–1.76)	< 0.001	51.3%	< 0.001	Random
ε4 vs. ε2+ε3+ε4 carrier	47	3511/18046	1.50 (1.32–1.70)	< 0.001	40.9%	0.002	Random
ε4ε4 vs. ε3ε3	30	1650/11634	3.23 (2.27–4.60)	< 0.001	0.0%	0.922	Fixed
ε3ε4 vs. ε3ε3	32	1696/11700	1.62 (1.25–2.10)	< 0.001	67.3%	< 0.001	Random
ε3ε4+ε4ε4 vs. ε3ε3	32	1696/11700	1.70 (1.33–2.19)	< 0.001	67.6%	< 0.001	Random
ε4ε4 vs. ε3ε3+ε3ε4	30	1650/11634	2.82 (1.99–3.98)	< 0.001	0.0%	0.962	Fixed
ε2 vs. ε3 allele	34	2072/13661	1.09 (0.87–1.37)	0.462	51.5%	< 0.001	Random
ε2 vs. ε2+ε3+ε4 allele	34	2072/13661	1.01 (0.82–1.24)	0.953	43.1%	0.005	Random
ε2 vs. ε2+ε3+ε4 carrier	32	1936/13591	0.93 (0.74–1.17)	0.545	42.3%	0.007	Random
ε2ε2 vs. ε3ε3	22	944/9708	1.74 (1.03–2.96)	0.039	0.0%	0.774	Fixed
ε3ε2 vs. ε3ε3	32	1346/10740	0.87 (0.73–1.04)	0.132	24.2%	0.110	Fixed
ε3ε2+ε2ε2 vs. ε3ε3	32	1346/10740	0.95 (0.72–1.23)	0.678	41.6%	0.008	Random
ε2ε2 vs. ε3ε3+ε3ε2	22	944/9708	1.84 (1.08–3.12)	0.024	0.0%	0.842	Fixed

P < 0.05 of association test is shown in bold.

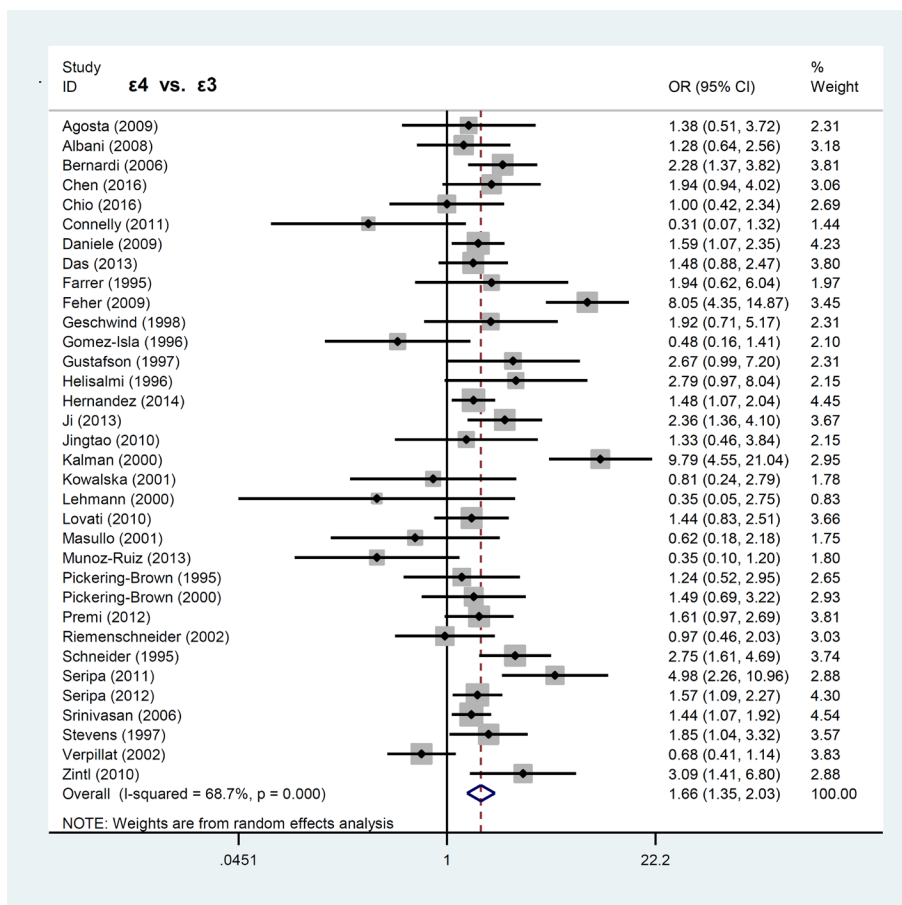


Figure 2: Forest plot of meta-analysis of the ε4 vs. ε3 allele model.

APOE $\epsilon 4$ and $A\beta$ deposition merits further investigation. In addition, *APOE* $\epsilon 4$ enhanced phosphorylation of tau protein in brains of transgenic mice [66]. Since FTLT-tau is a neuropathological subtype of FTLT [4, 67], abnormal Tau phosphorylation may be partly involved in the pathogenesis of FTLT by *APOE* $\epsilon 4$.

There are several limitations in this meta-analysis that need to be highlighted. Firstly, out of 51 case-control studies included in our pooled analysis, 19 studies [5, 6, 17, 18, 20, 21, 27, 35, 43, 44, 46, 47, 49, 50, 54, 55, 57–59] contained only allele or carrier data and did not provide information regarding the specific genotype frequencies of $\epsilon 3\epsilon 4$ and $\epsilon 3\epsilon 2$ that could have weakened the statistical output. Secondly, genetic heterogeneity existed between studies for majority of comparisons because of hospital based controls, lack of the pathology or autopsy confirmed FTLT diagnoses, clinical complexity, and pathological heterogeneity. Although poor quality studies were excluded based on NOS analysis, six medium quality articles [26, 28, 32, 39, 46, 47] were still included in the analysis. Hence, more high quality studies with large sample sizes are required to avoid false positives. Thirdly, our meta-analysis included only five articles based on Asian populations [22, 26, 36, 37, 39] compared to 46

articles based on Caucasian populations [5, 6, 9, 10, 13, 15–21, 23–25, 27–35, 38, 40–60]. Among these were 15 articles based on Italian populations [5, 6, 15–17, 19, 20, 23, 25, 41, 42, 48, 52, 53, 59]. In addition, only full-text articles in Chinese or English were collected for this meta-analysis. All these factors might lead to selection bias. Fourthly, bvFTL, the most frequent clinical subtype of FTLT is a clinical syndrome characterized by progressive changes of personality, abnormalities of social behavior and cognitive function, and lack of emotional response [4, 68]. Our subgroup analysis of bvFTL contained seven articles [6, 15, 21, 22, 53, 54, 56] that showed significant association with *APOE* $\epsilon 4$. It is probable that *APOE* $\epsilon 4$ may serve as a disease modifier of bvFTL. However, this result needs to be verified since our analysis was based on a small sample size. Similarly, only four articles for PNFA [6, 21, 48, 54] and five articles for SD [6, 21, 22, 48, 56] were available and therefore the role of *APOE* polymorphisms in PNFA and SD could not be determined conclusively. This was true of the subgroup analysis of FTLT with or without MND. Finally, in view of the unclear etiology of FTLT, more factors, including age at onset, male/female, pathological criteria, clinical presentation, living habits, the combination of *APOE* and

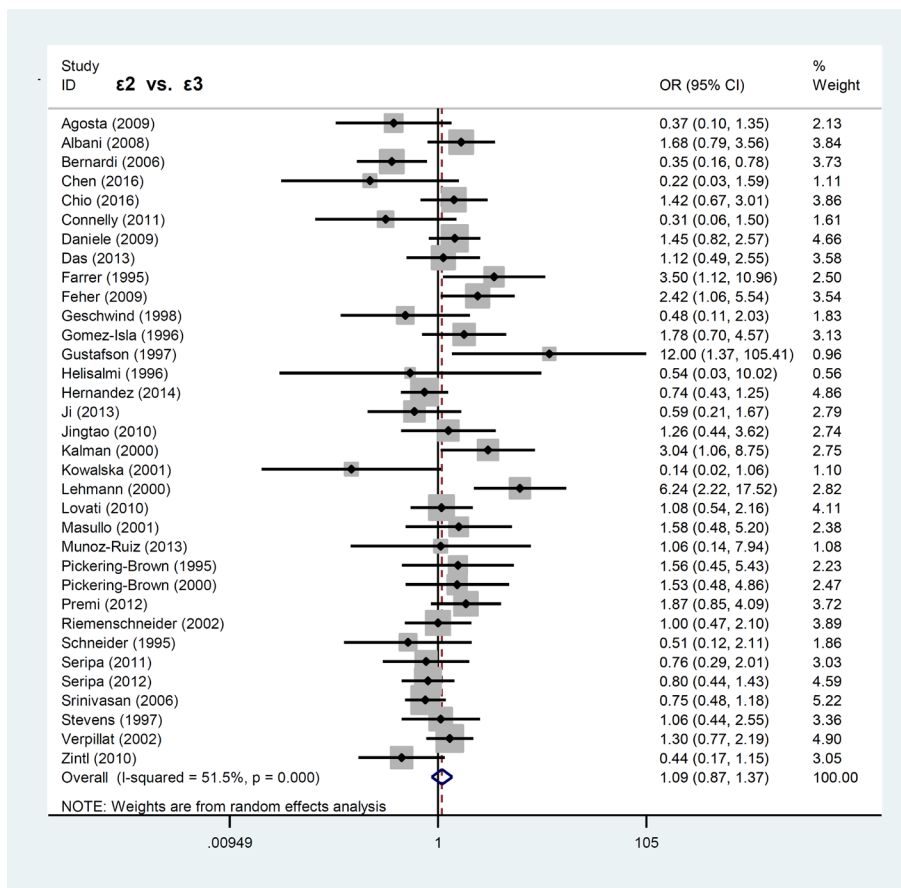


Figure 3: Forest plot of meta-analysis of the $\epsilon 2$ vs. $\epsilon 3$ allele model.

Table 2: Subgroup analysis of association between *APOE* $\epsilon 4$ and FTLN risks for $\epsilon 4$ vs. $\epsilon 3$, $\epsilon 4$ vs. $\epsilon 2$, and $\epsilon 4$ vs. $\epsilon 2+\epsilon 3+\epsilon 4$ allele models

Subgroup	$\epsilon 4$ vs. $\epsilon 3$				$\epsilon 4$ vs. $\epsilon 2$				$\epsilon 4$ vs. $\epsilon 2+\epsilon 3+\epsilon 4$			
	Study number	Sample size (case/control)	OR (95 % CI)	P	Study number	Sample size (case/control)	OR (95 % CI)	P	Study number	Sample size (case/control)	OR (95 % CI)	P
Ethnicity												
Caucasian	29	1854/11162	1.66 (1.31–2.09)	< 0.001	29	1854/11162	1.41 (1.01–1.97)	0.043	35	2199/12560	1.50 (1.28–1.77)	< 0.001
Asian	5	218/2499	1.72 (1.26–2.34)	0.001	5	218/2499	2.40 (1.12–5.11)	0.024	5	218/2499	1.65 (1.22–2.24)	0.001
Country												
Italy	10	839/2168	1.64 (1.30–2.07)	< 0.001	10	839/2168	1.57(0.93–2.65)	0.091	11	848/2193	1.55 (1.26–1.90)	< 0.001
China	3	113/2030	2.04 (1.36–3.07)	0.001	3	13/2030	2.99 (0.97–9.21)	0.056	3	113/2030	1.94 (1.30–2.90)	0.001
USA	4	106/3394	1.60 (0.75–3.40)	0.224	4	106/3394	1.29 (0.29–5.08)	0.733	5	169/3732	1.62 (1.06–2.49)	0.026
UK	4	345/962	1.39 (1.08–1.80)	0.012	4	345/962	0.74 (0.23–2.38)	0.609	4	345/962	1.32 (1.03–1.70)	0.028
Source of control												
PB	31	1912/13391	1.70 (1.36–2.11)	< 0.001	31	1912/13391	1.53 (1.11–2.12)	0.009	37	2257/14789	1.54 (1.32–1.80)	< 0.001
HB	3	160/270	1.25 (0.74–2.11)	0.400	3	160/270	1.19 (0.47–3.03)	0.715	3	160/270	1.20 (1.19–1.86)	0.488
Clinical subtypes												
bvFTD	4	373/2257	1.57 (1.246–1.99)	< 0.001	4	373/2257	2.14 (1.39–3.30)	0.001	5	400/2595	1.49 (1.19–1.86)	< 0.001
SD	2	59/956	1.09 (0.63–1.90)	0.755	2	59/956	1.31 (0.49–3.47)	0.587	2	59/956	1.09 (0.63–1.89)	0.747
PNFA	1	60/200	1.80 (1.02–3.15)	0.041	1	60/200	0.79 (0.30–2.04)	0.620	2	78/538	1.50 (0.91–2.48)	0.116
FTLD MND–	2	50/149	0.68 (0.29–1.59)	0.373	2	50/149	0.36 (0.11–1.17)	0.090	3	123/477	0.81 (0.53–1.23)	0.324
FTLD MND+	3	45/905	1.56 (0.90–2.71)	0.112	2	42/791	2.45 (0.79–7.57)	0.121	4	116/1233	1.30 (0.93–1.83)	0.125
NOS												
score > 6	28	1800/11889	1.67 (1.32–2.12)	< 0.001	28	1800/11889	1.64 (1.17–2.30)	0.004	34	2145/13287	1.54 (1.30–1.82)	< 0.001
score <= 6	6	272/1772	1.51 (1.09–2.10)	0.014	6	272/1772	0.98 (0.55–1.72)	0.937	6	272/1772	1.36 (0.99–1.88)	0.059

PB: population-based; HB: hospital-based; bvFTD: behavior variant frontotemporal dementia; SD: semantic dementia; PNFA: progressive non-fluent aphasia; FTLD: Frontotemporal lobar degeneration; MND: motor neuron disease; NOS: Newcastle-Ottawa scale; $P < 0.05$ is shown in bold.

Table 3: Subgroup analysis of association between *APOE* $\epsilon 3/\epsilon 4$ genotype frequency and FTLN risks for $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$ and $\epsilon 3\epsilon 4$ vs. $\epsilon 3\epsilon 3$ models

Subgroup	$\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$				$\epsilon 3\epsilon 4$ vs. $\epsilon 3\epsilon 3$			
	Study number	Sample size (case/control)	OR (95 % CI)	P	Study number	Sample size (case/control)	OR (95 % CI)	P
Ethnicity								
Caucasian	25	1447/9494	3.34 (2.31–4.83)	< 0.001	27	1493/9560	1.61 (1.19–2.16)	0.002
Asian	5	203/2140	2.20 (0.66–7.36)	0.199	5	203/2140	1.84 (1.29–2.63)	0.001
Country								
Italy	9	710/1850	3.71 (1.83–7.51)	< 0.001	10	738/1893	1.61 (1.28–2.03)	< 0.001
China	3	105/1735	4.36 (0.90–21.21)	0.068	3	105/1735	2.20 (1.37–3.51)	0.001
USA	4	91/2905	1.67 (0.42–6.64)	0.464	4	91/2905	1.58 (0.37–6.74)	0.535
UK	2	179/750	3.75 (1.65–8.54)	0.002	2	179/750	1.16 (0.66–2.02)	0.606
Source of control								
PB	29	1627/11474	3.28 (2.30–4.67)	< 0.001	31	1673/11540	1.65 (1.27–2.15)	< 0.001
HB	1	23/160	1.27 (0.06–27.36)	0.879	1	23/160	0.59 (0.13–2.69)	0.494
HWE								
$P > 0.05$	25	1481/10080	2.92 (1.99–4.30)	< 0.001	27	1527/10146	1.55 (1.20–2.01)	0.001
$P < 0.05$	5	169/1554	5.58 (2.31–13.47)	< 0.001	5	169/1554	1.95 (0.51–7.53)	0.332
Clinical subtypes								
bvFTD	3	310/1859	4.42 (1.93–10.09)	< 0.001	4	338/1902	1.48 (1.11–1.98)	0.008
SD	2	53/816	3.39 (0.82–13.91)	0.091	2	53/816	0.94 (0.46–1.92)	0.866
PNFA	1	56/185	1.28 (0.05–32.17)	0.879	1	56/185	1.85 (0.96–3.58)	0.066
FTLD MND–	1	19/103	1.61 (0.06–41.17)	0.774	1	19/103	0.55 (0.12–2.59)	0.449
FTLD MND+	2	30/734	3.04 (0.53–17.44)	0.212	2	30/734	1.50 (0.69–3.29)	0.306
NOS								
score > 6	26	1529/10171	3.32 (2.28–4.82)	< 0.001	28	1575/10237	1.67 (1.27–2.21)	< 0.001
Score <= 6	4	121/1463	2.58 (0.88–7.59)	0.084	4	121/1463	1.38 (0.82–2.32)	0.222

PB: population-based; HB: hospital-based; HWE: Hardy-Weinberg Equilibrium; bvFTD: behavior variant frontotemporal dementia; SD: semantic dementia; PNFA: progressive non-fluent aphasia; FTLD: Frontotemporal lobar degeneration; MND: motor neuron disease; NOS: Newcastle-Ottawa scale; $P < 0.05$ is shown in bold.

Table 4: Subgroup analysis of association between *APOE* $\epsilon 3/\epsilon 4$ genotype frequency and FTLD risks for $\epsilon 3\epsilon 4+\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$ and $\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3+\epsilon 3\epsilon 4$ models

Subgroup	Study number	$\epsilon 3\epsilon 4+\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$			$\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3+\epsilon 3\epsilon 4$			
		Sample size (case/control)	OR (95% CI)	P	Study number	Sample size (case/control)	OR (95% CI)	P
Ethnicity								
Caucasian	27	1493/9560	1.71 (1.28–2.27)	< 0.001	25	1447/9494	2.90 (2.02–4.17)	< 0.001
Asian	5	203/2140	1.82 (1.26–2.63)	0.001	5	203/2140	2.02 (0.61–6.72)	0.252
Country								
Italy	10	738/1893	1.67 (1.30–2.16)	< 0.001	9	710/1850	3.31 (1.63–6.72)	0.001
China	3	105/1735	2.21 (1.40–3.51)	0.001	3	105/1735	3.74 (0.77–18.13)	0.101
USA	4	91/2905	1.57 (0.43–5.77)	0.498	4	91/2905	1.21 (0.32–4.57)	0.774
UK	2	179/750	1.14 (0.40–3.27)	0.808	2	179/750	3.57 (1.58–8.08)	0.002
Source of control								
PB	31	1673/11540	1.74 (1.35–2.24)	< 0.001	29	1627/11474	2.85 (2.01–4.04)	< 0.001
HB	1	23/160	0.54 (0.12–2.45)	0.425	1	23/160	1.35 (0.06–28.97)	0.848
HWE								
$P > 0.05$	27	1527/10146	1.60 (1.24–2.06)	< 0.001	25	1481/10080	2.59 (1.77–3.79)	< 0.001
$P < 0.05$	5	169/1554	2.57 (0.88–7.51)	0.085	5	169/1554	4.38 (1.88–10.20)	0.001
Clinical subtypes								
bvFTD	4	338/1902	1.62 (1.22–2.14)	0.001	3	310/1859	3.96 (1.76–8.94)	0.001
SD	2	53/816	1.03 (0.54–1.95)	0.935	2	53/816	3.60 (0.88–14.71)	0.074
PNFA	1	56/185	1.80 (0.94–3.47)	0.077	1	56/185	1.09 (0.04–27.09)	0.959
FTLD MND–	1	19/103	0.52 (0.11–2.44)	0.408	1	19/103	1.75 (0.07–44.61)	0.734
FTLD MND+	2	30/734	1.54 (0.72–3.31)	0.263	2	30/734	2.71 (0.48–15.17)	0.257
NOS								
score > 6	28	1575/10237	1.75 (1.34–2.29)	< 0.001	26	1529/10171	2.86 (1.98–4.12)	< 0.001
Score ≤ 6	4	121/1463	1.43 (0.83–2.46)	0.203	4	121/1463	2.51 (0.85–7.40)	0.095

PB: population-based; HB: hospital-based; HWE: Hardy-Weinberg Equilibrium; bvFTD: behavior variant frontotemporal dementia; SD: semantic dementia; PNFA: progressive non-fluent aphasia; FTLD: Frontotemporal lobar degeneration; MND: motor neuron disease; NOS: Newcastle-Ottawa scale; $P < 0.05$ is shown in bold.

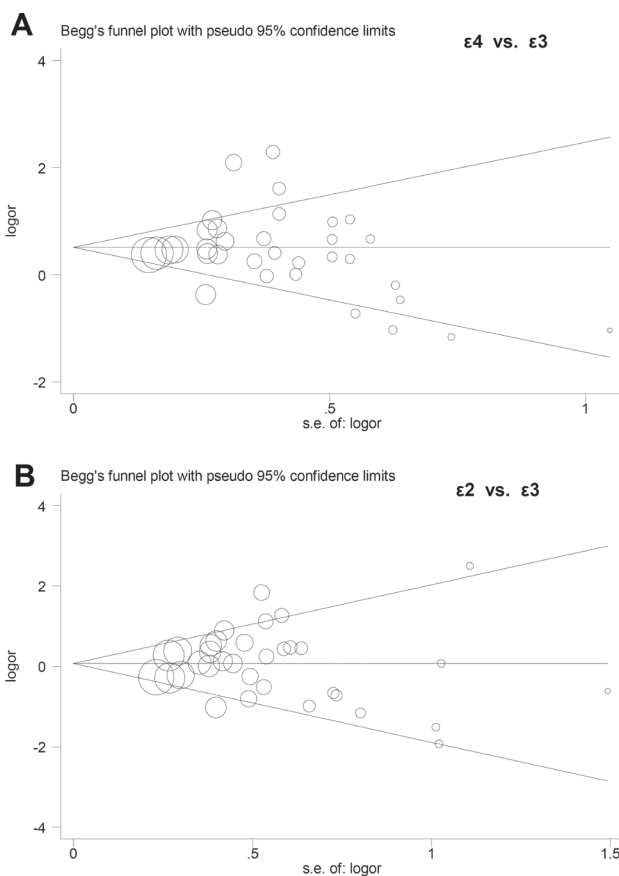


Figure 4: Begg's funnel plots of publication bias. (A) $\epsilon 4$ vs. $\epsilon 3$ allele model; (B) $\epsilon 2$ vs. $\epsilon 3$ allele model.

other related genes (e.g. *VCP*, *GRN*, *MAPT*) should be considered in future meta-analysis. Also, pathogenesis of *APOE* $\epsilon 4$ in the memory function, behavioral symptoms and brain morphological changes in FTLN-spectrum disease should be investigated.

In conclusion, this meta-analysis demonstrated that *APOE* $\epsilon 4$ was a genetic risk factor for FTLN patients in Caucasian and Asian populations, thereby corroborating the role of *APOE* genetic variants in FTLN. Also, our study demonstrated that *APOE* $\epsilon 2$ was not a susceptibility factor for FTLN.

MATERIALS AND METHODS

Database search and study selection

We searched four databases, including PubMed, CENTRAL, EMBASE and WOS until February 27th, 2017 with specific search terms listed in Supplementary

Table 7 and identified 488 records. After removing the duplicates by endnote software (Thomson Reuters), the remaining 376 records were screened according to our inclusion/exclusion criteria. We excluded the records of case reports, posters, books, reviews, meeting abstracts, meta-analysis, and the articles with non-FTLN, non-*ApoE*, non-clinical, non-mutation data. The remaining 92 full-text articles were then assessed to identify 51 eligible case-control studies while removing articles that lacked control or other usable data for this meta-analysis. The PRISMA was used in this study [69]. The PRISMA 2009 checklist is shown in Supplementary Table 8.

Quality assessment of eligible studies and data extraction

Three authors independently assessed the methodological quality of the selected case-control studies using the Newcastle-Ottawa Scale (NOS) (<http://www.>

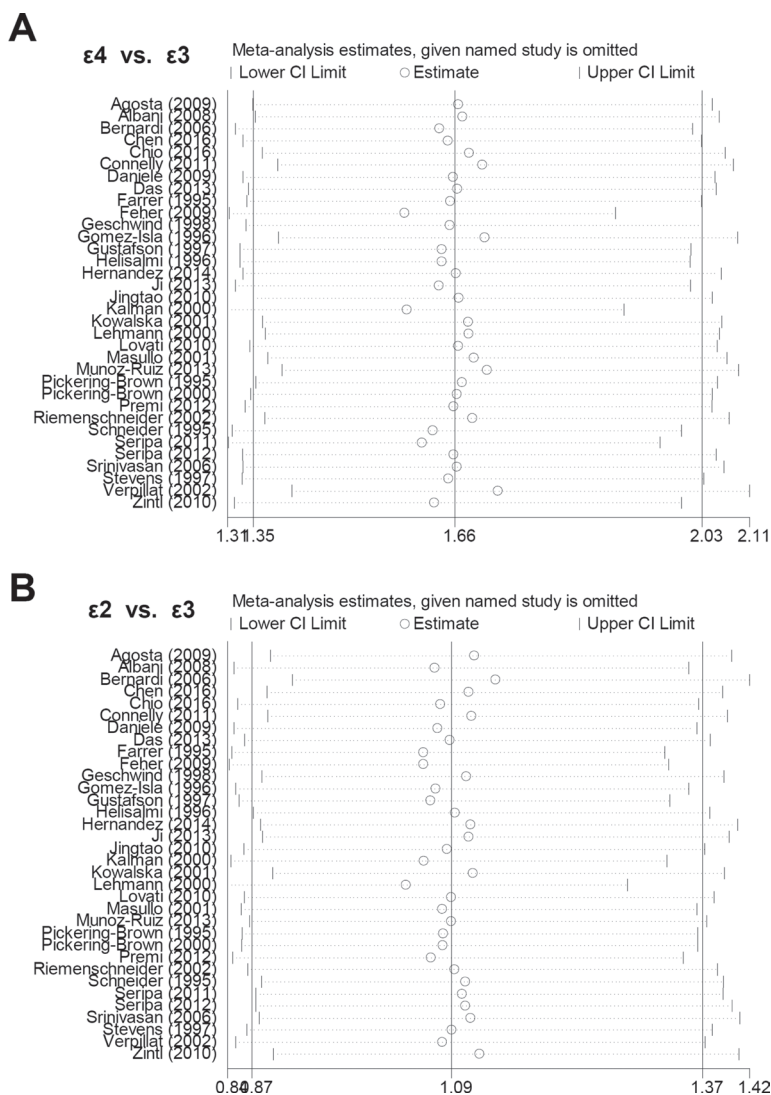


Figure 5: Sensitivity analyses. (A) $\epsilon 4$ vs. $\epsilon 3$ allele model; (B) $\epsilon 2$ vs. $\epsilon 3$ allele model.

ohri.ca/programs/clinical_epidemiology/oxford.asp) and extracted the relevant data. Studies with a NOS score > 6 were considered high quality, whereas studies with NOS score < 5 were considered poor and removed from the included studies. Whenever there was a disagreement, it was resolved by discussion among the three authors. The following information was collected from all the selected studies and summarized: first author, year of publication, country, ethnicity, genotype distributions ($\epsilon 3\epsilon 3$, $\epsilon 3\epsilon 2$, $\epsilon 2\epsilon 2$, $\epsilon 3\epsilon 4$, $\epsilon 4\epsilon 4$, and $\epsilon 2\epsilon 4$) in case group and control group, clinical subtypes of case, source of control, and genotyping assay. The first or the corresponding author was contacted by email whenever relevant data was not available.

Statistical analyses

Stata/SE 12.0 software (StataCorp, USA) was used for Mantel-Haenszel statistic, Q statistic and I^2 tests from P values, pooled ORs, and 95% CIs. $P < 0.05$ was considered statistically significant. Six genetic models, namely allele ($\epsilon 4$ vs. $\epsilon 3$; $\epsilon 2$ vs. $\epsilon 3$; $\epsilon 4$ vs. $\epsilon 2$; $\epsilon 4$ vs. $\epsilon 2 + \epsilon 3 + \epsilon 4$, $\epsilon 2$ vs. $\epsilon 2 + \epsilon 3 + \epsilon 4$), homozygote ($\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$, $\epsilon 2\epsilon 2$ vs. $\epsilon 3\epsilon 3$), heterozygote ($\epsilon 3\epsilon 4$ vs. $\epsilon 3\epsilon 3$, $\epsilon 3\epsilon 2$ vs. $\epsilon 3\epsilon 3$), dominant ($\epsilon 3\epsilon 4 + \epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3$, $\epsilon 3\epsilon 2 + \epsilon 2\epsilon 2$ vs. $\epsilon 3\epsilon 3$), recessive ($\epsilon 4\epsilon 4$ vs. $\epsilon 3\epsilon 3 + \epsilon 3\epsilon 4$, $\epsilon 2\epsilon 2$ vs. $\epsilon 3\epsilon 3 + \epsilon 3\epsilon 2$) or carrier ($\epsilon 4$ vs. $\epsilon 2 + \epsilon 3 + \epsilon 4$ carrier; $\epsilon 2$ vs. $\epsilon 2 + \epsilon 3 + \epsilon 4$ carrier) were used and Hardy-Weinberg Equilibrium (HWE) was calculated by chi-squared test. P values of Q statistic > 0.1 or I^2 values $\leq 25\%$ indicated heterogeneity between studies and the fixed-effect model was used for analysis. If not, the random-effect model was used. Subgroup analyses were performed based on ethnicity, country, source of control, clinical subtypes, HWE, and NOS score. Furthermore, Begg's funnel plot (Begg's test) and Egger's publication bias plot (Egger's test) was used to evaluate the potential publication bias. The P value of Begg's test and Egger's test > 0.05 was regarded as the absence of publication bias. Sensitivity analysis was also performed to evaluate the stability of statistical results.

ACKNOWLEDGMENTS AND FUNDING

This study was supported by National Natural Science Foundation of China (Grant Number: 81571057 and 81300947), Tianjin Science and Technology Support Programs (Grant Numbers: 13ZCZDSY01600), Tianjin Natural Science Foundation (Grant Number: 13JCYBJC21300), Key Research Project of Tianjin Public Health (Grant Number: 14KG117), the Science and Technology Project of Tianjin Municipal Health Bureau (Grant number: 2014KR10).

CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

REFERENCES

1. Pan XD, Chen XC. Clinic, neuropathology and molecular genetics of frontotemporal dementia: a mini-review. *Transl Neurodegener.* 2013; 2:8.
2. Tsai RM, Boxer AL. Therapy and clinical trials in frontotemporal dementia: past, present, and future. *J Neurochem.* 2016; 138:211–221.
3. Oeckl P, Steinacker P, Feneberg E, Otto M. Neurochemical biomarkers in the diagnosis of frontotemporal lobar degeneration: an update. *J Neurochem.* 2016; 138:184–192.
4. Li YQ, Tan MS, Yu JT, Tan L. Frontotemporal Lobar Degeneration: Mechanisms and Therapeutic Strategies. *Mol Neurobiol.* 2016; 53:6091–6105.
5. Venturelli E, Villa C, Scarpini E, Fenoglio C, Guidi I, Lovati C, Marcone A, Cortini F, Scalabrini D, Clerici F, Bresolin N, Mariani C, Cappa S, et al. Neuronal nitric oxide synthase C276T polymorphism increases the risk for frontotemporal lobar degeneration. *Eur J Neurol.* 2008; 15:77–81.
6. Galimberti D, Venturelli E, Villa C, Fenoglio C, Clerici F, Marcone A, Benussi L, Cortini F, Scalabrini D, Perini L, Restelli I, Binetti G, Cappa S, et al. MCP-1 A-2518G polymorphism: effect on susceptibility for frontotemporal lobar degeneration and on cerebrospinal fluid MCP-1 levels. *J Alzheimers Dis.* 2009; 17:125–133.
7. Lin-Lee YC, Kao FT, Cheung P, Chan L. Apolipoprotein E gene mapping and expression: localization of the structural gene to human chromosome 19 and expression of ApoE mRNA in lipoprotein- and non-lipoprotein-producing tissues. *Biochemistry.* 1985; 24:3751–3756.
8. Mahley RW. Apolipoprotein E: from cardiovascular disease to neurodegenerative disorders. *J Mol Med (Berl).* 2016; 94:739–746.
9. Stevens M, van Duijn CM, de Knijff P, van Broeckhoven C, Heutink P, Oostra BA, Niermeijer MF, van Swieten JC. Apolipoprotein E gene and sporadic frontal lobe dementia. *Neurology.* 1997; 48:1526–1529.
10. Riemenschneider M, Diehl J, Müller U, Förstl H, Kurz A. Apolipoprotein E polymorphism in German patients with frontotemporal degeneration. *J Neurol Neurosurg Psychiatry.* 2002; 72:639–641.
11. Ferrari R, Hernandez DG, Nalls MA, Rohrer JD, Ramasamy A, Kwok JB, Dobson-Stone C, Brooks WS, Schofield PR, Halliday GM, Hodges JR, Piguet O, Bartley L, et al. Frontotemporal dementia and its subtypes: a genome-wide association study. *Lancet Neurol.* 2014; 13:686–699.
12. Girard SL, Rouleau GA. Genome-wide association study in FTD: Divide to conquer. *Lancet Neurol.* 2014; 13:643–644.
13. Verpillat P, Camuzat A, Hannequin D, Thomas-Anterion C, Puel M, Belliard S, Dubois B, Didic M, Lacomblez L, Moreaud O, Golfier V, Campion D, Brice A, et al. Apolipoprotein E gene in frontotemporal dementia: an

- association study and meta-analysis. *Eur J Hum Genet.* 2002; 10:399–405.
14. Rubino E, Vacca A, Govone F, De Martino P, Pinessi L, Rainero I. Apolipoprotein E polymorphisms in frontotemporal lobar degeneration: a meta-analysis. *Alzheimers Dement.* 2013; 9:706–713.
 15. Agosta F, Vessel KA, Miller BL, Migliaccio R, Bonasera SJ, Filippi M, Boxer AL, Karydas A, Possin KL, Gorno-Tempini ML. Apolipoprotein E epsilon 4 is associated with disease-specific effects on brain atrophy in Alzheimer's disease and frontotemporal dementia. *Proc Natl Acad Sci USA.* 2009; 106:2018–2022.
 16. Albani D, Prato F, Fenoglio C, Batelli S, Dusi S, De Mauro S, Polito L, Lovati C, Galimberti D, Mariani C, Scarpini E, Forloni G. Association study to evaluate the serotonin transporter and apolipoprotein E genes in frontotemporal lobar degeneration in Italy. *J Hum Genet.* 2008; 53: 1029–1033.
 17. Bagnoli S, Piaceri I, Tedde A, Bessi V, Bracco L, Sorbi S, Nacmias B. TOMM40 polymorphisms in Italian Alzheimer's disease and frontotemporal dementia patients. *Neurol Sci.* 2013; 34:995–998.
 18. Balasa M, Sanchez-Valle R, Antonell A, Bosch B, Olives J, Rami L, Castellvi M, Molinuevo JL, Llado A. Usefulness of biomarkers in the diagnosis and prognosis of early-onset cognitive impairment. *J Alzheimers Dis.* 2014; 40:919–927.
 19. Bernardi L, Maletta RG, Tomaino C, Smirne N, Di Natale M, Perri M, Longo T, Colao R, Curcio SAM, Puccio G, Mirabelli M, Kawarai T, Rogava E, et al. The effects of APOE and tau gene variability on risk of frontotemporal dementia. *Neurobiol Aging.* 2006; 27:702–709.
 20. Boccardi M, Laakso MP, Bresciani L, Galluzzi S, Geroldi C, Beltramello A, Soininen H, Frisoni GB. The MRI pattern of frontal and temporal brain atrophy in fronto-temporal dementia. *Neurobiol Aging.* 2003; 24:95–103.
 21. Borroni B, Yancopoulou D, Tsutsui M, Padovani A, Sawcer SJ, Hodges JR, Spillantini MG. Association between tau H2 haplotype and age at onset in frontotemporal dementia. *Arch Neurol.* 2005; 62:1419–1422.
 22. Chen KL, Sun YM, Zhou Y, Zhao QH, Ding D, Guo QH. Associations between APOE polymorphisms and seven diseases with cognitive impairment including Alzheimer's disease, frontotemporal dementia, and dementia with Lewy bodies in southeast China. *Psychiatr Genet.* 2016; 26:124–131.
 23. Chio A, Brunetti M, Barberis M, Iazzolino B, Montuschi A, Ilardi A, Cammarosano S, Canosa A, Moglia C, Calvo A. The Role of APOE in the Occurrence of Frontotemporal Dementia in Amyotrophic Lateral Sclerosis. *JAMA Neurol.* 2016; 73:425–430.
 24. Connelly SJ, Mukaetova-Ladinska EB, Abdul-Ali Z, Alves da Silva J, Brayne C, Honer WG, Mann DM. Synaptic changes in frontotemporal lobar degeneration: correlation with MAPT haplotype and APOE genotype. *Neuropathol Appl Neurobiol.* 2011; 37:366–380.
 25. Daniele A, Matera MG, Seripa D, Acciarri A, Bizzarro A, Pilotto A, Masullo C. APOE epsilon 2/epsilon 4 genotype a risk factor for primary progressive aphasia in women. *Arch Neurol.* 2009; 66:910–912.
 26. Das G, Sadhukhan T, Sadhukhan D, Biswas A, Pal S, Ghosh A, Das SK, Ray K, Ray J. Genetic study on frontotemporal lobar degeneration in India. *Parkinsonism Relat Disord.* 2013; 19:487–489.
 27. Fabre SF, Forsell C, Viitanen M, Sjogren M, Wallin A, Blennow K, Blomberg M, Andersen C, Wahlund LO, Lannfelt L. Clinic-based cases with frontotemporal dementia show increased cerebrospinal fluid tau and high apolipoprotein E epsilon4 frequency, but no tau gene mutations. *Exp Neurol.* 2001; 168:413–418.
 28. Farrer LA, Abraham CR, Volicer L, Foley EJ, Kowall NW, McKee AC, Wells JM. Allele epsilon 4 of apolipoprotein E shows a dose effect on age at onset of Pick disease. *Exp Neurol.* 1995; 136:162–170.
 29. Feher A, Juhasz A, Rimanoczy A, Kalman J, Janka Z. Association between BDNF Val66Met polymorphism and Alzheimer disease, dementia with Lewy bodies, and Pick disease. *Alzheimer Dis Assoc Disord.* 2009; 23:224–228.
 30. Geschwind D, Karrim J, Nelson SF, Miller B. The apolipoprotein E epsilon4 allele is not a significant risk factor for frontotemporal dementia. *Ann Neurol.* 1998; 44:134–138.
 31. Gomez-Isla T, West HL, Rebeck GW, Harr SD, Growdon JH, Locascio JJ, Perls TT, Lipsitz LA, Hyman BT. Clinical and pathological correlates of apolipoprotein E epsilon 4 in Alzheimer's disease. *Ann Neurol.* 1996; 39:62–70.
 32. Gustafson L, Abrahamson M, Grubb A, Nilsson K, Fex G. Apolipoprotein-E genotyping in Alzheimer's disease and frontotemporal dementia. *Dement Geriatr Cogn Disord.* 1997; 8:240–243.
 33. Helisalmi S, Linnaranta K, Lehtovirta M, Mannermaa A, Heinonen O, Ryyänen M, Riekkinen P Sr, Soininen H. Apolipoprotein E polymorphism in patients with different neurodegenerative disorders. *Neurosci Lett.* 1996; 205:61–64.
 34. Hernandez I, Mauleon A, Rosense-Roca M, Alegret M, Vinyes G, Espinosa A, Sotolongo-Grau O, Becker JT, Valero S, Tarraga L, Lopez OL, Ruiz A, Boada M. Identification of misdiagnosed fronto-temporal dementia using APOE genotype and phenotype-genotype correlation analyses. *Curr Alzheimer Res.* 2014; 11:182–191.
 35. Ingelsson M, Fabre SF, Lilius L, Andersen C, Viitanen M, Almkvist O, Wahlund LO, Lannfelt L. Increased risk for frontotemporal dementia through interaction between tau polymorphisms and apolipoprotein E ϵ 4. *Neuroreport.* 2001; 12:905–909.
 36. Ji Y, Liu M, Huo YR, Liu S, Shi Z, Liu S, Wisniewski T, Wang J. Apolipoprotein E ϵ 4 Frequency Is Increased among Chinese Patients with Frontotemporal Dementia and Alzheimer's Disease. *Dement Geriatr Cogn Disord.* 2013; 36:163–170.

37. Jingtao W. The association study of apolipoprotein E gene polymorphism with different types of dementia in a Chinese Han population. *CAMS & PUMC*. 2010; 28–29.
38. Kalman J, Juhasz A, Majtenyi K, Rimanoczy A, Jakab K, Gardian G, Rasko I, Janka Z. Apolipoprotein E polymorphism in Pick's disease and in Huntington's disease. *Neurobiol Aging*. 2000; 21:555–558.
39. Kowalska A, Asada T, Arima K, Kumakiri C, Kozubski W, Takahashi K, Tabira T. Genetic analysis in patients with familial and sporadic frontotemporal dementia: two tau mutations in only familial cases and no association with apolipoprotein epsilon4. *Dement Geriatr Cogn Disord*. 2001; 12:387–392.
40. Lehmann DJ, Smith AD, Combrinck M, Barnetson L, Joachim C. Apolipoprotein E epsilon2 may be a risk factor for sporadic frontotemporal dementia. *J Neurol Neurosurg Psychiatry*. 2000; 69:404–405.
41. Lovati C, Galimberti D, Albani D, Bertora P, Venturelli E, Cislaghi G, Guidi I, Fenoglio C, Cortini F, Clerici F, Finazzi D, Forloni G, Scarpini E, et al. APOE epsilon2 and epsilon4 influence the susceptibility for Alzheimer's disease but not other dementias. *Int J Mol Epidemiol Genet*. 2010; 1:193–200.
42. Masullo C, Daniele A, Fazio VM, Seripa D, Gravina C, Filippini V, Grossi D, Fragassi N, Nichelli P, Leone M, Gainotti G. The Apolipoprotein E genotype in patients affected by syndromes with focal cortical atrophy. *Neurosci Lett*. 2001; 303:87–90.
43. Minthon L, Hesse C, Sjogren M, Englund E, Gustafson L, Blennow K. The apolipoprotein E epsilon4 allele frequency is normal in fronto-temporal dementia, but correlates with age at onset of disease. *Neurosci Lett*. 1997; 226:65–67.
44. Morenas-Rodriguez E, Cervera-Carles L, Vilaplana E, Alcolea D, Carmona-Iragui M, Dols-Icardo O, Ribosa-Nogue R, Munoz-Llahuna L, Sala I, Belen Sanchez-Saudinos M, Blesa R, Clarimon J, Fortea J, et al. Progranulin Protein Levels in Cerebrospinal Fluid in Primary Neurodegenerative Dementias. *J Alzheimers Dis*. 2016; 50:539–546.
45. Munoz-Ruiz MA, Hartikainen P, Hall A, Mattila J, Koikkalainen J, Herukka SK, Julkunen V, Vanninen R, Liu Y, Lotjonen J, Soininen H. Disease state fingerprint in frontotemporal degeneration with reference to Alzheimer's disease and mild cognitive impairment. *J Alzheimers Dis*. 2013; 35:727–739.
46. Pickering-Brown SM, Owen F, Isaacs A, Snowden J, Varma A, Neary D, Furlong R, Daniel SE, Cairns NJ, Mann DM. Apolipoprotein E epsilon4 allele has no effect on age at onset or duration of disease in cases of frontotemporal dementia with pick- or microvacuolar-type histology. *Exp Neurol*. 2000; 163:452–456.
47. Pickering-Brown SM, Siddons M, Mann DM, Owen F, Neary D, Snowden JS. Apolipoprotein E allelic frequencies in patients with lobar atrophy. *Neurosci Lett*. 1995; 188:205–207.
48. Premi E, Pilotto A, Alberici A, Papetti A, Archetti S, Seripa D, Daniele A, Masullo C, Garibotto V, Paghera B, Caobelli F, Padovani A, Borroni B. FOXP2, APOE, and PRNP: new modulators in primary progressive aphasia. *J Alzheimers Dis*. 2012; 28:941–950.
49. Rosso SM, Van Swieten JC, Roks G, Van Duijn CM, Heutink P, Cruts M, Van Broeckhoven C. Apolipoprotein E4 in the temporal variant of frontotemporal dementia. *J Neurol Neurosurg Psychiatry*. 2002; 72:820.
50. Ruiz A, Dols-Icardo O, Bullido MJ, Pastor P, Rodriguez-Rodriguez E, Lopez de Munain A, de Pancorbo MM, Perez-Tur J, Alvarez V, Antonell A, Lopez-Arrieta J, Hernandez I, Tarraga L, et al. Assessing the role of the TREM2 p.R47H variant as a risk factor for Alzheimer's disease and frontotemporal dementia. *Neurobiol Aging*. 2014; 35:444.e441–444.
51. Schneider JA, Gearing M, Robbins RS, de l'Aune W, Mirra SS. Apolipoprotein E genotype in diverse neurodegenerative disorders. *Ann Neurol*. 1995; 38:131–135.
52. Seripa D, Bizzarro A, Panza F, Acciarri A, Pellegrini F, Pilotto A, Masullo C. The APOE gene locus in frontotemporal dementia and primary progressive aphasia. *Arch Neurol*. 2011; 68:622–628.
53. Seripa D, Bizzarro A, Pilotto A, Palmieri O, Panza F, D'Onofrio G, Gravina C, Archetti S, Daniele A, Borroni B, Padovani A, Masullo C. TOMM40, APOE, and APOC1 in primary progressive aphasia and frontotemporal dementia. *J Alzheimers Dis*. 2012; 31:731–740.
54. Short RA, Graff-Radford NR, Adamson J, Baker M, Hutton M. Differences in tau and apolipoprotein E polymorphism frequencies in sporadic frontotemporal lobar degeneration syndromes. *Arch Neurol*. 2002; 59:611–615.
55. Sleegers K, Roks G, Theuns J, Aulchenko YS, Rademakers R, Cruts M, Van Gool WA, Van Broeckhoven C, Heutink P, Oostra BA, Van Swieten JC, Van Duijn CM. Familial clustering and genetic risk for dementia in a genetically isolated Dutch population. *Brain*. 2004; 127:1641–1649.
56. Srinivasan R, Davidson Y, Gibbons L, Payton A, Richardson AM, Varma A, Julien C, Stopford C, Thompson J, Horan MA, Pendleton N, Pickering-Brown SM, Neary D, et al. The apolipoprotein E epsilon4 allele selectively increases the risk of frontotemporal lobar degeneration in males. *J Neurol Neurosurg Psychiatry*. 2006; 77:154–158.
57. Steenland K, MacNeil J, Seals R, Levey A. Factors affecting survival of patients with neurodegenerative disease. *Neuroepidemiology*. 2010; 35:28–35.
58. van Blitterswijk M, Mullen B, Wojtas A, Heckman MG, Diehl NN, Baker MC, DeJesus-Hernandez M, Brown PH, Murray ME, Hsiung GY, Stewart H, Karydas AM, Finger E, et al. Genetic modifiers in carriers of repeat expansions in the C9ORF72 gene. *Mol Neurodegener*. 2014; 9:38.
59. Villa C, Ghezzi L, Fenoglio C, Clerici F, Marcone A, Benussi L, Ghidoni R, Gallone S, Serpente M, Cantoni C, Ridolfi E, Bonsi R, Cerami C, et al. Genetics and expression

- analysis of the specificity protein 4 gene (SP4) in patients with Alzheimer's disease and frontotemporal lobar degeneration. *J Alzheimers Dis.* 2012; 31:537–542.
60. Zintl M, Petkov M, Schmitz G, Hajak G, Klunemann HH. [Frontotemporal dementia in association with a family history of dementia and ApoE polymorphism]. [Article in German]. *Nervenarzt.* 2010; 81:75–78.
 61. Shi Z, Liu S, Xiang L, Wang Y, Liu M, Liu S, Han T, Zhou Y, Wang J, Cai L, Gao S, Ji Y. Frontotemporal dementia-related gene mutations in clinical dementia patients from a Chinese population. *J Hum Genet.* 2016.
 62. Rogers BS, Lippa CF. A clinical approach to early-onset inheritable dementia. *Am J Alzheimers Dis Other Demen.* 2012; 27:154–161.
 63. Holtzman DM, Bales KR, Tenkova T, Fagan AM, Parsadanian M, Sartorius LJ, Mackey B, Olney J, McKeel D, Wozniak D, Paul SM. Apolipoprotein E isoform-dependent amyloid deposition and neuritic degeneration in a mouse model of Alzheimer's disease. *Proc Natl Acad Sci USA.* 2000; 97:2892–2897.
 64. Castellano JM, Kim J, Stewart FR, Jiang H, DeMattos RB, Patterson BW, Fagan AM, Morris JC, Mawuenyega KG, Cruchaga C, Goate AM, Bales KR, Paul SM, et al. Human apoE isoforms differentially regulate brain amyloid-beta peptide clearance. *Sci Transl Med.* 2011; 3:89ra57.
 65. Mann DM, McDonagh AM, Pickering-Brown SM, Kowa H, Iwatsubo T. Amyloid beta protein deposition in patients with frontotemporal lobar degeneration: relationship to age and apolipoprotein E genotype. *Neurosci Lett.* 2001; 304:161–164.
 66. Brecht WJ, Harris FM, Chang S, Tesseur I, Yu GQ, Xu Q, Dee Fish J, Wyss-Coray T, Buttini M, Mucke L, Mahley RW, Huang Y. Neuron-specific apolipoprotein e4 proteolysis is associated with increased tau phosphorylation in brains of transgenic mice. *J Neurosci.* 2004; 24:2527–2534.
 67. Mackenzie IR, Neumann M, Bigio EH, Cairns NJ, Alafuzoff I, Kril J, Kovacs GG, Ghetti B, Halliday G, Holm IE, Ince PG, Kamphorst W, Revesz T, et al. Nomenclature and nosology for neuropathologic subtypes of frontotemporal lobar degeneration: an update. *Acta Neuropathol.* 2010; 119:1–4.
 68. Sorbi S, Hort J, Erkinjuntti T, Fladby T, Gainotti G, Gurvit H, Nacmias B, Pasquier F, Popescu BO, Rektorova I, Religa D, Rusina R, Rossor M, et al. EFNS-ENS Guidelines on the diagnosis and management of disorders associated with dementia. *Eur J Neurol.* 2012; 19:1159–1179.
 69. Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLoS Med.* 2009; 6:e1000097.