

Baicalin promotes the bacteriostatic activity of lysozyme on *S. aureus* in mammary glands and neutrophilic granulocytes in mice

Xuejiao Gao^{1,*}, Mengyao Guo^{2,*}, Zecai Zhang¹, Peng Shen¹, Zhengtao Yang¹, Naisheng Zhang¹

¹Department of Clinical Veterinary Medicine, College of Veterinary Medicine, Jilin University, Changchun, Jilin Province 130062, People's Republic of China

²Department of Clinical Veterinary Medicine, College of Veterinary Medicine, Huazhong Agricultural University, Wuhan 430070, People's Republic of China

*These authors have contributed equally to this work

Correspondence to: Naisheng Zhang, email: zhangns@jlu.edu.cn

Keywords: lysozyme, baicalin, *S. aureus*, bacteriostasis, mastitis

Received: July 20, 2016

Accepted: November 22, 2016

Published: February 08, 2017

ABSTRACT

***Staphylococcus aureus* causes mastitis as a result of community-acquired or nosocomial infections. Lysozyme (LYSO) is an enzyme that is upregulated in many organisms during the innate immune response against infection by bacterial pathogens. Baicalin is a bioactive flavonoid that can bind to enzymes, often to potentiate their effect. Here we tested the effects of baicalin on the activity of LYSO using the *S. aureus* mastitis mouse model and neutrophilic granulocyte model of *S. aureus* infection. In our experiments, *S. aureus* counts decreased with increasing baicalin concentration. Furthermore, qPCR and western blot analyses showed that LYSO expression was unaffected by baicalin, while fluorescence quenching and UV fluorescence spectral analyses showed that baicalin binds to LYSO. To test whether this binding increased LYSO activity, we assessed LYSO-induced bacteriostasis in the presence of baicalin. Our results showed that LYSO-induced *S. aureus* bacteriostasis increased with increasing concentrations of baicalin, and that baicalin binding to LYSO synergistically increased the antibacterial activity of LYSO. These results demonstrate that baicalin enhances LYSO-induced bacteriostasis during the innate immune response to *S. aureus*. They suggest baicalin is a potentially useful therapeutic agent for the treatment of bacterial infections.**

INTRODUCTION

Staphylococcus aureus, a major gram-positive pathogen, is classified as an opportunistic bacterial pathogen, which infects a diverse array of hosts [1]. *S. aureus* can be spread both through community-acquired and nosocomial infections [2]. Mastitis, which occurs most commonly by pathogenic infection during the postpartum period, poses a serious problem for humans [3] and other animals [4]. *S. aureus* is a common infectious pathogen that causes mastitis in both humans and animals [5]. *S. aureus* can infect mammary glands, resulting in serious long-term injury to mammary gland cells and ultimately damaging the structure and lactation function of the breast [6]. Until recently, there was no efficacious treatment for

S. aureus-induced mastitis in either humans or animals. Further research is still needed to improved prophylactic and therapeutic approaches to combat *S. aureus*-induced mastitis.

Local innate immunity helps to initiate and coordinate homeostatic and defense responses in mammary glands. Lysozyme (LYSO) is a major component of the innate immune system that helps fight against infection by bacterial pathogens and viruses. Indeed, LYSO contributes to surveillance of mammalian cells membranes, fortification of the phagocytic activity of neutrophils and macrophages [7], and secretion by polymorphonuclear leukocytes [8]. LYSO breaks bacterial cell walls via hydrolysis of peptidoglycan [9] and is thus particularly effective against Gram-positive bacteria.

It is extensively used in the pharmaceutical and food industries due to its anti-inflammatory, anti-viral, immune modulatory, anti-histaminic and anti-tumor effects [10].

Baicalin (Figure 1), a bioactive flavonoid isolated from the root of *scutellaria baicalensis*, exerts multiple biological functions [11–14]. Recent, studies have reported that baicalin regulates Toll-like receptors (TLRs) [15], NOD-like receptors (NODR) and other immune receptors in the innate and adaptive systems [16]. Previous studies have also suggested that LYSO can enhance the activity of some antibacterial drugs [17]. Here, we studied the effect of baicalin on the activity of LYSO in the progression of the innate immune response against infection by *S. aureus* pathogens.

RESULTS

S. aureus quantity in mammary gland tissues and neutrophil

The effect of baicalin on *S. aureus* quantity was measured by colony counting. Initially, the MIC of baicalin with *S. aureus* was confirmed at 1024 µg/ml. The concentration of baicalin was below 1024 µg/ml for treatment on tissues and neutrophils. There was no *S. aureus* in the control group in either the mammary gland tissues or neutrophils. The *S. aureus* counts were significantly increased in the MG of tissues and 0 µg/ml baicalin of neutrophils. Baicalin treatment significantly inhibited *S. aureus* growth in tissues and neutrophils in a dose dependent manner (Table 1).

Effect of baicalin on the expression of LYSO

Lysozyme is the major component of the innate immune system against infection by bacterial pathogens, particularly Gram-positive bacteria [18]. We analyzed the effect of baicalin on LYSO expression using qPCR and western blotting (Figure 2). Compared with controls,

LYSO mRNA and protein levels were increased by *S. aureus* infection in mammary tissues and neutrophils. Baicalin showed no effect on the expression of LYSO. LYSO mRNA and protein levels remained high in the presence of *S. aureus* infection without baicalin treatment. Varying baicalin concentrations did not affect LYSO levels either.

Binding of baicalin and LYSO

We used spectroscopy to measure potential interactions between baicalin and LYSO (Figure 3). Baicalin displayed fluorescence quenching with LYSO. The emission peak of LYSO moved from 337 nm to 294 nm. Fluorescence spectroscopy experiments showed that the fluorescence intensity of LYSO was reduced by baicalin in a dose-dependent manner (Figure 3B). We also measured UV fluorescence, which ranged between 200 and 500 nm in wavelength. These results showed that the UV fluorescence spectroscopy of LYSO was also altered by baicalin in a dose-dependent manner (Figure 3D).

Baicalin promotes the activity of LYSO

Micrococcus lysolei is typically used to measure the activity of LYSO by nephelometry. We used this method and our results confirmed the coactivation of baicalin and LYSO. The LYSO enzymatic activity value in the absence of baicalin was 47217.6 U/mg. The presence of baicalin enhanced LYSO activity in a dose-dependent manner (Figure 4).

Baicalin enhances the bacteriostatic effect of LYSO on *S. aureus*

The size of the bacteriostatic circle is the direct method for confirming bacteriostatic action. We performed antibacterial experiments *in vitro* to measure the bacteriostatic effects of LYSO on *S. aureus* in the presence

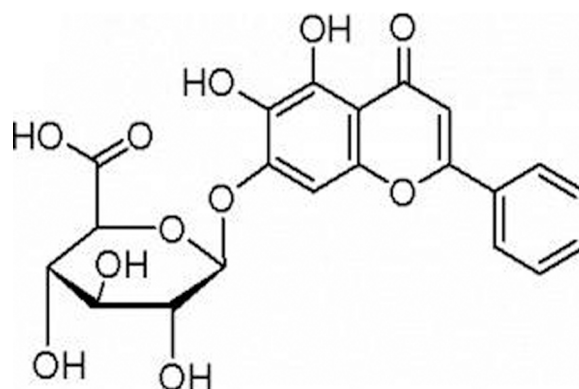


Figure 1: Chemical structure of baicalin.

Table 1: Effect of baicalin on count of *S. aureus* in mammary gland and neutrophils

Mammary tissues	CG	<i>S. aureus</i>	25 µg/g	50 µg/g	100 µg/g
(CFU/g)	0	2.2×10^{11}	2.7×10^9	2.2×10^7	2.2×10^5
neutrophils	CG	<i>S. aureus</i>	25 µg/ml	50 µg/ml	100 µg/ml
(CFU/mL)	0	2.2×10^8	1.2×10^6	2.4×10^5	1.3×10^4

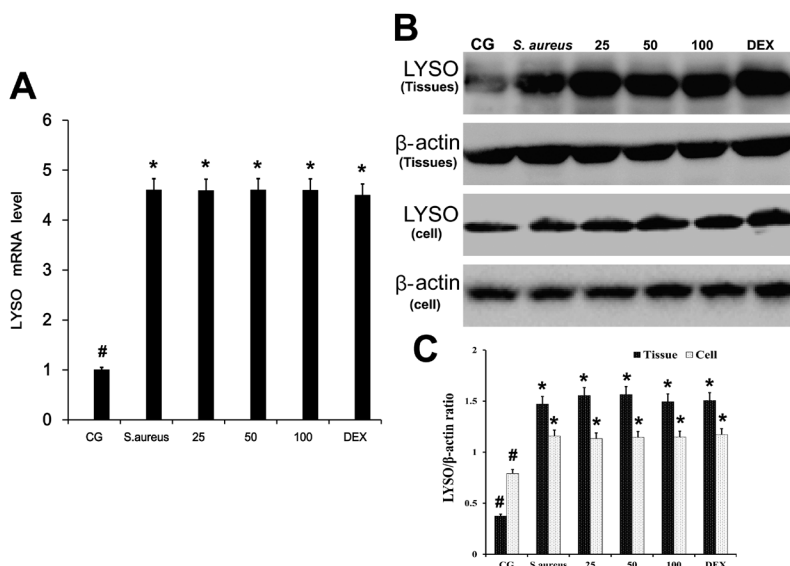


Figure 2: Effect of baicalin on the expression of LYSO. A. LYSO mRNA levels in mammary tissue. **B.** LYSO protein levels. **C.** LYSO/β-actin ratio analysis. CG is the control group, *S. aureus* is the microbionation group without baicalin treatment, and the baicalin administration group consists of, 25 mg/kg, 50 mg/kg and 100 mg/kg in the animals and 25 µg/ml, 50 µg/ml and 100 µg/ml in cells. β-actin served as the control. *p < 0.05 indicates a significant difference from the CG; #p < 0.05 indicates a significant difference from *S. aureus*.

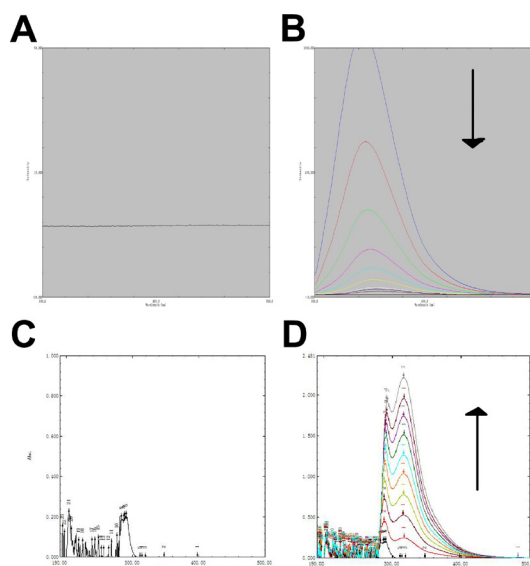


Figure 3: Spectroscopy experiment on the binding of baicalin and LYSO. A. Fluorescence spectroscopy of baicalin. **B.** Fluorescence spectroscopy of baicalin at different concentrations with LYSO. **C.** UV Fluorescence spectroscopy of LYSO. **D.** The effect of baicalin on LYSO with UV Fluorescence spectroscopy. The arrows indicate the concentrations of baicalin in increasing order (0, 10, 20, 30, 40, 50, 60, 70, 80 µg/ml).

of baicalin (Figure 5). Baicalin alone did not have any bacteriostatic effects on *S. aureus* at concentrations of 10, 20, 40 and 80 µg/ml. On the other hand, LYSO exerted antibacterial effects on *S. aureus* at 100 µg/ml. Furthermore, the size of the LYSO bacteriostatic circle was increased by baicalin in a dose-dependent manner. *Micrococcus lysolei* was used for the case-controlled study.

DISCUSSION

S. aureus is the main pathogen causing mastitis in both humans and animals [3]. Mastitis, an inflammatory disease of mammary glands, is caused by infections

during the postpartum period [19] and is closely related to the immune state of partial-breast [20]. An increase in the count of neutrophils is a sign of mastitis in mammary glands [21]. While polymorphonuclear leukocytes are the major cell type that control bacterial infections in mammals [22], LYSO is a major component against infection by bacterial pathogens in neutrophils [8]. Previous studies showed that baicalin treatment could significantly reduce the colony count of *S. aureus* in mammary tissues [15] and in neutrophils [1].

In the present study, the MIC of baicalin with *S. aureus* was confirmed at 1024 µg/ml, and the concentration of baicalin treatment on tissues and neutrophils was below 1024 µg/ml. Previous studies have shown that drugs can

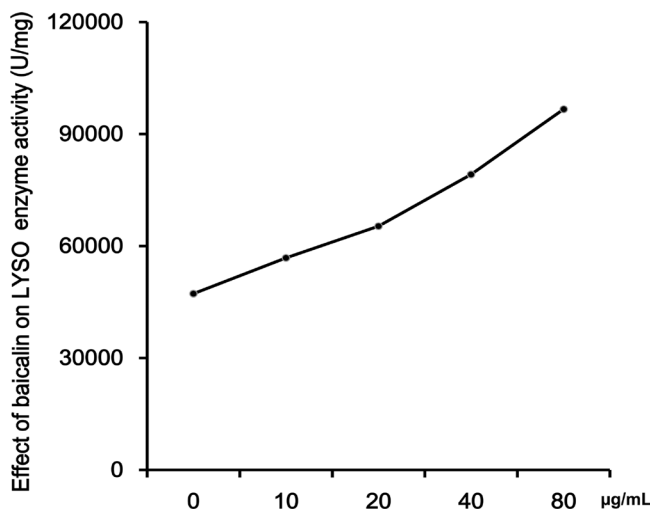


Figure 4: Effects of baicalin on the activity of LYSO. The coactivation of baicalin and LYSO was confirmed. Effects of baicalin on the activity of LYSO were analyzed at increasing concentrations of baicalin (0, 10, 20, 40, 80 µg/ml).

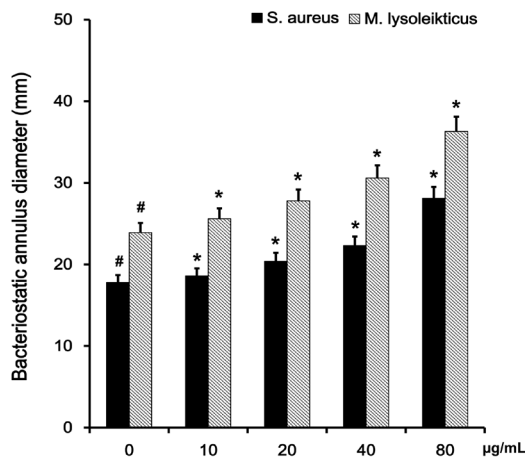


Figure 5: Effects of baicalin on the size of the bacteriostatic circle of LYSO. The experiment to measure the antibacterial effect of LYSO on *S. aureus* with baicalin was performed *in vitro*. *Micrococcus lysolei* was used for the case-controlled study. The concentrations of baicalin used were 0, 10, 20, 40, 80 µg/ml. *p < 0.05 indicates a significant difference from 0 µg/ml baicalin.

Table 2: Oligonucleotide primers used for qPCR

Name	Primer sequence	Product size (bp)
LYSO	Sense: 5'- CCAGCCTCCAGTCACCAT -3' Anti-sense: 5'-TCGTTTTACTCCAATTTAT-3'	378
β -actin	Sense: 5'-TAAAACGCAGCTCAGTAACAGTCGG-3' Anti-sense: 5'-TGCAATCCTGTGGCATCCATGAAAC-3'	182

increase the expression or activity of enzymes [23]. In our study here, the expression of LYSO was not affected by baicalin in either mammary tissues or neutrophils. However, our fluorescence spectroscopy experiments on baicalin combined with lysozyme were consistent with previous studies [24]. LYSO is a small globular protein, consisting of 129 amino acid residues with six tryptophans, two tyrosines and four disulfide bonds and exhibits helical (α -helix), pleated sheet (β -sheet), turn (β -turn) and random coil secondary structure. These structural elements allow LYSO to bind drugs to aid in the treatment of some illnesses [25]. LYSO exhibits bacteriostatic activity and its interactions with some drugs (e.g., antibiotics) can have a synergistic effect [26, 27].

We used *M. lysolei* (as typically done) to measure the activity of LYSO with nephelometry in the absence and presence of baicalin. LYSO breaks bacterial cell walls made of peptidoglycan via hydrolysis of the β (1–4) glucosidic bond between N-acetylmuramic acid and N-acetylglucosamine [28, 29]. *M. lysolei* possesses a peptidoglycan wall [30]. Our results showed that baicalin enhanced the activity of LYSO in a dose-dependent manner. Since *S. aureus* is a Gram-positive bacterium [31], this suggests that its peptidoglycan cell wall was destroyed, resulting in apoptosis. Measuring the size of the bacteriostatic circle is the standard method to test for bacteriostatic action [32]. We analyzed the antibacterial activity of LYSO on *S. aureus* under different concentrations of baicalin. Our results showed that with increasing concentrations of baicalin, the size of the bacteriostatic circle of LYSO on *S. aureus* increased. A previous study showed that baicalin could increase the antiseptic action of acetoamycin on *S. aureus*. Our results demonstrated that baicalin could also enhance the antiseptic activity of LYSO on *S. aureus*.

A previous study by our group showed that baicalin played an important role in immune regulation [15]. Local innate immunity helps to maintain homeostasis and to trigger defense responses in mammary glands [33]. LYSO is a major component in the innate immune system against infection by bacterial pathogens. In the present study, we found that baicalin inhibited the growth of the *S. aureus* by enhancing the antiseptic activity of LYSO secreted by neutrophils in mammary gland tissues. Our results here warrant further

studies to firmly establish whether baicalin should be administered as a drug to treat *S. aureus* mastitis.

MATERIALS AND METHODS

S. aureus

S. aureus (ATCC 35556) was purchased from the American Type Culture Collection (ATCC, USA). *S. aureus* were resuspended in 1,000 μ l PBS (2×10^8 CFU per 30 μ l). Fifty milliliters of culture aliquots were centrifuged and washed with phosphate-buffered saline (PBS) prior to resuspension. A 100- μ l suspension was used per breast. The *S. aureus* suspension was injected via the teat canal of breast to induce infection in the mammary gland.

MIC of baicalin

Baicalin (purity >99.9 %) was purchased from Sigma Chemical Company. The minimal inhibitory concentration (MIC) of baicalin with *S. aureus* was detected using the broth micro dilution method, according to the American Institute of Standardization of Clinical Laboratory (CLSI) guidelines. Baicalin was diluted multiple times with bouillon culture medium in 96-microwells plates. The initial concentration was 4096 μ g/ml. Next, 100 μ l of bacterium suspension were added to each well. The reaction mixture was incubated for 24 h at 37°C. The value was evaluated by measuring the change in absorbance at 600 nm using a 96-well plate reader. The percentage of growth (test/*S. aureus*) was shown as the MIC.

Experimental animal and drug administration

Sixty adult female BALB/c mice (6-8 w old, weighing 25-28 g) were used in the present study and were provided by the Center of Experimental Animals of Baiqien Medical College of Jilin University in China. The procedures were performed according to the NIH Guidelines for the care and use of Laboratory Animals and were approved by the Institutional Animal Care and Use Committee of Jilin University. Baicalin was dissolved in physiological saline and intraperitoneally injected three times at 6 h, 12 h and 24 h after the

microbionation in the mammary gland. Mice were postpartum in lactation period and divided into three groups as follows: 1) Microbionation group (MG), the mouse model of *S. aureus* mastitis without drug treatment; 2) baicalin administration group, which was subjected to mastitis by *S. aureus* and intraperitoneally administered baicalin at 25 mg/kg, 50 mg/kg, or 100 mg/kg, as previously described [15]; 3) control group (CG), in which mice were treated with normal saline as the vehicle control at the same volume and time point as baicalin for the baicalin administration group. *S. aureus* culture broth was centrifuged and washed with phosphate-buffered saline (PBS). Then, *S. aureus* were resuspended in PBS (1×10^7 CFU per 10 μ l). A 100- μ l suspension was administered per breast. The *S. aureus* suspension was injected via teat canal to induce a mammary gland infection. The mammary gland tissues were weighed and homogenized with phosphate buffered saline (w/v: 1/9) on ice, and then coated bacteria underwent colony counting, while others were centrifuged at $2,000 \times g$ for 40 min at 4°C. The supernatant was collected for LYSO detection.

Neutrophil granulocyte cell culture and drug treatment

Neutrophil granulocytes were separated from fractions of heart blood obtained from mice using a Percoll separation medium density gradient. The purified neutrophils were seeded at a concentration of 3×10^7 cells/well into 6-well plates and cultured in RPMI1640 medium supplemented with 10% fetal bovine serum in a humidified atmosphere of 5% CO₂. After 2 h, different concentrations of baicalin were added to the wells of 6-well plates and incubated for 1 h. The concentrations were 0, 10, 20, 40 and 80 μ g/ml. The last well was the control with neither drug nor bacteria. Infection was induced using *S. aureus*. The viability of the bacteria was routinely confirmed by counting the number of colonies to determine the CFU/ml. Bactericidal antibacterial assays were carried out for 2 h at 37°C at a multiplicity of infection of 1:5. Then, the dilution was plated on TBS agar, and colonies were counted.

Quantitative real-time polymerase chain reaction

Total RNA was isolated from the tissue and PMN cell samples using the TRIzol reagent and synthesized first-strand cDNA using oligo(dT) primers and Superscript II reverse transcriptase, according to the manufacturer's instructions (Invitrogen, USA). Synthesized cDNA was diluted five times with sterile water and stored at -80°C. The Primer Premier software (PREMIER Biosoft International, USA) was used to design specific primers for LYSO and β -actin based on known sequences (Table 2). Quantitative real-time PCR was performed on an ABI PRISM 7500 Detection System (Applied

Biosystems, USA). Reactions were performed in a 25- μ l reaction mixture, under the following conditions: 95°C for 30 s, followed by 35 cycles of 95°C for 15 s, 63°C for 30 s and 60°C for 30 s. The results (fold changes) were expressed as $2^{-\Delta\Delta Ct}$. β -actin served as the reference gene.

Western blotting analyses

The total protein of the mammary gland tissue and PMN cell samples was extracted according to the manufacturer's recommended protocol (Invitrogen, Beijing, China). The protein concentrations were determined using the BCA Protein Assay Kit and equal amounts of protein (50 μ g) for fractionation on 10% SDS polyacrylamide gels. The proteins were then transferred onto polyvinylidene difluoride membranes and blocked. The membranes were then incubated with primary antibodies (LYSO and β -actin). After several washes, the membranes were incubated with secondary antibodies. Protein expression was measured using an Enhanced Chemiluminescence Detection System. β -actin was used as a loading control.

Fluorescence measurements

Fluorescence spectroscopy experiments were executed by Fluorescence Spectroscopy (SHIMADZU Co. Japan). Spectra were measured at two temperatures (298 k and 310 k), with an excitation wavelength at 280 nm and an emission wavelength from 300 to 500 nm. The slit widths for both the excitation and emission channels were 5 nm. The samples were placed in a 1 \times 1 cm path length quartz cuvette (2 ml). To eliminate the inner filter effects of LYSO and baicalin, the fluorescence intensities were corrected using the equation: $F_{cor} = F_{obs}10^{(A1+A2)/2}$, where F_{obs} is the observed fluorescence, and $A1$ and $A2$ are the sum of the absorbance of LYSO and baicalin at the measured condition, respectively. UV fluorescence measurements were also used, and the wavelength range was between 200-500 nm.

Antibacterial activity of LYSO correlates positively with varying baicalin concentration

Micrococcus lysodeikticus (*M. lysodeikticus*), Gram-positive bacteria, was usually a positive result for the antibacterial activity of LYSO. We selected *M. lysodeikticus* and *S. aureus*. The two bacteria were inoculated in sterile nutrient broth at 37°C for 24 h, and they were then spread on the surface of nutrient agar discs, and seeded in 20- μ l test culture containing 1×10^8 CFU/ml. The antibacterial activity of baicalin in the presence of LYSO was assessed using the agar disc diffusion method. Whatman No. 1 filter papers (diameter 4 mm) impregnated in 100 μ g/ml LYSO solution with different concentrations (0, 10, 20, 40, 80 μ g/ml) of baicalin were placed on the surface of the nutrient agar. The control disk

was placed with water to assess the effect of water on the pathogens. The plates were incubated at 37°C for 24 h, and antibacterial activity was measured based on the inhibitory zone. The antibacterial activity test was also assessed by the value of OD_{600nm} using UV-spectrophotometry. The two bacteria were inoculated in sterile nutrient broth at 37°C for 24 h, and then each 20- μ l aliquot was spread onto 4-ml sterile nutrient broth and incubated at 37°C for 24 h.

Data analyses

Statistical analyses were performed using the SPSS software package (ver. 13 for Windows; SPSS Inc., Chicago, IL, USA). Significance was determined using one-way ANOVA with a significance level of *p-value* < 0.05. The data were assessed using the Tukey-Kramer method for multiple comparisons. All values were expressed as the mean \pm SD.

ACKNOWLEDGMENTS

This work was supported by grants from the National Natural Science Foundation of China (nos. 31272622, 31201925, and 31502130) and the Graduate Innovation Fund of Jilin University (Grant no. 2015034).

CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

REFERENCES

- Schweizer ML, Bossen A, McDanel JS, Dennis LK. Staphylococcus aureus Colonization before Infection Is Not Associated with Mortality among S. aureus-Infected Patients: A Meta-analysis. *Infection Control and Hospital Epidemiology*. 2012; 33: 796-802. doi: 10.1086/666628.
- Li B, Li J, Pan X, Ding G, Cao H, Jiang W, Zheng J, Zhou H. Artesunate protects sepsis model mice challenged with Staphylococcus aureus by decreasing TNF-alpha release via inhibition TLR2 and Nod2 mRNA expressions and transcription factor NF-kappaB activation. *Int Immunopharmacol*. 2010; 10: 344-50. doi: 10.1016/j.intimp.2009.12.006.
- Zadoks RN, Middleton JR, McDougall S, Katholm J, Schukken YH. Molecular epidemiology of mastitis pathogens of dairy cattle and comparative relevance to humans. *J Mammary Gland Biol Neoplasia*. 2011; 16: 357-72. doi: 10.1007/s10911-011-9236-y.
- Sinha MK, Thombare NN, Mondal B. Subclinical mastitis in dairy animals: incidence, economics, and predisposing factors. *Scientific World Journal*. 2014; 2014: 523984. doi: 10.1155/2014/523984.
- Reinoso EB, El-Sayed A, Lammler C, Bogni C, Zschock M. Genotyping of Staphylococcus aureus isolated from humans, bovine subclinical mastitis and food samples in Argentina. *Microbiol Res*. 2008; 163: 314-22. doi: 10.1016/j.micres.2006.05.013.
- Guo MY, Zhang NS, Li DP, Liang DJ, Liu ZC, Li FY, Fu YH, Cao YG, Deng XM, Yang ZT. Baicalin plays an anti-inflammatory role through reducing nuclear factor-kappa B and p38 phosphorylation in S. aureus-induced mastitis. *International Immunopharmacology*. 2013; 16: 125-30. doi: 10.1016/j.intimp.2013.03.006.
- Paviani V, Queiroz RF, Marquez EF, Di Mascio P, Augusto O. MS Characterization of the Dityryptophan Cross-Links in Lysozyme Submitted to Enzymatically and Photolytically Generated Carbonate Radicals. *Free Radical Biology and Medicine*. 2014; 76: S159-S. doi: 10.1016/j.freeradbiomed.2014.10.068.
- Biggar WD. Effect of phorbol myristate acetate on cellular metabolism and lysozyme release from alveolar macrophages and polymorphonuclear leukocytes. *Infect Immun*. 1978; 21: 669-71.
- Pushkaran AC, Nataraj N, Nair N, Gotz F, Biswas R, Mohan CG. Understanding the Structure-Function Relationship of Lysozyme Resistance in Staphylococcus aureus by Peptidoglycan O-Acetylation Using Molecular Docking, Dynamics, and Lysis Assay. *J Chem Inf Model*. 2015; 55: 760-70. doi: 10.1021/ci500734k.
- Figueiredo TA, Ludovice AM, Sobral RG. Contribution of peptidoglycan amidation to beta-lactam and lysozyme resistance in different genetic lineages of Staphylococcus aureus. *Microb Drug Resist*. 2014; 20: 238-49. doi: 10.1089/mdr.2014.0042.
- Jeong HS, Gu GE, Jo AR, Bang JS, Yun HY, Baek KJ, Kwon NS, Park KC, Kim DS. Baicalin-induced Akt activation decreases melanogenesis through downregulation of microphthalmia-associated transcription factor and tyrosinase. *European Journal of Pharmacology*. 2015; 761: 19-27. doi: 10.1016/j.ejphar.2015.04.028.
- Liao P, Liu L, Wang B, Li W, Fang X, Guan S. Baicalin and geniposide attenuate atherosclerosis involving lipids regulation and immunoregulation in ApoE-/- mice. *Eur J Pharmacol*. 2014; 740: 488-95. doi: 10.1016/j.ejphar.2014.06.039.
- Zhou BR, Yin HB, Xu Y, Wu D, Zhang ZH, Yin ZQ, Permatasari F, Luo D. Baicalin protects human skin fibroblasts from ultraviolet A radiation-induced oxidative damage and apoptosis. *Free Radic Res*. 2012; 46: 1458-71. doi: 10.3109/10715762.2012.726355.
- Zhang Y, Huang Y, Deng X, Xu Y, Gao Z, Li H. Iron overload-induced rat liver injury: Involvement of protein tyrosine nitration and the effect of baicalin. *Eur J Pharmacol*. 2012; 680: 95-101. doi: 10.1016/j.ejphar.2012.01.010.
- Guo M, Cao Y, Wang T, Song X, Liu Z, Zhou E, Deng X, Zhang N, Yang Z. Baicalin inhibits Staphylococcus aureus-induced apoptosis by regulating TLR2 and TLR2-related apoptotic factors in the mouse mammary glands.

- Eur J Pharmacol. 2014; 723: 481-8. doi: 10.1016/j.ejphar.2013.10.032.
16. Nayak MK, Agrawal AS, Bose S, Naskar S, Bhowmick R, Chakrabarti S, Sarkar S, Chawla-Sarkar M. Antiviral activity of baicalin against influenza virus H1N1-pdm09 is due to modulation of NS1-mediated cellular innate immune responses. *Journal of Antimicrobial Chemotherapy*. 2014; 69: 1298-310. doi: 10.1093/jac/dkt534.
 17. Kiristi M, Singh VV, de Avila BEF, Uygun M, Soto F, Uygun DA, Wang J. Lysozyme-Based Antibacterial Nanomotors. *Acs Nano*. 2015; 9: 9252-9. doi: 10.1021/acsnano.5b04142.
 18. Arabski M, Konieczna I, Tusinska E, Wasik S, Relich I, Zajac K, Kaminski ZJ, Kaca W. The use of lysozyme modified with fluorescein for the detection of Gram-positive bacteria. *Microbiological Research*. 2015; 170: 242-7. doi: 10.1016/j.micres.2014.05.004.
 19. Aryeetey RNO, Marquis GS, Timms L, Lartey A, Brakohiapa L. Subclinical mastitis is common among Ghanaian women lactating 3 to 4 months postpartum. *Journal of Human Lactation*. 2008; 24: 263-7. doi: 10.1177/0890334408316077.
 20. Swinkels JM, Hogeveen H, Zadoks RN. A partial budget model to estimate economic benefits of lactational treatment of subclinical *Staphylococcus aureus* mastitis. *Journal of Dairy Science*. 2005; 88: 4273-87.
 21. Burvenich C, Monfardini E, Mehrzad J, Capuco AV, Paape MJ. Role of neutrophil polymorphonuclear leukocytes during bovine coliform mastitis: physiology or pathology? *Verh K Acad Geneesk Belg*. 2004; 66: 97-150; discussion -3.
 22. Wanahita A, Goldsmith EA, Musher DM, Clarridge JE, Rubio J, Krishnan B, Trial J. Interaction between human polymorphonuclear leukocytes and streptococcus milleri group bacteria. *Journal of Infectious Diseases*. 2002; 185: 85-90. doi: 10.1086/338145.
 23. Lin SM, Wang SW, Ho SC, Tang YL. Protective effect of green tea (-)-epigallocatechin-3-gallate against the monoamine oxidase B enzyme activity increase in adult rat brains. *Nutrition*. 2010; 26: 1195-200. doi: 10.1016/j.nut.2009.11.022.
 24. Huang Y, Cui LJ, Wang JM, Huo K, Chen C, Zhan WH, Dou YH. Comparative studies on interactions of baicalin, baicalin and scutellarin with lysozyme. *European Journal of Medicinal Chemistry*. 2011; 46: 6039-45. doi: 10.1016/j.ejmech.2011.10.016.
 25. Aravind UK, Mathew J, Aravindakumar CT. Transport studies of BSA, lysozyme and ovalbumin through chitosan/polystyrene sulfonate multilayer membrane. *Journal of Membrane Science*. 2007; 299: 146-55. doi: 10.1016/j.memsci.2007.04.036.
 26. Oliver WT, Wells JE. Lysozyme as an alternative to growth promoting antibiotics in swine production. *Journal of Animal Science and Biotechnology*. 2015; 6. doi: ARTN 3510.1186/s40104-015-0034-z.
 27. Biziulevicus GA, Biziuleviciene G, Kazlauskaite J. Lysozyme and similar lytic enzyme preparations should be considered antibiotics. *Medical Hypotheses*. 2007; 68: 1420-. doi: 10.1016/j.mehy.2006.11.016.
 28. Benachour A, Ladjouzi R, Le Jeune A, Hebert L, Thorpe S, Courtin P, Chapot-Chartier MP, Prajsnar TK, Foster SJ, Mesnage S. The Lysozyme-Induced Peptidoglycan N-Acetylglucosamine Deacetylase PgdA (EF1843) Is Required for *Enterococcus faecalis* Virulence. *Journal of Bacteriology*. 2012; 194: 6066-73. doi: 10.1128/Jb.00981-12.
 29. Rae CS, Geissler A, Adamson PC, Portnoy DA. Mutations of the *Listeria monocytogenes* Peptidoglycan N-Deacetylase and O-Acetylase Result in Enhanced Lysozyme Sensitivity, Bacteriolysis, and Hyperinduction of Innate Immune Pathways. *Infection and Immunity*. 2011; 79: 3596-606. doi: 10.1128/iai.00077-11.
 30. Cooke DG, Blackwell LF. Clearing of suspensions of *Micrococcus lysodeikticus* catalysed by lysozymes from hen, goose, and turkey egg whites, human milk, and phage T4. Assessment of potential as signal generators for homogeneous enzyme immunoassays for urinary steroids. *Journal of Immunoassay & Immunochemistry*. 2007; 28: 67-90. doi: 10.1080/15321810701209704.
 31. Hepburn L, Prajsnar TK, Klapholz C, Moreno P, Loynes CA, Ogryzko NV, Brown K, Schiebler M, Hegyi K, Antrobus R, Hammond KL, Connolly J, Ochoa B, et al. Innate immunity. A Spatzle-like role for nerve growth factor beta in vertebrate immunity to *Staphylococcus aureus*. *Science*. 2014; 346: 641-6. doi: 10.1126/science.1258705.
 32. Jordan MJ, Lax V, Rota MC, Loran S, Sotomayor JA. Influence of the Bioclimatic Area on the Polyphenolic Composition, and Antioxidant and Bacteriostatic Activities of *Rosmarinus officinalis*. *Natural Product Communications*. 2013; 8: 817-22.
 33. Reardon C, Duncan GS, Brustle A, Brenner D, Tusche MW, Olofsson PS, Rosas-Ballina M, Tracey KJ, Mak TW. Lymphocyte-derived ACh regulates local innate but not adaptive immunity. *Proc Natl Acad Sci U S A*. 2013; 110: 1410-5. doi: 10.1073/pnas.1221655110.