

Associations between single-nucleotide polymorphisms of human exonuclease 1 and the risk of hepatocellular carcinoma

Shengkui Tan¹, Ruoyun Qin², Xiaonian Zhu¹, Chao Tan³, Jiale Song¹, Linyuan Qin¹, Liu Liu², Xiong Huang², Anhua Li³, Xiaoqiang Qiu²

¹Department of Epidemiology, School of Public Health, Guilin Medical University, Guilin 541004, Guangxi, People's Republic of China

²Department of Epidemiology, School of Public Health, Guangxi Medical University, Nanning 530021, Guangxi, People's Republic of China

³Guangxi Center for Disease Prevention and Control, Nanning 530021, Guangxi, People's Republic of China

Correspondence to: Xiaoqiang Qiu, email: xqiu9999@sina.com

Keywords: hepatocellular carcinoma (HCC), human exonuclease 1 (hEXO1), single-nucleotide polymorphisms (SNPs), interaction

Received: May 31, 2016

Accepted: October 17, 2016

Published: November 23, 2016

ABSTRACT

Human exonuclease 1 (*hEXO1*) is an important nuclease involved in mismatch repair system that contributes to maintain genomic stability and modulate DNA recombination. This study is aimed to explore the associations between single-nucleotide polymorphisms (SNPs) of *hEXO1* and the hereditary susceptibility of hepatocellular carcinoma (HCC). SNPs rs1047840, rs1776148, rs3754093, rs4149867, rs4149963, and rs1776181 of *hEXO1* were examined from a hospital-based case-control study including 1,196 cases (HCC patients) and 1,199 controls (non-HCC patients) in Guangxi, China. We found the rs3754093 AG genotype decreased the risk of HCC (OR=0.714, 95% CI: 0.539~0.946). According to the results of stratification analysis, rs3754093 mutant genotype AG/GG decreased the risk of HCC with some HCC protective factors such as non-smoking, non-alcohol consumption and non-HCC family history, but also decreased the risk of HCC with HBV infection. Moreover, it was correlated to non-tumor metastasis and increased the survival of HCC patients. The results from gene-environment interaction assay indicated all *hEXO1* SNPs interacted with smoking, alcohol consumption, HBV infection in pathogenesis of HCC. However, gene-gene interaction assay suggested the interaction between rs3754093 and other 5 SNPs were associated with reducing the HCC risk. These results suggest rs3754093 exhibits a protective activity to decrease the incidence risk of HCC in Guangxi, China. In addition, all SNPs in this study interacted with environment risk factors in pathogenesis of HCC.

INTRODUCTION

Hepatocellular carcinoma (HCC) is a very common digestive system carcinoma which is the fifth most prevalent cancer and the third most frequent cause of cancer mortality globally [1]. Each year there are approximately 630,000 new cases of HCC in the world and more than half of the new cases occur in China [2]. It is noticed that the Southern Guangxi has one of the highest occurrence of liver cancer in China [3]. Epidemiological studies suggest that the etiology of HCC is a complicated disease caused by a multi-stage process involving multiple genetic or environmental factors including alcohol consumption, tobacco use, hepatitis

B virus (HBV) and hepatitis C virus (HCV) infection, as well as HCC-family history [4–7]. However, only a minority of people with established risk factors eventually develop HCC, suggesting that other environmental and/or genetic factors may play a role in HCC development. In genomic levels, dysfunctions of some oncogenes and tumor suppressor genes induced the progressive growth of malignant cells, and caused HCC.

DNA mismatch repair (MMR) is a component of the DNA repair mechanism, which plays a key role in maintaining genomic stability, preventing gene mutation, the process of DNA replication, and modulates DNA recombination and mediates cell cycle arrest [8]. Human

Table 1: Demographic information of the study objects

Characteristics	Controls (n=1199)	Cases (n=1196)	t/χ^2	<i>P</i>
Age(years, \pm s)	48.28 \pm 11.69	48.58 \pm 10.84	-0.648	0.517
Gender			0.042	0.837
Male	1045	1039		
Female	154	157		
Nation			3.005	0.223
Han	753	791		
Zhuang	423	386		
Others	23	19		
Smoking			140.90	0.000
Yes	1008	749		
No	191	447		
Alcohol Consumption			141.74	0.000
Yes	1036	785		
No	163	411		
HBV infection			1375.31	0.000
Yes	98	195		
No	1101	1001		
HCC family history			42.76	0.000
Yes \bar{x}	1189	1130		
No	10	66		

exonuclease 1 (*hEXO1*) which located at chromosome 1q42-q43, contains one untranslated exon followed by 13 coding exons and encodes an 846 amino acid protein [9–11]. *EXO1* is an important member of DNA MMR system which has been well documented to implicate in DNA replication, DNA repair, DNA restructuring, and maintain the stability of telomeres [12]. *EXO1* can interact physically with the MMR proteins MSH2 and MLH1 in yeast and human cells, and with MSH3 in human cells [9, 13–17]. Wei indicated that mammalian *EXO1* is responsible for mutation prevention and mice *EXO1* inactivation reduced survival time and increased risk of lymphoma [18]. In addition, some studies have indicated SNPs of *hEXO1* were associated with various cancers including lung cancer, colorectal cancer, gastric cancer, breast cancer, oral cancer, and other tumors [10, 19–25]. A number of studies have reported a SNP of the *hEXO1* gene, K589E (rs1047840), is associated with human lung cancer, breast cancer, oral cancer and gastric cancer risk in Chinese Taiwan population [24, 26–28]. Jin [29] and Luo [30] reported the *hEXO1* K589E was associated with human lung cancer and cervical cancer susceptibility in Chinese Mainland population. The *hEXO1* K589E polymorphism may be a genetic susceptibility factor of

HCC in the Turkish population [31], and increase the risk of colorectal cancer in the Polish population [32].

However, the association between SNPs of *hEXO1* and hereditary susceptibility of HCC has not been investigated in China. In this study, we conducted a screening on *hEXO1* from NIEHS database to seek candidate SNPs in Chinese population. Minor allele frequency (MAF) of six SNPs (rs1047840, rs1776148, rs3754093, rs4149867, rs4149963, and rs1776181) was greater than 0.05 in Chinese population and had potential functions. Therefore, these 6 SNPs were selected to investigate their frequency distributions and associations with HCC in Guangxi, China. We expect the study will provide scientific basis for prevention and treatment of HCC.

RESULTS

Demographic information of HCC patients and controls

The demographic information of this study is presented in Table 1. There were no statistical differences in the age, sex and nation distribution between HCC patients and controls ($P>0.05$). However, 4 risk factors

Table 2: Distribution of genotypes and risk of HCC

Genotypes	Controls, n (%)	Cases, n (%)	OR (95%CI) ^a	OR (95%CI) ^b
rs1047840				
AA	54(4.50)	64(5.35)	1.000	1.000
AG	378(31.53)	388(32.44)	0.866(0.587~1.278)	0.769(0.419~1.410)
GG	767(63.97)	744(62.21)	0.818(0.562~1.192)	0.789(0.440~1.417)
AG/GG	1145(95.50)	1132(94.65)	0.834(0.575~1.209)	0.782(0.439~1.394)
rs1776148				
AA	38(3.17)	43(3.60)	1.000	1.000
AG	362(30.19)	355(29.68)	0.867(0.547~1.373)	0.614(0.312~1.207)
GG	799(66.64)	798(66.72)	0.883(0.564~1.380)	0.547(0.284~1.054)
AG/GG	1161(96.83)	1153(96.40)	0.878(0.563~1.368)	0.568(0.297~1.086)
rs3754093				
AA	401(33.44)	441(36.87)	1.000	1.000
AG	580(48.37)	557 (46.57)	0.873(0.731~1.044)	0.714(0.539~0.946)
GG	218(18.18)	198(16.55)	0.826(0.653~1.045)	0.822(0.567~1.193)
AG/GG	798(66.56)	755(63.12)	0.906(0.766~1.071)	0.741 (0.569~0.966)
rs4149867				
CC	858(71.56)	718(68.39)	1.000	1.000
CT	316(26.35)	347 (29.01)	1.312(1.094~1.574)	0.962 (0.723~1.281)
TT	25(2.09)	31(2.59)	1.482(0.867~2.533)	0.767(0.337~1.749)
CT/TT	341(28.44)	378(31.60)	1.325(1.110~1.581)	0.945 (0.715~1.248)
rs4149963				
CC	1057(88.15)	1047(87.54)	1.000	1.000
CT	129(10.76)	124(10.37)	0.970(0.748~1.260)	0.805 (0.531~1.222)
TT	13(1.09)	25(2.09)	1.941(0.988~3.815)	1.270(0.410~3.932)
CT/TT	142(11.85)	149(12.46)	1.059(0.829~1.354)	0.846 (0.570~1.257)
rs1776181				
CC	644(53.71)	609(50.92)	1.000	1.000
CT	445(37.11)	447(37.37)	1.062(0.895~1.261)	0.939 (0.716~1.230)
TT	110(9.17)	140(11.71)	1.346(1.024~1.769)	0.873(0.563~1.353)
CT/TT	555(46.28)	587(49.08)	1.118(0.953~1.313)	0.925 (0.717~1.192)

^a:OR and 95%CI without adjusting. ^b:Adjusted by logistic regression for age, gender, nations, smoking, alcohol consumption, HBV infection, and HCC family history.

including HBV infection, alcohol consumption, smoking and HCC family history showed a significantly statistical differences between HCC patients and controls ($P<0.05$).

Distribution of genotypes and risk of HCC

The allele and genotype distributions of the six SNPs in this study and their associations with HCC

risk are presented at Table 2. The distribution of all the SNPs’ genotypes in the controls obeyed Hardy-Weinberg equilibrium (HWE) ($P> 0.05$). The chi-square test also showed distribution of all the SNPs’ genotypes in the cases and the controls had no statistic difference ($P>0.05$). Following adjust the nation, gender, age, HBV infection, alcohol consumption, smoking, and HCC family history in Logistic regression models, the rs3754093AG (adjusted

Table 3: Results of rs3754093 stratification analysis

Variables	Genotypes	Controls (n=1199)	Cases (n=1196)	OR (95%CI) ^a	P
Smoking					
No	AA	336	278	1.000	0.029
	AG/GG	672	471	0.701(0.510~0.963)	
Yes	AA	65	163	1.000	0.246
	AG/GG	126	284	0.485(0.143~1.646)	
Alcohol Consumption					
No	AA	345	296	1.000	0.005
	AG/GG	691	489	0.644(0.473~0.877)	
Yes	AA	56	145	1.000	0.716
	AG/GG	107	266	1.104(0.648~1.882)	
HBV Infection					
No	AA	377	73	1.000	0.417
	AG/GG	724	122	0.872(0.626~1.214)	
Yes	AA	24	368	1.000	0.010
	AG/GG	74	633	0.532(0.328~0.862)	
HCC family history					
No	AA	399	413	1.000	0.039
	AG/GG	790	717	0.756(0.579~0.986)	
Yes	AA	2	28	1.000	0.544
	AG/GG	8	38	0.958(0.836~1.099)	

^a: Adjusted by logistic regression for age, gender, nations, smoking, alcohol consumption, HBV infection, and HCC family history.

OR=0.714; 95% CI=0.539-0.946) genotypes exhibited an activity to reduce the risk of HCC ($P<0.05$).

Stratification analysis

The distribution of HCC risk factors including smoking, alcohol consumption, HBV infection, and HCC family history in the cases and the controls had statistic difference. Therefore, we conducted a stratification analysis based on these factors (Table 3 and Supplementary Tables S1-S5). As shown in Table 3, the interaction of rs3754093 and HCC risk factors was assessed for the possible combined effect on HCC risk.

Associations between *hEXO1* SNPs and the clinicopathological features of HCC

We evaluated the associations of *hEXO1* SNPs with various clinicopathological features of HCC including: tumor size, tumor number, TNM staging (tumor-node-metastasis staging, a cancer staging notation system that

describes the stage of a cancer which originates from a solid tumor with alphanumeric codes), combined with liver cirrhosis, AFP level, and tumor metastasis (Table 4-9). The G allele (AG/GG genotype) of rs3754093 was associated with non-tumor metastasis (adjusted OR: 1.535, 95% CI: 1.023-2.538, $P<0.05$). However, other 5 SNPs of *hEXO1* were not significantly associated with any of the clinicopathological features. Because tumor metastasis is directly related to the prognosis of HCC patients, we performed a univariate survival analysis shown in Figure 1. In accordance with above results, HCC patients with allele G (AG/GG genotype) had a significantly increased risk for death than patients without allele G (AA genotype) (HR= 1.493, 95% CI = 1.102-2.024, $P=0.01$) (Figure 1).

Gene-environment interaction and SNP-SNP interaction

As shown in Table 10, all the SNPs were found to interact with smoking, alcohol consumption, HBV infection in the pathogenesis of HCC. The results of

Table 4: The associations between rs3754093 and clinical characteristics of hepatocellular carcinoma patients

Variables	AA	AG/GG	OR(95%CI) ^a	OR(95%CI) ^b
tumor size				
≥5cm	85(34.56)	161(63.44)	1.000	1.000
<5cm	217(29.17)	527(70.83)	1.282(0.944~1.742)	1.165 (0.876~1.963)
tumor number				
solitary	267(30.58)	606(69.42)	1.000	1.000
multiple	35(29.91)	82(70.09)	1.032(0.677~1.573)	0.972 (0.596~2.248)
TNM staging				
T1+T2	262(31.30)	575(68.70)	1.000	1.000
T3+T4	40(26.14)	113(74.86)	1.287(0.872~1.899)	1.196 (0.756~1.972)
Combined with liver cirrhosis				
Yes	100(23.98)	317(76.02)	1.000	1.000
No	202(35.25)	371(64.75)	0.579(0.437~0.769)	0.585 (0.446~0.756)
AFP level (ng/ml)				
≥400	205 (29.45)	491(70.55)	1.000	1.000
<400	97 (37.16)	164(62.84)	0.706 (0.523~0.952)	0.681 (0.434~1.051)
tumor metastasis				
Yes	53(40.15)	79(59.85)	1.000	1.000
No	249(29.02)	609(70.98)	1.641(1.124~2.394)	1.535 (1.023~2.538)

^a: OR (95% CI) not adjusted; ^b: OR (95% CI) adjusted for age, gender, nations, smoking, alcohol consumption, HBV infection, and HCC family history.

SNP-SNP interaction suggested the interaction between rs3754093 and other 5 SNPs (rs1047840, rs1776148, rs4149867, rs4149963 and rs1776181) was associated with reducing the HCC risk (Table 11).

DISCUSSION

In this study, we investigated six SNPs of the *hEXO1* gene and their association with hereditary susceptibility for HCC in the population of Guangxi, China. Among these six SNPs, we found rs3754093 was significantly associated with a higher susceptibility, and exhibited a protective activity in HCC patients.

hEXO1 is an endonuclease which plays a critical role in both 5'-3' and 3'-5' mispair-dependent excision repair to maintain the overall integrity of the MMR protein complex in human MMR system [33, 34]. Alteration of *MMR* gene can cause cell DNA mismatch repair by increasing cell spontaneous mutation frequency, down-regulating tumor suppressor genes and up-regulating oncogenes, eventually leading to the occurrence of tumors [35]. Therefore, *hEXO1* gene is an important target gene in association with the risk of various malignancies [23, 25, 36]. Moreover, *hEXO1* activates mutations of target genes

through cutting DNA of telomere [43], which is a kind of special protein-DNA structure that exists in eukaryotic cell linear chromosomes [37] and plays an important role in maintaining cell proliferation ability and protecting chromosome integrity [38]. Through its role in these recombinational events, such as repairing of DNA double-strand breaks and maintaining of telomere stabilization, *hEXO1* functions in carcinogenesis of various tumors [18, 26].

In consistent with its key role in carcinogenesis, *hEXO1* rs3754093 showed a protective activity and decreased the risk for death in HCC patients in our study. Furthermore, function prediction showed *hEXO1* rs3754093 was a transcription factor binding site (TFBS). Previous reports showed that SNPs on TFBS could alter the affinity between transcription factors and specific DNA sequences, then cause expression alterations of specific genes [39–41]. That is to say, the polymorphism of rs3754093 could alter the expression level of *hEXO1* through changing its binding affinity to transcription factors. And then the expression change of *hEXO1* might either alter the procedure of MMR to result in gene mutations [42], or increase the risk of cancer through telomere cutting to cause genomic instability [13].

Table 5: The associations between rs1047840 and clinical characteristics of hepatocellular carcinoma patients

Variables	AA	AG/GG	OR(95%CI) ^a	OR(95%CI) ^b
tumor size				
≥5cm	18(7.32)	228(92.68)	1.000	1.000
<5cm	33(4.64)	711(95.36)	0.588(0.325~1.064)	0.582(0.202~1.678)
tumor number				
solitary	48(5.50)	825(94.50)	1.000	1.000
multiple	3(2.56)	114(97.44)	2.211(0.678~7.215)	2.131(0.268~16.913)
TNM staging				
T1+T2	48(5.73)	789(94.27)	1.000	1.000
T3+T4	3(1.96)	150(98.04)	3.042(0.935~9.893)	2.828(0.359~22.287)
Combined with liver cirrhosis				
Yes	21(5.04)	396(94.36)	1.000	1.000
No	30(5.24)	543(94.86)	0.960(0.541~1.702)	1.115(0.402~3.089)
AFP level (ng/ml)				
≥400	33(4.74)	663(95.26)	1.000	1.000
<400	18(6.89)	243(93.10)	0.672(0.371~1.216)	0.749(0.264~2.130)
tumor metastasis				
Yes	6(4.55)	126(95.45)	1.000	1.000
No	45(5.24)	813(94.76)	1.162(0.486~2.781)	1.201(0.257~5.681)

^a: OR (95% CI) not adjusted; ^b: OR (95% CI) adjusted for age, gender, nations, smoking, alcohol consumption, HBV infection, and HCC family history.

This might be the mechanism of association between *hEXO1* rs3754093 and HCC. In addition, we did not find the other 5 SNPs associated with HCC pathogenesis, indicating that the 5 SNPs didn't alter expression level of *hEXO1*.

From gene-environment interaction analysis, we found the six *hEXO1* SNPs were associated with such environment factors, smoking, alcohol consumption and HBV infection. These environmental factors are well documented risk factors of HCC and each of them is an independent strong cause of HCC as previous studies showed. Meta-analysis of epidemiologic studies suggested tobacco use was associated with the increasing of HCC risk [43]. Koh also reported that the tobacco use increased the risk of HCC in the Singapore Chinese population [44]. Both alcohol and tobacco are synergistic risk factors for HCC [45]. In addition, the HBV and HCV infection also plays an important role in increasing the risk of HCC [46], and also is a major factor of HCC risk in Guangxi, China [47]. The chronic HBV infection resulted in the hepatic inflammation reaction, fibrosis, hepatocirrhosis and finally induced liver cancer [46]. We draw a similar conclusion on the effects of these environment factors in development of HCC with previous reports. Moreover, all the six *hEXO1*

SNPs in the study have interaction effects with some environment factors on HCC risk.

Previous studies have reported the SNP-SNP interaction was significantly associated with the pathological process of breast cancer [48], colon cancer [49], oesophageal squamous cell carcinoma [50] and HCC [51], respectively. We found the interaction between rs3754093 and other SNPs (rs1047840, rs1776148, rs4149867, rs4149963 and rs1776181) was significantly associated with reduced the risk of HCC. These results suggested the SNP-SNP interaction may decrease the HCC risk.

HCC is a serious cancer with a complicated pathogenesis related to lots of genes and environmental factors. However, the mechanism of HCC incidence has not been totally understood. In this study, we found the *hEXO1* rs3754093 was a protective factor to reduce the risk of HCC. Moreover, the combined effects of SNP-SNP interactions may decrease the risk of HCC in Guangxi, which has a high incidence of HCC in China. As the association between rs3754093 and the risk of HCC is only an epidemiological link right now, further function study on the SNP should be conducted to validate the current finding.

Table 6: The associations between rs1776148 and clinical characteristics of hepatocellular carcinoma patients

Variables	AA	AG/GG	OR(95%CI) ^a	OR(95%CI) ^b
tumor size				
≥5cm	6(2.44)	240(97.56)	1.000	1.000
<5cm	24(3.23)	720(96.77)	1.333(0.539~3.301)	1.587(0.317~7.937)
tumor number				
solitary	27(3.09)	846(96.91)	1.000	1.000
multiple	3(2.56)	114(97.44)	1.213(0.362~4.062)	1.028(0.121~8.760)
TNM staging				
T1+T2	30(3.58)	807(96.42)	1.000	1.000
T3+T4	6(3.92)	147(96.08)	0.911(0.373~2.227)	1.128(0.569~2.233)
Combined with liver cirrhosis				
Yes	3(0.72)	414(99.28)	1.000	1.000
No	27(4.71)	546(95.29)	0.147(0.044~0.486)	0.154(0.019~1.260)
AFP level (ng/ml)				
≥400	27(3.88)	669(96.12)	1.000	1.000
<400	3(1.02)	291(98.98)	3.915(1.178~13.007)	4.391(0.536~35.971)
tumor metastasis				
Yes	6(4.55)	126(95.45)	1.000	1.000
No	30(3.50)	828(96.50)	0.761(0.310~1.865)	0.764(0.164~3.606)

^a: OR (95% CI) not adjusted; ^b: OR (95% CI) adjusted for age, gender, nations, smoking, alcohol consumption, HBV infection, and HCC family history.

Table 7: The associations between rs4149867 and clinical characteristics of hepatocellular carcinoma patients

Variables	AA	AG/GG	OR(95%CI) ^a	OR(95%CI) ^b
tumor size				
≥5cm	165(67.07)	81(32.93)	1.000	1.000
<5cm	501(67.34)	243(32.66)	1.125(0.533~2.373)	0.638(0.357~1.138)
tumor number				
solitary	609(69.76)	264(30.24)	1.000	1.000
multiple	78(66.66)	39(33.34)	1.153(0.765~1.739)	1.080(0.521~2.238)
TNM staging				
T1+T2	588(70.25)	249(29.75)	1.000	1.000
T3+T4	96(64.00)	54(36.00)	1.328(0.922~1.913)	1.250(0.646~2.419)
Combined with liver cirrhosis				
Yes	273(65.94)	141(34.06)	1.000	1.000
No	414(72.25)	159(27.05)	0.744(0.566~0.977)	0.781(0.480~1.270)
AFP level (ng/ml)				
≥400	483(69.40)	213(30.60)	1.000	1.000
<400	204(69.39)	90(30.61)	1.000(0.744~1.345)	0.988(0.587~1.665)

(Continued)

Variables	AA	AG/GG	OR(95%CI) ^a	OR(95%CI) ^b
tumor metastasis				
Yes	99(75.00)	33(25.00)	1.000	1.000
No	582(68.31)	270(31.69)	0.719(0.472~1.093)	0.726(0.348~1.515)

^a: OR (95% CI) not adjusted; ^b: OR (95% CI) adjusted for age, gender, nations, smoking, alcohol consumption, HBV infection, and HCC family history.

Table 8: The associations between rs4149963 and clinical characteristics of hepatocellular carcinoma patients

Variables	AA	AG/GG	OR(95%CI) ^a	OR(95%CI) ^b
tumor size				
≥5cm	218(88.62)	28(11.38)	1.000	1.000
<5cm	669(89.91)	75(10.08)	1.146(0.723~1.815)	1.648(0.775~3.508)
tumor number				
solitary	780(89.34)	93(10.65)	1.000	1.000
multiple	104(88.89)	13(11.11)	1.048(0.567~1.940)	1.610(0.614~4.220)
TNM staging				
T1+T2	738(88.17)	99(11.83)	1.000	1.000
T3+T4	131(87.33)	19(2.67)	1.081(0.640~1.828)	0.586(0.189~1.814)
Combined with liver cirrhosis				
Yes	360(86.96)	51(13.04)	1.000	1.000
No	516(90.05)	57(9.95)	0.780(0.522~1.164)	0.759(0.378~1.527)
AFP level (ng/ml)				
≥400	603(86.64)	93(13.36)	1.000	1.000
<400	252(85.71)	42(14.29)	1.081(0.730~1.601)	0.420(0.168~1.049)
tumor metastasis				
Yes	117(88.64)	15(11.36)	1.000	1.000
No	762(88.81)	96(11.19)	1.018(0.571~1.814)	1.308(0.506~3.382)

^a: OR (95% CI) not adjusted; ^b: OR (95% CI) adjusted for age, gender, nations, smoking, alcohol consumption, HBV infection, and HCC family history.

Table 9: The associations between rs1776181 and clinical characteristics of hepatocellular carcinoma patients

Variables	AA	AG/GG	OR(95%CI) ^a	OR(95%CI) ^b
tumor size				
≥5cm	390(52.42)	354(41.58)	1.000	1.000
<5cm	120(48.78)	126(51.22)	1.157(0.867~1.544)	1.183 (0.711~1.966)
tumor number				
solitary	450(51.55)	423(48.45)	1.000	1.000
multiple	60(51.28)	57(48.72)	1.101(0.687~1.487)	0.919 (0.464~1.819)
TNM staging				
T1+T2	427(49.82)	410(50.18)	1.000	1.000
T3+T4	80(60.00)	70(40.0)	0.911(0.643~1.291)	0.850 (0.545~1.426)

(Continued)

Variables	AA	AG/GG	OR(95%CI) ^a	OR(95%CI) ^b
Combined with liver cirrhosis				
Yes	231(55.80)	183(44.20)	1.000	1.000
No	279(48.69)	294(51.31)	1.330(1.032~1.715)	1.295 (0.828~2.028)
AFP level (ng/ml)				
≥400	357(51.29)	339(48.71)	1.000	1.000
<400	153(52.04)	141(47.96)	0.971(0.739~1.275)	0.882 (0.597~1.558)
tumor metastasis				
Yes	72(54.55)	60(45.45)	1.000	1.000
No	435(51.06)	423(48.94)	0.857(0.593~1.239)	0.857 (0.450~1.632)

^a: OR (95% CI) not adjusted; ^b: OR (95% CI) adjusted for age, gender, nations, smoking, alcohol consumption, HBV infection, and HCC family history

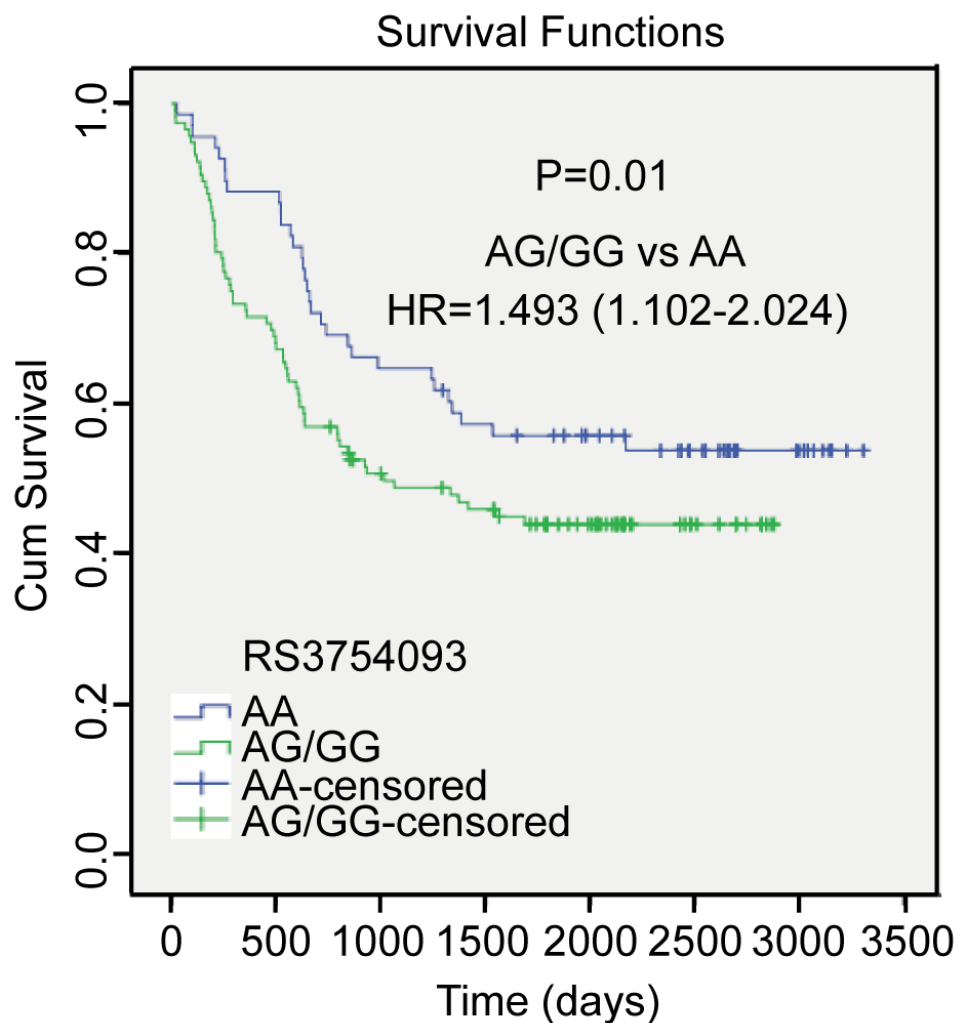


Figure 1: Kaplan-Meier overall survival curve for HCC patients based on rs3754093 genotypes. *P* value is from the log-rank test. HR with 95% CI was from univariate analysis of OS and DFS.

Table 10: Gene-environment interaction

Factors	β	S.E.	Wald χ^2	OR (95%CI) ^a	P
rs1047840 × Smoking	0.428	0.093	21.020	1.534(1.278~1.842)	0.000
rs1047840 × Alcohol Consumption	0.336	0.096	12.225	1.399(1.159~1.689)	0.000
rs1047840 × HBV Infection	2.102	0.074	809.417	8.818(7.078~9.456)	0.000
rs1047840 × HCC family history	0.229	0.235	0.949	1.257(0.793~1.293)	0.330
rs1776148 × Smoking	0.424	0.093	20.652	1.527(1.272~1.833)	0.000
rs1776148 × Alcohol Consumption	0.318	0.095	11.066	1.374(1.139~1.657)	0.001
rs1776148 × HBV Infection	2.080	0.073	820.029	8.003(6.941~9.288)	0.000
rs1776148 × HCC family history	0.236	0.223	1.022	1.266(0.801~2.001)	0.312
rs3754093 × Smoking	0.421	0.103	16.756	1.524(1.245~1.864)	0.000
rs3754093 × Alcohol Consumption	0.367	0.105	12.101	1.443(1.174~1.775)	0.001
rs3754093 × HBV Infection	2.326	0.092	641.686	10.235(8.549~12.253)	0.000
rs3754093 × HCC family history	0.186	0.269	0.477	1.204(0.711~2.041)	0.490
rs4149867 × Smoking	0.462	0.124	13.852	1.587(1.244~2.023)	0.000
rs4149867 × Alcohol Consumption	0.385	0.134	8.240	1.469(1.130~1.910)	0.004
rs4149867 × HBV Infection	3.238	0.126	664.917	25.485(19.925~32.597)	0.000
rs4149867 × HCC family history	0.391	0.340	1.324	1.478(0.760~2.876)	0.250
rs4149963 × Smoking	0.610	0.149	16.769	1.840(1.374~2.464)	0.000
rs4149963 × Alcohol Consumption	0.454	0.159	8.276	1.580(1.157~2.158)	0.004
rs4149963 × HBV Infection	3.707	0.134	762.612	40.703(31.296~52.965)	0.000
rs4149963 × HCC family history	0.350	0.401	0.760	1.419(0.646~3.113)	0.383
rs1776181 × Smoking	0.497	0.110	20.283	1.644(1.324~2.042)	0.000
rs1776181 × Alcohol Consumption	0.347	0.115	9.139	1.414(1.130~1.771)	0.003
rs1776181 × HBV Infection	2.578	0.103	620.820	13.175(10.756~16.137)	0.000
rs1776181 × HCC family history	0.372	0.316	1.384	1.450(0.781~2.695)	0.239

^a: Adjusted by logistic regression for age, gender, nations, smoking, alcohol consumption, HBV infection, and HCC family history.

Table 11: SNP-SNP interaction on HCC risk

Factors	β	S.E.	Wald χ^2	OR (95%CI) ^a	P
rs1047840 × rs1776148	-0.202	0.113	3.190	0.817(0.654~1.020)	0.074
rs1047840 × rs3754093	-0.148	0.065	5.262	0.862(0.759~0.979)	0.022
rs1047840 × rs4149867	-0.055	0.069	0.627	0.947(0.826~1.084)	0.429
rs1047840 × rs4149963	-0.098	0.096	1.045	0.906(0.751~1.094)	0.307
rs1047840 × rs1776181	-0.061	0.063	0.924	0.941(0.831~1.065)	0.337
rs1776148 × rs3754093	-0.177	0.065	7.302	0.838(0.737~0.953)	0.007
rs1776148 × rs4149867	-0.052	0.070	0.547	0.950(0.828~1.089)	0.460
rs1776148 × rs4149963	-0.151	0.098	2.367	0.860(0.709~1.042)	0.124
rs1776148 × rs1776181	-0.067	0.063	1.104	0.937(0.827~1.059)	0.293

(Continued)

Factors	β	S.E.	Wald χ^2	OR (95%CI) ^a	P
rs3754093 × rs4149867	-0.134	0.065	4.193	0.875(0.769~0.994)	0.041
rs3754093 × rs4149963	-0.171	0.084	4.153	0.843(0.715~0.993)	0.042
rs3754093 × rs1776181	-0.128	0.059	4.660	0.880(0.783~0.988)	0.031
rs4149867 × rs4149963	-0.104	0.098	1.129	0.901(0.744~1.092)	0.288
rs4149867 × rs1776181	-0.090	0.060	2.301	0.914(0.813~1.027)	0.129
rs4149963 × rs1776181	-0.066	0.083	2.628	0.936(0.795~1.102)	0.428

^a: Adjusted by logistic regression for age, gender, nations, smoking, alcohol consumption, HBV infection, and HCC family history.

MATERIALS AND METHODS

Study subjects

The cases (n=1,199) were newly diagnosed as HCC patients without chemotherapy and radiotherapy who recruited from the Affiliated Tumor Hospital of Guangxi Medical University between June 2007 and March 2014. All HCC patients were diagnosed according to the World Health Organization (WHO) Classification of HCC diagnostic criteria. The controls (n=1,196) were non-tumor patients from the First Affiliated Hospital of Guangxi Medical University during the same period. Inclusion and exclusion criteria for participants: 1. Inclusion criteria: (1) Cases: new HCC patients hospitalized in the period of study, and were pathologically confirmed. (2) Controls: patients without HCC hospitalized in the same period with the cases. 2. Exclusion criteria: (1) Cases: have been treated, HCV infected, with other tumors. (2) Controls: suffering from cancer, HCV infected. This study was respectively approved by the Institutional Review Board of the Affiliated Tumor Hospital of Guangxi Medical University and First Affiliated Hospital of Guangxi Medical University. The informed consent was obtained from each subject in this study.

Questionnaire survey and blood sample collection

Questionnaires were designed after consulting experts. Data of questionnaire survey was collected from a structured interview, and conducted by trained interviewers after a pre-investigation, which including the demographic characteristics (name, nation, gender, age), smoking alcohol consumption, hepatitis B virus (HBV) infection and HCC family history. 2 mL peripheral blood sample was collected from each study object. Genome DNA was extracted from the blood samples. All laboratory and questionnaire data were coded, entered by two investigators with a logical and consistency test, and verified using EpiData 3.1 (www.epidata.dk/download.php). Neither the laboratory nor the data entry personnel had any knowledge of the subjects' case-control status.

DNA extraction and genotyping

The blood was collected from the all subjects using an EDTA-K₂ anticoagulant blood vessel. The DNA was extracted according to the phenol-chloroform method and stored at -80°C. The genotyping was performed in an Applied Biosystems 7500 Fast Real-Time quantitative polymerase chain reaction (qPCR) system with the following TaqMan SNP Genotyping Assays kit (Applied Biosystems) according to the manufacturer's instructions. Each PCR reaction mixture (10 μ L) contained DNA template (0.4 μ L), ddH₂O (4.35 μ L), 2 \times TaqMan Universal PCR Master Mix (5 μ L), and 20 \times SNP Genotyping Assay Mix (0.25 μ L). The genomic DNA was amplified at 95°C for 10 min, followed by 40 cycles of 95°C for 15s and 60°C for 60s. All genotyping reagents and analytical software were purchased from Applied Biosystems. Approximately 5% of the samples were randomly repeated to validate the genotyping procedures, and the concordance rate was 100%. The results of the genotyping were analyzed with 7500 Fast System V1.4.0 SDS software.

Survival analysis

356 HCC patients were brought into a survival analysis cohort. The patients were followed up from July 2007 to May 2016. Survival time was counted the first day after surgery and end up when the patients appeared metastasis and recurrence or death. Survival time was calculated in months. Till the end of follow-up, a total of 31 patients were lost to follow-up, 325 patients with complete follow-up data, follow-up rate was 91.3%.

Statistical analysis

Data entry and consistency check were conducted on the EpiData3.1 software. The SPSS 20.0 software was used for the statistical analyses. Logistic regression models were used to estimate odds ratio (OR) and 95% confidence interval (CI). Gene-environment interactions and gene-gene interactions were performed through

multiplicative model of binary logistic regression. Hardy-Weinberg Equilibrium (HWE) in the controls was tested by Haploview 4.2 software. Kaplan–Meier method, Log–rank test and COX regression were used on survival analysis.

ACKNOWLEDGMENTS

This study was supported by the grant from the National Natural Science Foundation of China (81160359).

CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

REFERENCES

1. Bosetti C, Turati F and La Vecchia C. Hepatocellular carcinoma epidemiology. *Best practice & research*. 2014; 28:753-770.
2. Ding J and Wang H. Multiple interactive factors in hepatocarcinogenesis. *Cancer letters*. 2014; 346:17-23.
3. Zhang CY, Huang TR, Yu JH, Zhang ZQ, Li JL, Deng W, Ye SY, Zhou DN and He ZF. Epidemiological analysis of primary liver cancer in the early 21st century in Guangxi province of China. *Chinese journal of cancer*. 2010; 29:545-550.
4. Flemming JA, Yang JD, Vittinghoff E, Kim WR and Terrault NA. Risk prediction of hepatocellular carcinoma in patients with cirrhosis: the ADRESS-HCC risk model. *Cancer*. 2014; 120:3485-3493.
5. Jun CH, Hong HJ, Chung MW, Park SY, Cho SB, Park CH, Joo YE, Kim HS, Choi SK and Rew JS. Risk factors for hepatocellular carcinoma in patients with drug-resistant chronic hepatitis B. *World journal of gastroenterology*. 2013; 19:6834-6841.
6. Paranagua-Vezozzo DC, Ono SK, Alvarado-Mora MV, Farias AQ, Cunha-Silva M, Franca JI, Alves VA, Sherman M and Carrilho FJ. Epidemiology of HCC in Brazil: incidence and risk factors in a ten-year cohort. *Annals of hepatology*. 2014; 13:386-393.
7. Toyoda H, Kumada T, Tada T, Kiriya S, Tanikawa M, Hisanaga Y, Kanamori A, Kitabatake S and Ito T. Risk factors of hepatocellular carcinoma development in non-cirrhotic patients with sustained virologic response for chronic hepatitis C virus infection. *Journal of gastroenterology and hepatology*. 2015; 30:1183-1189.
8. Muro Y, Sugiura K, Mimori T and Akiyama M. DNA mismatch repair enzymes: genetic defects and autoimmunity. *Clinica chimica acta; international journal of clinical chemistry*. 2015; 442:102-109.
9. Schmutte C, Marinescu RC, Sadoff MM, Guerrette S, Overhauser J and Fishel R. Human exonuclease I interacts with the mismatch repair protein hMSH2. *Cancer research*. 1998; 58:4537-4542.
10. Tishkoff DX, Amin NS, Viars CS, Arden KC and Kolodner RD. Identification of a human gene encoding a homologue of *Saccharomyces cerevisiae* EXO1, an exonuclease implicated in mismatch repair and recombination. *Cancer research*. 1998; 58:5027-5031.
11. Wilson DM, 3rd, Carney JP, Coleman MA, Adamson AW, Christensen M and Lamerdin JE. Hex1: a new human Rad2 nuclease family member with homology to yeast exonuclease I. *Nucleic acids research*. 1998; 26:3762-3768.
12. Orans J, McSweeney EA, Iyer RR, Hast MA, Hellinga HW, Modrich P and Beese LS. Structures of human exonuclease I DNA complexes suggest a unified mechanism for nuclease family. *Cell*. 2011; 145:212-223.
13. Jager AC, Rasmussen M, Bisgaard HC, Singh KK, Nielsen FC and Rasmussen LJ. HNPCC mutations in the human DNA mismatch repair gene hMLH1 influence assembly of hMutLalpha and hMLH1-hEXO1 complexes. *Oncogene*. 2001; 20:3590-3595.
14. Rasmussen LJ, Rasmussen M, Lee B, Rasmussen AK, Wilson DM, 3rd, Nielsen FC and Bisgaard HC. Identification of factors interacting with hMSH2 in the fetal liver utilizing the yeast two-hybrid system. In vivo interaction through the C-terminal domains of hEXO1 and hMSH2 and comparative expression analysis. *Mutation research*. 2000; 460:41-52.
15. Schmutte C, Sadoff MM, Shim KS, Acharya S and Fishel R. The interaction of DNA mismatch repair proteins with human exonuclease I. *The Journal of biological chemistry*. 2001; 276:33011-33018.
16. Tishkoff DX, Boeger AL, Bertrand P, Filosi N, Gaida GM, Kane MF and Kolodner RD. Identification and characterization of *Saccharomyces cerevisiae* EXO1, a gene encoding an exonuclease that interacts with MSH2. *Proceedings of the National Academy of Sciences of the United States of America*. 1997; 94:7487-7492.
17. Tran PT, Simon JA and Liskay RM. Interactions of Exo1p with components of MutLalpha in *Saccharomyces cerevisiae*. *Proceedings of the National Academy of Sciences of the United States of America*. 2001; 98:9760-9765.
18. Wei K, Clark AB, Wong E, Kane MF, Mazur DJ, Parris T, Kolas NK, Russell R, Hou H, Jr., Kneitz B, Yang G, Kunkel TA, Kolodner RD, Cohen PE and Edelman W. Inactivation of Exonuclease I in mice results in DNA mismatch repair defects, increased cancer susceptibility, and male and female sterility. *Genes & development*. 2003; 17:603-614.
19. Chang JS, Yeh RF, Wiencke JK, Wiemels JL, Smirnov I, Pico AR, Tihan T, Patoka J, Miike R, Sison JD, Rice T and Wensch MR. Pathway analysis of single-nucleotide polymorphisms potentially associated with glioblastoma multiforme susceptibility using random forests. *Cancer Epidemiol Biomarkers Prev*. 2008; 17:1368-1373.

20. Hansen LT, Thykjaer T, Orntoft TF, Rasmussen LJ, Keller P, Spang-Thomsen M, Edmonston TB, Schmutte C, Fishel R and Petersen LN. The role of mismatch repair in small-cell lung cancer cells. *Eur J Cancer*. 2003; 39:1456-1467.
21. Nielsen FC, Jager AC, Lutzen A, Bundgaard JR and Rasmussen LJ. Characterization of human exonuclease 1 in complex with mismatch repair proteins, subcellular localization and association with PCNA. *Oncogene*. 2004; 23:1457-1468.
22. Sun X, Zheng L and Shen B. Functional alterations of human exonuclease 1 mutants identified in atypical hereditary nonpolyposis colorectal cancer syndrome. *Cancer research*. 2002; 62:6026-6030.
23. Thompson E, Meldrum CJ, Crooks R, McPhillips M, Thomas L, Spigelman AD and Scott RJ. Hereditary non-polyposis colorectal cancer and the role of hPMS2 and hEXO1 mutations. *Clinical genetics*. 2004; 65:215-225.
24. Wang HC, Chiu CF, Tsai RY, Kuo YS, Chen HS, Wang RF, Tsai CW, Chang CH, Lin CC and Bau DT. Association of genetic polymorphisms of EXO1 gene with risk of breast cancer in Taiwan. *Anticancer research*. 2009; 29:3897-3901.
25. Wu Y, Berends MJ, Post JG, Mensink RG, Verlind E, Van Der Sluis T, Kempinga C, Sijmons RH, van der Zee AG, Hollema H, Kleibeuker JH, Buys CH and Hofstra RM. Germline mutations of EXO1 gene in patients with hereditary nonpolyposis colorectal cancer (HNPCC) and atypical HNPCC forms. *Gastroenterology*. 2001; 120:1580-1587.
26. Bau DT, Wang HC, Liu CS, Chang CL, Chiang SY, Wang RF, Tsai CW, Lo YL, Hsiung CA, Lin CC and Huang CY. Single-nucleotide polymorphism of the Exo1 gene: association with gastric cancer susceptibility and interaction with smoking in Taiwan. *The Chinese journal of physiology*. 2009; 52:411-418.
27. Hsu NY, Wang HC, Wang CH, Chiu CF, Tseng HC, Liang SY, Tsai CW, Lin CC and Bau DT. Lung cancer susceptibility and genetic polymorphisms of Exo1 gene in Taiwan. *Anticancer research*. 2009; 29:725-730.
28. Tsai MH, Tseng HC, Liu CS, Chang CL, Tsai CW, Tsou YA, Wang RF, Lin CC, Wang HC, Chiu CF and Bau DT. Interaction of Exo1 genotypes and smoking habit in oral cancer in Taiwan. *Oral oncology*. 2009; 45:e90-94.
29. Jin G, Wang H, Hu Z, Liu H, Sun W, Ma H, Chen D, Miao R, Tian T, Jin L, Wei Q, Huang W, Lu D and Shen H. Potentially functional polymorphisms of EXO1 and risk of lung cancer in a Chinese population: A case-control analysis. *Lung cancer (Amsterdam, Netherlands)*. 2008; 60:340-346.
30. Luo X, Hong XS, Xiong XD, Zeng LQ and Lim CE. A single nucleotide polymorphism in EXO1 gene is associated with cervical cancer susceptibility in Chinese patients. *Int J Gynecol Cancer*. 2012; 22:220-225.
31. Bayram S, Akkiz H, Bekar A, Akgollu E and Yildirim S. The significance of Exonuclease 1 K589E polymorphism on hepatocellular carcinoma susceptibility in the Turkish population: a case-control study. *Molecular biology reports*. 2012; 39:5943-5951.
32. Kabzinski J, Mucha B, Cuchra M, Markiewicz L, Przybylowska K, Dziki A, Dziki L and Majsterek I. Efficiency of Base Excision Repair of Oxidative DNA Damage and Its Impact on the Risk of Colorectal Cancer in the Polish Population. *Oxidative medicine and cellular longevity*. 2016; 2016:3125989.
33. Knudsen NO, Nielsen FC, Vinther L, Bertelsen R, Holten-Andersen S, Liberti SE, Hofstra R, Kooi K and Rasmussen LJ. Nuclear localization of human DNA mismatch repair protein exonuclease 1 (hEXO1). *Nucleic acids research*. 2007; 35:2609-2619.
34. Liberti SE and Rasmussen LJ. Is hEXO1 a cancer predisposing gene? *Mol Cancer Res*. 2004; 2:427-432.
35. Bak ST, Sakellariou D and Pena-Diaz J. The dual nature of mismatch repair as antimutator and mutator: for better or for worse. *Frontiers in genetics*. 2014; 5:287.
36. Jagmohan-Changur S, Poikonen T, Vilkki S, Launonen V, Wikman F, Orntoft TF, Moller P, Vasen H, Tops C, Kolodner RD, Mecklin JP, Jarvinen H, Bevan S, et al. EXO1 variants occur commonly in normal population: evidence against a role in hereditary nonpolyposis colorectal cancer. *Cancer research*. 2003; 63:154-158.
37. Martinez P and Blasco MA. Replicating through telomeres: a means to an end. *Trends in biochemical sciences*. 2015; 40:504-515.
38. Konvalinova H, Dvorakova Z, Renciuik D, Bednarova K, Kejnovska I, Trantirek L, Vorlickova M and Sagi J. Diverse effects of naturally occurring base lesions on the structure and stability of the human telomere DNA quadruplex. *Biochimie*. 2015; 118:15-25.
39. Buroker NE. Regulatory SNPs and transcriptional factor binding sites in ADRBK1, AKT3, ATF3, DIO2, TBXA2R and VEGFA. *Transcription*. 2015; 5:e964559.
40. Hesp K, Smant G and Kammenga JE. *Caenorhabditis elegans* DAF-16/FOXO transcription factor and its mammalian homologs associate with age-related disease. *Experimental gerontology*. 2015; 72:1-7.
41. Mundade R, Ozer HG, Wei H, Prabhu L and Lu T. Role of ChIP-seq in the discovery of transcription factor binding sites, differential gene regulation mechanism, epigenetic marks and beyond. *Cell cycle (Georgetown, Tex)*. 2014; 13:2847-2852.
42. Nogueira GA, Lourenco GJ, Oliveira CB, Marson FA, Lopes-Aguiar L, Costa EF, Lima TR, Liutti VT, Leal F, Santos VC, Rinck-Junior JA and Lima CS. Association between genetic polymorphisms in DNA mismatch repair-related genes with risk and prognosis of head and neck squamous cell carcinoma. *International journal of cancer*. 2015; 137:810-818.
43. Lee YC, Cohet C, Yang YC, Stayner L, Hashibe M and Straif K. Meta-analysis of epidemiologic studies on cigarette smoking and liver cancer. *International journal of epidemiology*. 2009; 38:1497-1511.

44. Koh WP, Robien K, Wang R, Govindarajan S, Yuan JM and Yu MC. Smoking as an independent risk factor for hepatocellular carcinoma: the Singapore Chinese Health Study. *British journal of cancer*. 2011; 105:1430-1435.
45. Marrero JA, Fontana RJ, Fu S, Conjeevaram HS, Su GL and Lok AS. Alcohol, tobacco and obesity are synergistic risk factors for hepatocellular carcinoma. *Journal of hepatology*. 2005; 42:218-224.
46. Chuang SC, La Vecchia C and Boffetta P. Liver cancer: descriptive epidemiology and risk factors other than HBV and HCV infection. *Cancer letters*. 2009; 286:9-14.
47. Yeh FS, Yu MC, Mo CC, Luo S, Tong MJ and Henderson BE. Hepatitis B virus, aflatoxins, and hepatocellular carcinoma in southern Guangxi, China. *Cancer research*. 1989; 49:2506-2509.
48. Sapkota Y, Mackey JR, Lai R, Franco-Villalobos C, Lupichuk S, Robson PJ, Kopciuk K, Cass CE, Yasui Y and Damaraju S. Assessing SNP-SNP interactions among DNA repair, modification and metabolism related pathway genes in breast cancer susceptibility. *PloS one*. 2013; 8:e64896.
49. Goodman JE, Mechanic LE, Luke BT, Ambs S, Chanock S and Harris CC. Exploring SNP-SNP interactions and colon cancer risk using polymorphism interaction analysis. *International journal of cancer*. 2006; 118:1790-1797.
50. Wang Y, Vogelsang M, Schafer G, Matejcic M and Parker MI. MicroRNA polymorphisms and environmental smoke exposure as risk factors for oesophageal squamous cell carcinoma. *PloS one*. 2013; 8:e78520.
51. Bei CH, Bai H, Yu HP, Yang Y, Liang QQ, Deng YY, Tan SK and Qiu XQ. Combined effects of six cytokine gene polymorphisms and SNP-SNP interactions on hepatocellular carcinoma risk in Southern Guangxi, China. *Asian Pac J Cancer Prev*. 2014; 15:6961-6967.